

Environment Protection Authority

GPO Box 1550 HOBART TAS 7001 Australia

Enquiries: Megan Burgoyne
Phone: +61 3 6165 4592
Email: Megan.Burgoyne@epa.tas.gov.au
Web: www.epa.tas.gov.au
Our Ref: D25-45392



7 March 2025

Henrique Batista
Chief Executive Officer
Huon Aquaculture Company Pty Ltd
ACN 067 386 109
GPO Box 987
HOBART TAS 7001

Email: hbatista@huonaqua.com.au

Dear Henrique Batista

AMENDED ANTIBIOTIC RESIDUE MONITORING SURVEY – ENVIRONMENTAL LICENCE NO. 9882/4 - MF141 (SOUTH OF ZUIDPOOL ROCK)

As per my correspondence dated 21 February 2025, Huon Aquaculture Company Pty Ltd (Huon) is required to conduct an Antibiotic Residue Survey for Marine Farming Lease No. 141 (South of Zuidpool Rock), in accordance with the *Antibiotic Residue Monitoring Schedule – MF141 – Feb 2025* (the Schedule).

This office was notified on 16 February 2025 of the presence of fatty globules along the beach at Verona Sands. Samples were subsequently collected and were determined to be consistent with fish oil. Since that date, the EPA has received several additional reports of congealed fish oil globules and fish carcasses washing up on a number of beaches in the southern D'Entrecasteaux Channel region.

As a result of these reports and laboratory results received to date, I have determined that additional antibiotic residue monitoring at four ambient beach sites in the vicinity of Marine Farming Lease No. 141 is required. Please find enclosed an amended Schedule (*AMENDED Antibiotic Residue Monitoring Schedule – MF141 – 7 Mar 2025*) which outlines the additional sediment and water monitoring requirements. All amendments to the schedule are marked in red.

Please note that these additional samples are scheduled to be collected on 12 March 2025, in conjunction with all other Day 14 monitoring required by the enclosed Schedule.

Huon Aquaculture Company Pty Ltd must notify this office immediately if there is any change to the information provided for this treatment event.

If you have any queries or feedback, please contact Megan Burgoyne on (03) 6165 4592 or via email Megan.Burgoyne@epa.tas.gov.au.

Yours sincerely

Darryl Cook
DIRECTOR FINFISH COMPLIANCE
Delegate for the Director, Environment Protection Authority

Enc: *AMENDED Antibiotic Residue Monitoring Schedule - MF141 – 7 Mar 2025*

cc: mwhittle@huonaqua.com.au asmak@huonaqua.com.au compliance@huonaqua.com.au

AMENDED ANTIBIOTIC RESIDUE MONITORING SCHEDULE - MF141
7 MARCH 2025

This Schedule supersedes the previous version. Changes are marked in red.

Preamble

Risk matrix to determine intensity and extent of monitoring program

The appropriate level of monitoring, including key elements to be included in surveys, is determined based on the following risk matrix.

	Monitoring element	Sediment	Wild fish tissue	Oysters	Other
	Element description	- within lease - outside of lease - at reference sites	- within lease - outside of lease	Oysters grown for human consumption in vicinity of lease	As determined by the Director depending on site-specific circumstances
Intensity and duration of medication event					
Low	<50 kg active ingredient/ lease (single event)	No	No		
Medium	> 50 kg active ingredient/ lease (per calendar year, combined for multiple events)	Yes (30% of treated cages to be sampled) + external sites	Yes		
High	Sustained, intensive treatment: >500 kg / year and / or > 2 weeks duration	Yes (50% of treated cages to be sampled) + external sites	Yes	The nearest production leases in the Port Esperance and Little Taylors Bay growing areas are approx. 8.5 and 6 km away, respectively.	Benthic Video Surveys of all four treated pen bays to be conducted on day 1 (following treatment) of the sampling schedule.
Special considerations	Vicinity of oysters grown and harvested for human consumption				
Colour coding		Determined to be necessary	Determined not to be necessary	Required to address specific circumstances	Not applicable

I. Outline of Requirements

An Antibiotic Residue Monitoring Program is to be conducted with respect to the finfish farming activity ('the activity') undertaken by Huon Aquaculture Company Pty Ltd under Environmental Licence No. 9882/4, on Marine Farming Lease No. 141 (South of Zuidpool Rock) ('the lease') located in the D'Entrecasteaux Channel and Huon River Marine Farming Development Plan area.

The monitoring program is to be undertaken in accordance with the specifications in this Schedule or as otherwise specified in writing by the Director, EPA. The need for ongoing monitoring will be reviewed on request depending on the results of the monitoring.

Survey reports must be submitted by the licence holder to the Salmon Environmental Regulation Division by email to Salmon.Reg@epa.tas.gov.au in accordance with section 2.6 of this schedule. The final consolidated monitoring report is to be made publicly available, either by the licence holder and/or on the EPA website.

The video surveys and sampling must be conducted at each of the sites identified in sections 2.1, 2.2, 2.3, 2.4 and 2.5 as shown on the maps in Attachment 1. All positional requirements (prescribed sample sites) of the survey are to be located and recorded using a GPS.

All sample collection for a given sampling event is to be conducted on the same day to the extent practical. If all of the samples for a given sampling event cannot be collected on the same day, the sampling event may be conducted over two consecutive days.

If a scheduled sampling event coincides with a weekend or public holiday, samples are to be taken on the day immediately before or after the scheduled date.

Any sample or measurement required below must be taken and processed in accordance with the following:

- sampling and measuring must be undertaken by a person with appropriate training, experience, and knowledge of the relevant procedure;
- the integrity of samples must be preserved prior to delivery to a laboratory;
- sample analysis or measurement must be conducted by a laboratory or testing facility accredited by the National Association of Testing Authorities (NATA), or a laboratory or testing facility approved in writing by the Director, EPA for the specified test; and
- details of methods employed in taking samples and measurements and results of sample analysis must be retained for at least three (3) years after the date of collection.

The Antibiotic Residue Monitoring Program must include the following components:

- 2.1 Benthic video surveys
- 2.2 Sediment sampling
- 2.3 Wild fish sampling
- 2.4 Water sampling
- 2.5 Congealed fish oil sampling
- 2.6 Reporting of results

2. Antibiotic Residue Monitoring Program Specifications

2.1 Benthic Video Surveys

Survey locations:

Benthic video surveys must be conducted at the following locations:

a) Cage sites:

All four pen bays undergoing treatment must be surveyed.

Surveys must be conducted on the first day following completion of treatment (day 1) and in accordance with the benthic video survey conditions in Environmental Licence No. 9882/4.

Table 1: Benthic video survey sites

Pen Bay	Site Category	Easting	Northing	Survey Day
ZN05	In-lease	512208	5201912	Day 1 after completion of medication
ZS03	In-lease	509512	5199513	Day 1 after completion of medication
ZS04	In-lease	509606	5199611	Day 1 after completion of medication
ZS12	In-lease	509103	5199315	Day 1 after completion of medication

2.2 Sediment sampling

Sampling locations:

Samples must be collected from the following locations:

a) Cage sites:

In total three pen bays are to be sampled.

Samples must be taken at the edge of the medicated cage, downstream of prevailing water currents, and the position must be recorded at the time of collection.

b) Outside of lease sites:

Four (4) transects on **Zuidpool North** and five (5) transects on **Zuidpool South**, are to be sampled.

These transects are positioned at right angles to the marine farming lease boundaries. The transect samples are to be taken at points which are at a distance of 35 m, 100 m and 500 m from the lease boundary.

c) Ambient sites:

Sediment samples are to be taken at six (6) ambient sites.

BEMP Site M6 (Central Mid Channel) is the same site as referred to in Environmental Licence No. 9882/4. The SW site has been located in-line with the predominate current flow to the southwest of the lease. Sites C10.2 and C11.2 are baseline survey control sites for the lease. Sample sites located off of Ventenat Point (VENPT), Bruny Island, and Lomas Point (LOMPT), at the southern entrance to Port Esperance, have been established to assess potential impact on nearby commercial shellfish leases and to address some community concern.

d) Ambient beach sites:

Sediment samples are to be taken at twelve (12) ambient beach transect sites.

Three sites have been selected along four of the beaches known to have had congealed fish oil globules wash ashore. The four beaches to be sampled include Verona Sands (VS), Jetty Beach (JB), Conleys Beach (CB) and Roaring Bay Beach (RBB). Approximate coordinates for each site are available in Table 2. The samples for these sites are to be collected on day 14 and 28, after

completion of the medication event (Attachment 2). Monitoring locations, duration and frequency may be increased if OTC is detected in the samples.

All beach sediment samples should be collected along the tide line and directly under congealed fish oil globules, if present. If congealed fish oil globules are present, a sample of the globules must also be collected in accordance with the requirements listed under Section 2.5 of this document.

Table 2: Sediment sampling sites

Site Name	Site Category & Purpose	Easting	Northing	Sampling Day
Pen Bay ZN05	In-lease	512208	5201912	Day 1, 14, 28 & 56 after completion of medication
ZN05-NE35	Transect outside lease	512426	5202137	As above
ZN05-NE100	Transect outside lease	512451	5202189	As above
ZN05-NE500	Transect outside lease	512727	5202480	As above
ZN05-NW35	Transect outside lease	511853	5202232	As above
ZN05-NW100	Transect outside lease	511804	5202272	As above
ZN05-NW500	Transect outside lease	511509	5202545	As above
ZN05-SE35	Transect outside lease	512500	5201648	As above
ZN05-SE100	Transect outside lease	512552	5201603	As above
ZN05-SE500	Transect outside lease	512852	5201341	As above
ZN05-SW35	Transect outside lease	511726	5201382	As above
ZN05-SW100	Transect outside lease	511691	5201329	As above
ZN05-SW500	Transect outside lease	511400	5201047	As above
Pen Bay ZS04	In-lease	509606	5199611	Day 1, 14, 28 & 56 after completion of medication
ZS04-NE35	Transect outside lease	509733	5199737	As above
ZS04-NE100	Transect outside lease	509790	5199786	As above
ZS04-NE500	Transect outside lease	510058	5200081	As above
ZS04-NW35	Transect outside lease	509351	5199850	As above
ZS04-NW100	Transect outside lease	509302	5199891	As above
ZS04-NW500	Transect outside lease	509007	5200163	As above
ZS04-SE35	Transect outside lease	509985	5199251	As above
ZS04-SE100	Transect outside lease	510025	5199205	As above
ZS04-SE500	Transect outside lease	510323	5198929	As above
Pen Bay ZS12	In-lease	509103	5199315	Day 1, 14, 28 & 56 after completion of medication
ZS12-NW35	Transect outside lease	508959	5199439	As above
ZS12-NW100	Transect outside lease	508913	5199490	As above
ZS12-NW500	Transect outside lease	508612	5199754	As above
ZS12-SW35	Transect outside lease	508906	5199105	As above
ZS12-SW100	Transect outside lease	508857	5199063	As above
ZS12-SW500	Transect outside lease	508578	5198773	As above
M6 (BEMP Site)	Ambient	515245	5204098	As above
C10.2	Ambient	509720	5201970	As above
C11.2	Ambient	512045	5199647	As above
SW	Ambient	506848	5196755	As above
LOMPT	Ambient	505131	5200810	As above
VENPT	Ambient	514491	5200332	As above
VS-W	Ambient beach	512504	5208539	Day 14 and 28 after completion of medication
VS-C	Ambient beach	512799	5208444	As above

Site Name	Site Category & Purpose	Easting	Northing	Sampling Day
VS-E	Ambient beach	513174	5208273	Day 14 and 28 after completion of medication
LJB-W	Ambient beach	512385	5188287	As above
LJB-C	Ambient beach	512618	5188187	As above
LJB-E	Ambient beach	512906	5188194	As above
CB-N	Ambient beach	515071	5196351	As above
CB-C	Ambient beach	514943	5195898	As above
CB-S	Ambient beach	514640	5195500	As above
RBB-W	Ambient beach	506718	5206827	As above
RBB-C	Ambient beach	507227	5206909	As above
RBB-E	Ambient beach	507657	5206891	As above

Sites provided in Table 2 are approximate only and are to be adjusted as necessary for practical reasons. Maps of the sampling site locations are provided in Attachment I.

Field personnel are to use best professional judgment on the day of sampling to adjust sites. Actual sampling sites for each sampling event must be recorded and included in reports required under section 2.6.

Sampling schedule:

Pen, transect and ambient sediment samples are to be collected from the end of the treatment period (= day 1 of sampling) and then 14, 28 and 56 days after the initial sampling date.

Subject to the prior approval from the EPA, any sediment sites returning results below LoR for two consecutive sampling events may be discontinued. If concentrations >LoR are detected on day 56, then the monitoring program may be extended for those sites. A survey calendar showing actual survey dates is provided in Attachment 2.

Sample collection:

Fresh, clean corers are to be used at each site. If this is not possible, the corer is to be thoroughly cleaned and rinsed with seawater between each site, with samples taken from the sites furthest from the lease area first (i.e. ambient sites, 100 m, 35 m and then cages).

At each sample site, triplicate cores (50 mm internal diameter and 75 mm depth) are to be taken. Duplicates for each site are to be composited and processed as one sample by the laboratory, with the third sample to be retained in frozen storage in the event that further analysis is required.

For each sample, the top 25 mm (surface) and subsequent 50 mm (bottom) are to be processed as separate sub-samples. For the duplicate composited samples, the top 25 mm from two replicate cores are to be combined into one jar. Similarly, the bottom 50 mm from the same two cores are to be combined into a single jar. The third replicate core is also to be split into separate surface and bottom sub-samples prior to freezing, to prevent migration of analyte(s) between sub-samples during storage.

Additional instructions provided by the analysing laboratory regarding sampling methodology, handling and storage are to be observed. Any inconsistencies between laboratory instructions and these requirements should be discussed with the EPA's Salmon Environmental Regulation Division.

Table 3: Number of sites and associated samples for lab analysis

Sites	No. sites	Depth layer	No. of sub-samples (triplicates)	No. samples analysed*	No. samples retained*
Cage sites	3	surface	9	3 (composite)	3
		bottom	9	3 (composite)	3
Transect sites	27	surface	81	27 (composite)	27
		bottom	81		54
Ambient sites	6	surface	18	6 (composite)	6
		bottom	18		12
Beach sites	12	surface	36	12 (composite)	12
		bottom	36		24
Total	48		288	51	141

* Initially surface & bottom sample analysis is required for cage sites only. For other sites, review of surface sample results will determine whether bottom samples need to be analysed. Until this determination has been made, all samples are to remain in storage.

There are four scheduled sampling events resulting in an initial estimate of **204 samples** to be analysed.

Table 4: Sample analyte(s)

Parameter	Abbreviation	Comments
Oxytetracycline Hydrochloride in sediment	OTC	Results must be reported using the lowest possible Limit of Reporting offered by the analysing laboratory

2.3 Wild fish sampling:

Wild fish are to be caught on day 1 and day 77 after completion of the medication event (1,000 degree days at bottom water temperatures ranging from 15.9 °C in February, 16.2 °C in March and 15.8 °C in April equating to 77 days).

A permit must be obtained to take fish for sampling under the *Living Marine Resources Management Act 1995*. Permits can be organised through the Wild Fisheries Management Branch of NRE Tas ([Fisheries Permits](#)).

Lease area

A minimum of 12 fish are to be collected from within each block of the lease site following medication (ideally these 12 fish should consist of 6 pelagic and 6 benthic species but in the event that only one species can be captured then a minimum of 12 fish of that species must be caught and processed).

'Pooled' samples for each species consisting of 3 individuals are to be compiled in the laboratory. Ideally, fish retained for sampling from the lease area should have pellets in the gut. This can be checked by purging from the gut or through gut content analysis once the tail section of the fish has been removed (see below). In the event that some of the collected fish are found not to contain pellets in the gut, these fish may be retained and analysed as part of a pooled sample.

A total of 4 pooled samples are to be processed for each block within the lease area.

Ambient sites

The ambient site for wild fish sampling is at BEMP Site M6 (Central Mid Channel), southwest of MF141 (SW), Lomas Point (LOMPT) and Ventenat Point (VENPT) The sampling protocol for ambient site fish

sampling is the same as for lease area sites, i.e. 12 fish should be sampled comprising 4 pooled samples of 3 individual fish.

Table 5: Summary of wild fish sampling

Site Name	Site category & purpose	Easting	Northing	Sampling Day	Comments
Zuidpool North - wild fish	In-lease	512208	5201912	Day 1 & 77 after completion of medication event	Sampling to be undertaken within central/northeastern part of the lease block
Zuidpool South - wild fish	In-lease	509512	5199513	Day 1 & 77 after completion of medication event	Sampling to be undertaken within northwestern and southwestern parts of the lease block
M6 (BEMP)	Ambient	515245	5204098	As above	
SW	Ambient	506848	5196755	As above	
LOMPT	Ambient	505131	5200810	As above	
VENPT	Ambient	514491	5200332	As above	

Sites provided in Table 5 are approximate only and are to be adjusted as necessary for practical reasons. Field personnel are to use best professional judgment on the day of sampling to adjust locations. Actual sampling sites for each sampling event must be recorded and included in reports required under section 2.6.

Note:

- If more than 3 species of fish are caught at a site, a minimum of one pooled sample per species will be required.
- Samples are to be taken first at the ambient site and then the lease site.

The species, length and gut content must be recorded for all fish retained and the GPS location of captures recorded. For lease area fish, the tail of the fish is to be cut off posterior to the anus and bagged prior to investigation of the fish's gut contents – if pellets are present then the sample is to be retained. Ambient site fish are to be processed in the same fashion, with species, length and gut content to be recorded for each sample (i.e. it is assumed that none of the fish sampled at ambient sites have any pellets in their gut, but this must be confirmed).

All sampling gear and sample preparation equipment used in the field must be thoroughly cleaned. Measures to minimise contamination risk from medicated pellets coming into contact with the blade should be implemented, such as using a separate blade/instrument when cutting the abdomen vs checking for gut contents. Alternatively, if this is not possible, the knife blade is to be thoroughly cleaned between cuts.

Flesh samples from the tail of the fish with pellets in their gut from lease area sites and from fish with no pellets from ambient sites are to be retained in separate bags with appropriate labelling (date, location, species, length, gut content), chilled immediately and frozen as soon as practicable.

Laboratory-based processing of samples will involve slicing a flesh sample off the tail section. The sample is to be removed by filleting the flesh off the backbone, thereby limiting inclusion of the vertebrae in the sample. The flesh sample is to have skin on and can include lateral/pin bones. The

remaining tail section is to be bagged and held in frozen storage in the event additional analyses are required.

Each individual fish sample within a pooled batch should be of approximately equal weight. The weight (g) of each fish sample and that of each pooled batch is to be recorded. Details of each fish within each pooled batch are also to be recorded and reported.

Table 6: Sample analyte(s)

Parameter	Abbreviation	Comments
Oxytetracycline Hydrochloride in fish tissue	OTC	Results must be reported using the lowest possible Limit of Reporting offered by the analysing laboratory

2.4 Water sampling

Water samples are to be collected from four ambient beach sites. The beaches selected are known to have had congealed fish oil globules wash ashore and include Verona Sands (VS), Roaring Bay Beach (RBB), Little Jetty Beach (LJB) and Conleys Beach (CB). Approximate coordinates for each site are available in Table 7. The samples for these sites are to be collected on day 14 and 28, after completion of the medication event (Attachment 2). Monitoring locations, duration and frequency may be increased if OTC is detected in the samples.

Table 7: Water sampling sites

Site Name	Site Category & Purpose	Easting	Northing	Sampling Day
VS	Ambient beach	512799	5208426	Day 14 and 28 after completion of medication
LJB	Ambient beach	512618	5188228	As above
CB	Ambient beach	514883	5195898	As above
RBB	Ambient beach	507227	5206883	As above

A single 5 L water sample is to be collected from each site and must be analysed for OTC in marine waters (Table 8). Samples must be collected using clean equipment and in a manner to avoid any contamination.

Each sample must be collected outside of the surf zone and the sampler should endeavour to minimise any sediment disturbance when collecting the sample. An extension pole may assist with the collection of the samples, particularly if collecting them from the shore.

Samples are to only be collected in labelled 5 L bottles provided by the analysing laboratory and should be delivered to that laboratory within 24 hours of collection.

The sampler must ensure that any additional instructions provided by the testing laboratory regarding sampling methodology, handling and storage are to be observed. Any inconsistencies between laboratory instructions and these requirements should be discussed with the EPA's Salmon Environmental Regulation Division.

Table 8: Sample analyte(s)

Parameter	Abbreviation	Comments
Oxytetracycline Hydrochloride in marine waters	OTC	Results must be reported using the lowest possible Limit of Reporting offered by the analysing laboratory

2.5 Congealed fish oil sampling

Samples of congealed fish oil globules are to be collected, if observed following a diligent search, when conducting the ambient beach monitoring on day 14 and 28 of this monitoring schedule (Attachment 2). These samples are to be analysed for OTC (Table 9). Monitoring locations, duration and frequency may be increased congealed fish oil globules continue to wash onto the beaches or if OTC is detected in the samples.

As advised under Section 2.2 (d) of this document, **only congealed fish oil globules present at the beach sediment sample sites are required to be collected and analysed.**

If present, collect the globule or a representative sample of all of the globules present, and place them in a labelled, sterile glass sample jar provided by the analysing laboratory. These samples should be delivered to that laboratory within 24 hours of collection.

The sampler must ensure that any additional instructions provided by the testing laboratory regarding sampling methodology, handling and storage are observed. Any inconsistencies between laboratory instructions and these requirements should be discussed with the EPA's Salmon Environmental Regulation Division.

Relevant sampling information, including site coordinates, must be recorded. Photos must be taken of the globules to be collected and should include an object, such as a ruler, to provide scale. All sampling information, results and photos collected must be reported to the EPA. Reporting requirements are listed in Attachment 3.

Table 9: Sample analyte(s)

Parameter	Abbreviation	Comments
Oxytetracycline Hydrochloride in fish oil	OTC	Results must be reported using the lowest possible Limit of Reporting offered by the analysing laboratory

2.6 Reporting results to EPA

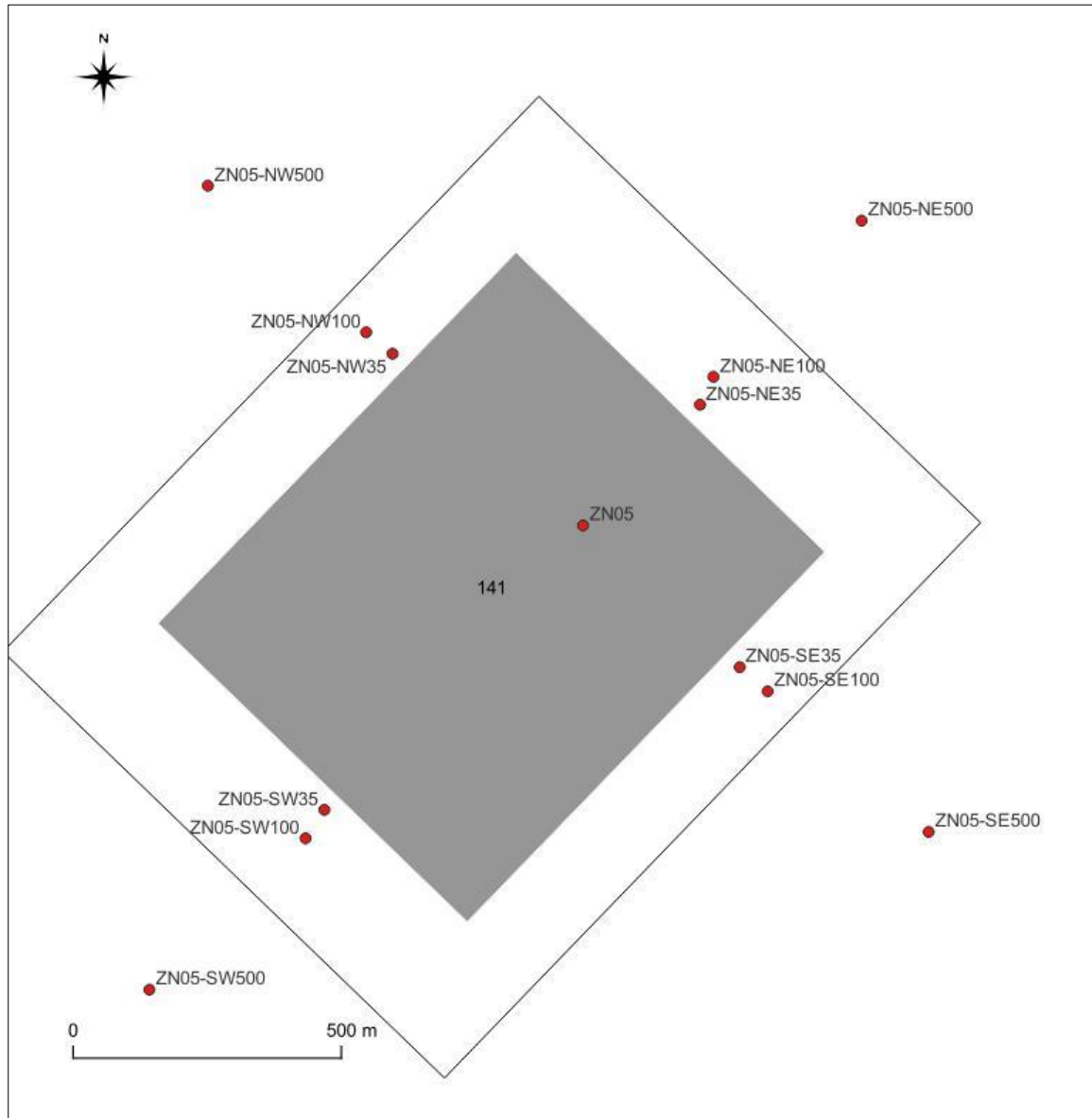
Laboratory reports are required to be sent to the EPA via email (salmon.reg@epa.tas.gov.au) after each round of sampling as soon as practicable.

A written report is to be submitted to EPA no later than 4 weeks after each sampling event or 2 weeks after the laboratory results becoming available, whichever is the later. Separate reports can be provided for sediment and wild fish sampling. Each report is to be accompanied by the raw data in an agreed form, submitted electronically. Report scope and format is to comply with the requirements of Attachment 3.

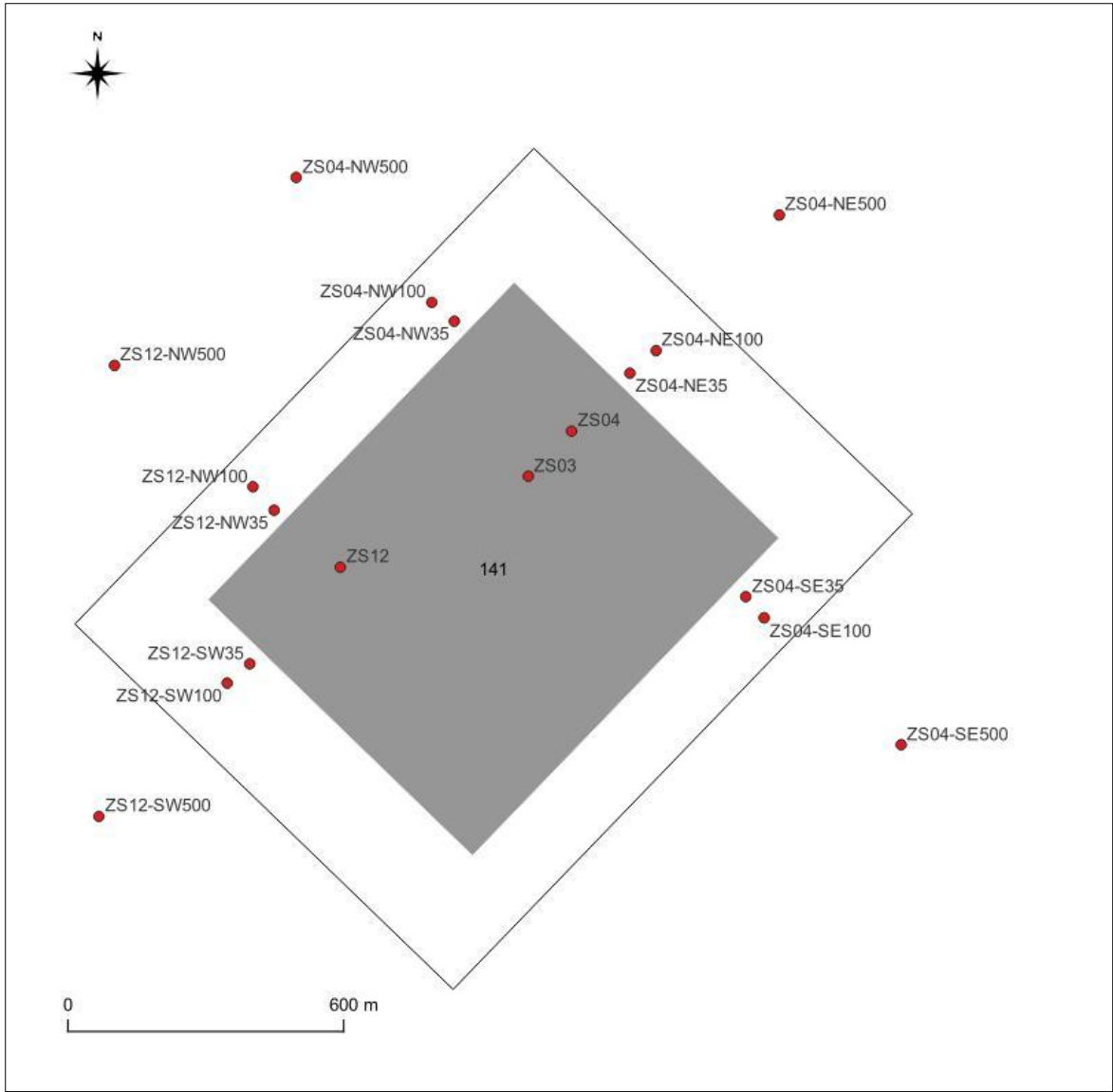
A consolidated report is to be provided no later than 4 weeks after the final sampling event (noting that the monitoring may be extended by the Director as described above). All monitoring results must be summarised, accompanied by a full analysis and graphical presentation of the data. Report scope and format is to comply with the requirements outlined in Attachment 3.

Attachment I

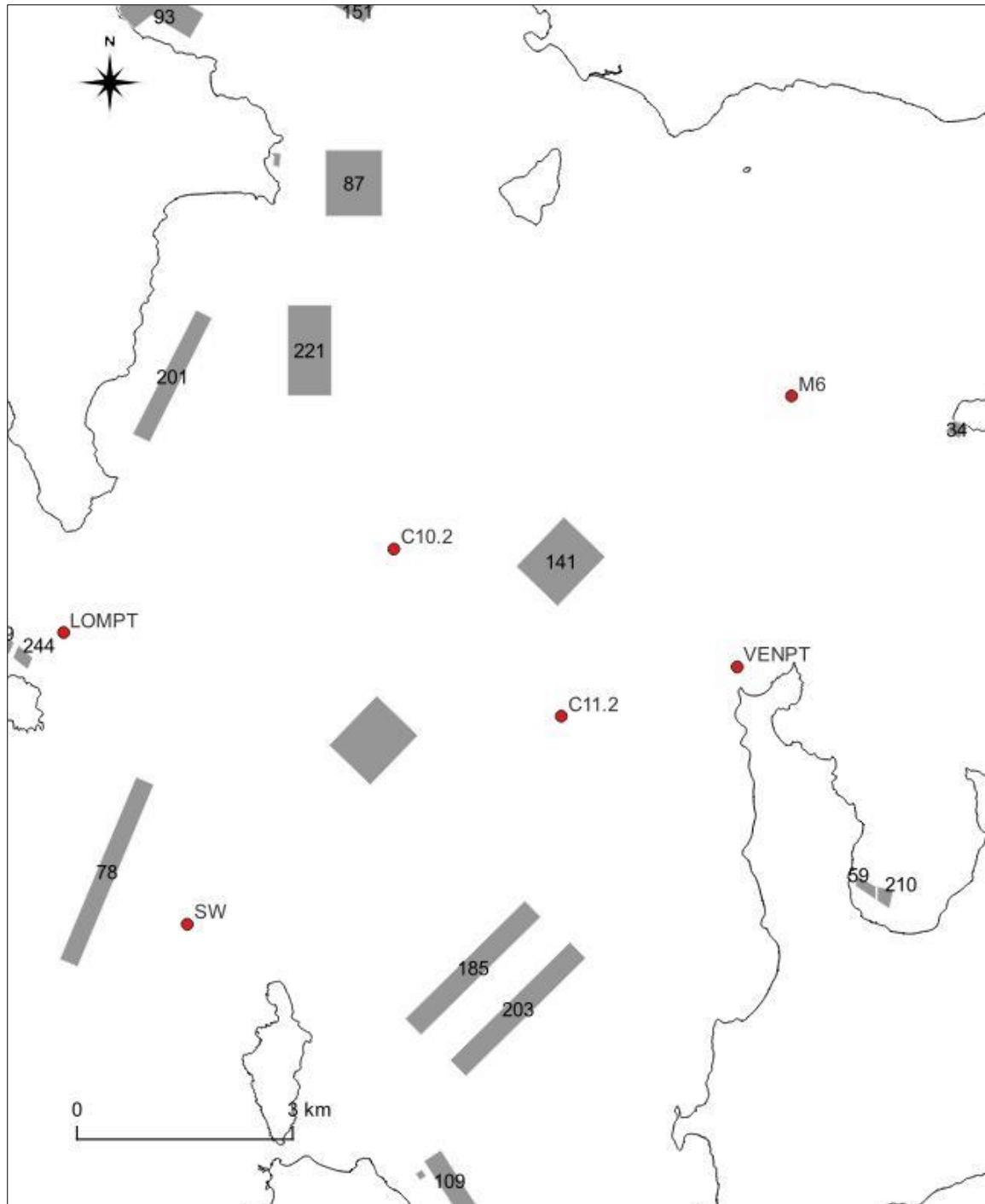
SAMPLE SITES



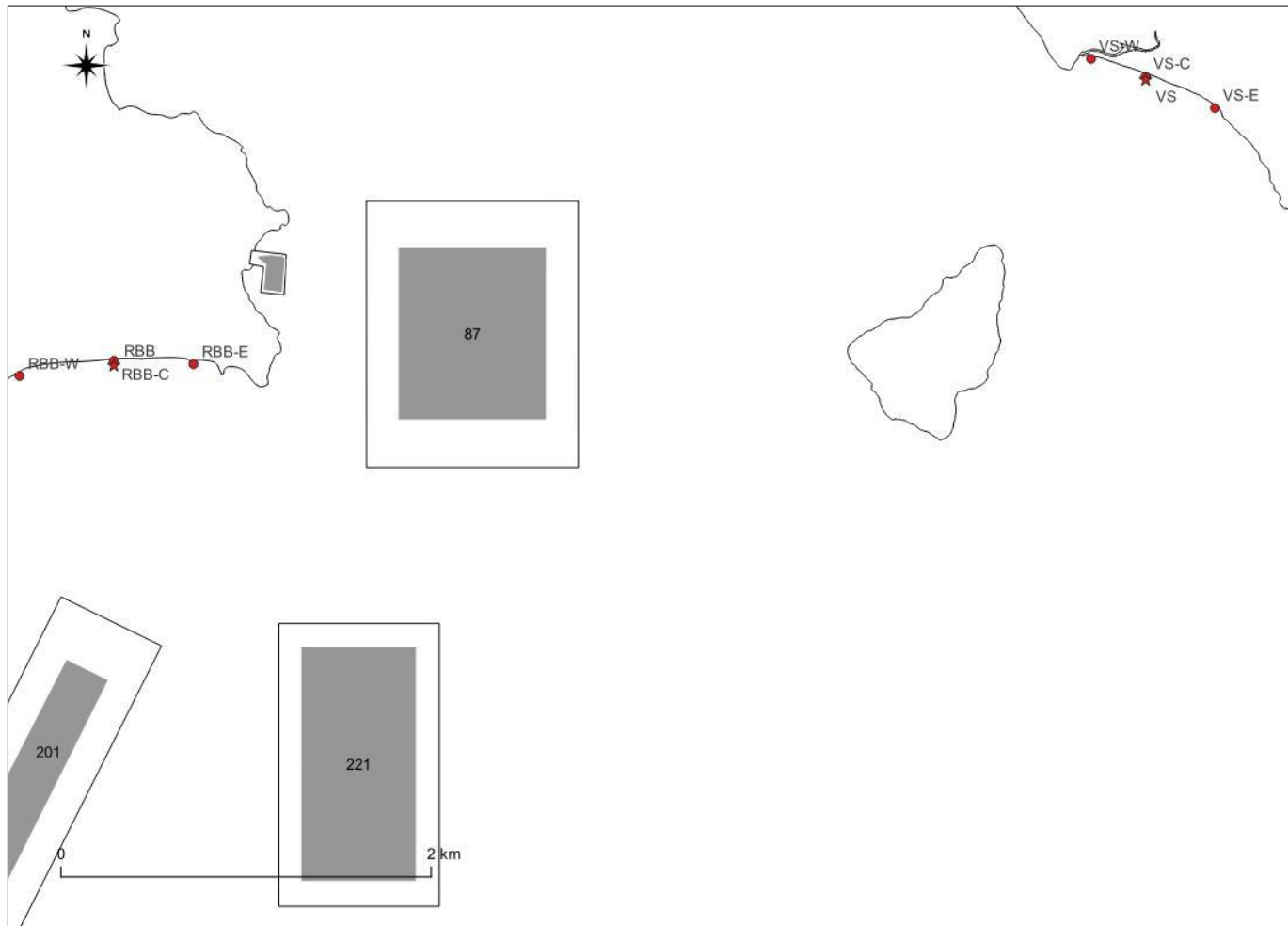
Map I: Map of antibiotic residue survey monitoring sites on the northern block (Zuidpool North) of Marine Farming Lease 141 (South of Zuidpool Rock).



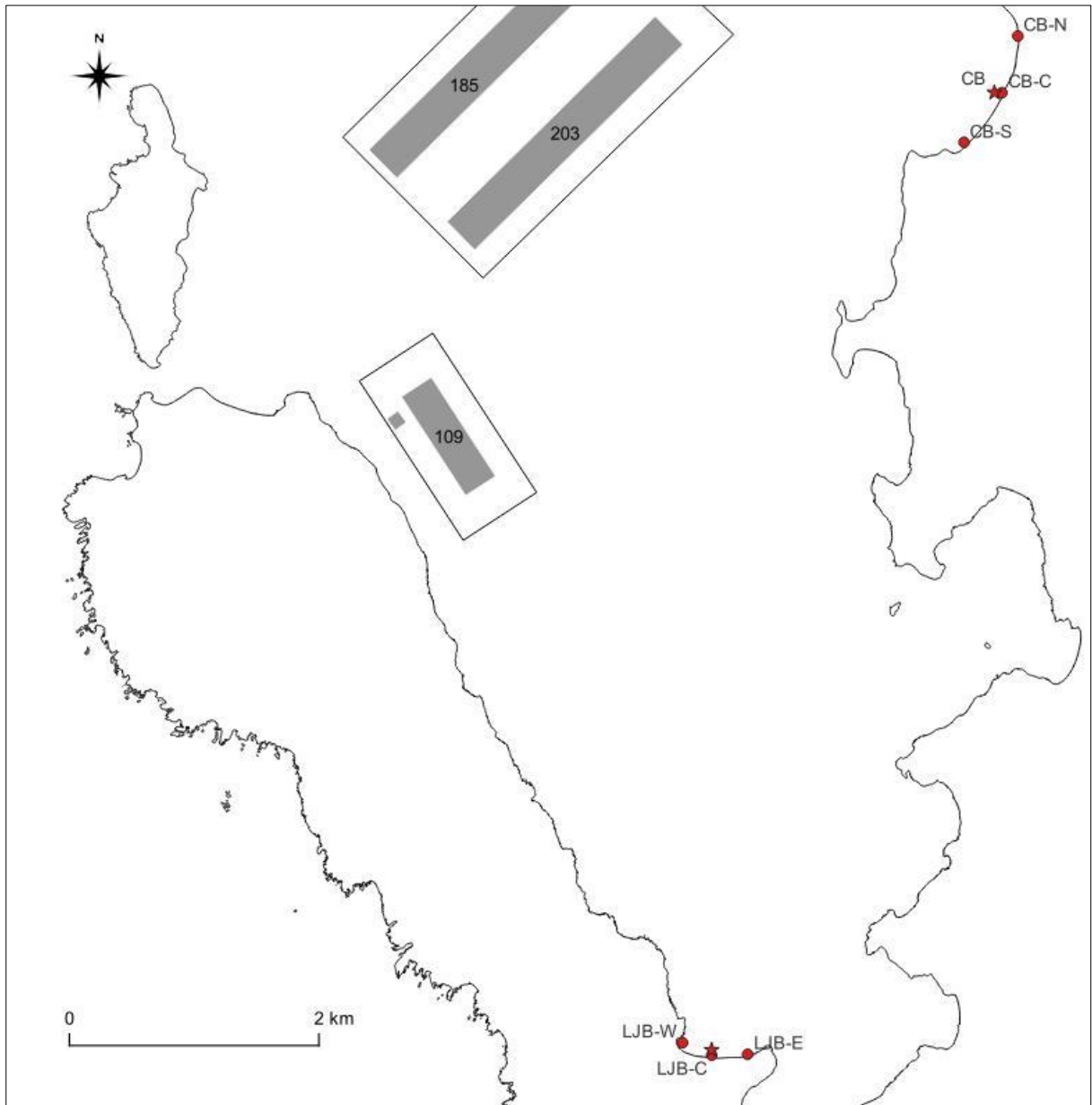
Map 2: Map of antibiotic residue survey monitoring sites on the southern block (Zuidpool South) of Marine Farming Lease 141 (South of Zuidpool Rock).



Map 3: Map of antibiotic residue survey monitoring sites external to Marine Farming Lease 141 (South of Zuidpool Rock).



Map 4: Map of additional antibiotic residue survey monitoring sites (sediment and water) on Roaring Bay Beach and Verona Sands (Pickup Beach), southern D'Entrecasteaux Channel.



Map 5: Map of additional antibiotic residue survey monitoring sites (sediment and water) on Conleys Beach and Lighthouse Jetty Beach, Bruny Island

Attachment 2

OVERVIEW OF SAMPLING PROGRAM ELEMENTS AND DATES

Date	Running days	Sediment sampling (pens)	Sediment sampling (transects & off-lease)	Wild fish sampling	Benthic Video Surveys (pens)	Sediment & globule sampling (beaches)	Water sampling (beaches)	Comments
13/02/2025	1 (Start)							
14/02/2025	2							
15/02/2025	3							
16/02/2025	4							
17/02/2025	5							
18/02/2025	6							
19/02/2025	7							
20/02/2025	8							
21/02/2025	9							
22/02/2025	10							
23/02/2025	11							
24/02/2025	12							
25/02/2025	13							
26/02/2025	14 (Finish)							
27/02/2025	15	X	X	X	X			Day 1 of sampling
28/02/2025	16							
01/03/2025	17							
02/03/2025	18							
03/03/2025	19							
04/03/2025	20							
05/03/2025	21							
06/03/2025	22							
07/03/2025	23							
08/03/2025	24							
09/03/2025	25							
10/03/2025	26							
11/03/2025	27							
12/03/2025	28	X	X			X	X	Day 14 of sampling
13/03/2025	29							
14/03/2025	30							
15/03/2025	31							
16/03/2025	32							
17/03/2025	33							
18/03/2025	34							
19/03/2025	35							
20/03/2025	36							
21/03/2025	37							
22/03/2025	38							
23/03/2025	39							
24/03/2025	40							
25/03/2025	41							
26/03/2025	42	X	X			X	X	Day 28 of sampling
23/04/2025	70	X	X					Day 56 of sampling
30/04/2025	77			X				Day 63 of sampling
30/05/2025		Submit Consolidated Report						

Attachment 3

REPORT REQUIREMENTS

Marine Farming Lease No: 141 (South of Zuidpool Rock)

Environmental Licence No: 9882/4

Licence holder/company: Huon Aquaculture Company Pty Ltd

Survey details: Date of field work, sampling personnel, contact persons, analysing laboratory and contextual information regarding weather conditions and other relevant observations.

Treatment details: Treatment type, quantity (active ingredient), application method, commencement date and duration for each cage.

Methods and results: The methods used for sample collection and analysis are to be described to allow verification that the requirements of this Monitoring Schedule have been fulfilled. The laboratory's Limits of Reporting in relation to the analyte(s) need to be specified.

Sediment sampling:

Laboratory results are to be tabulated with reference to site name & purpose (in-lease, out-of-lease transects, ambient sites), **site coordinates**, sampling depth, sampling date, and number of days from completion of antibiotics treatment. A description of sample collection (e.g. types of cores) and processing must also be provided. A map showing all sampling sites and site names is to be included.

Wild fish sampling:

Laboratory results are to be tabulated with reference to site name & purpose (in-lease & ambient site), **site coordinates**, sampling date and number of days from completion of antibiotics treatment. A description of sample collection and processing to obtain pooled samples must also be provided. A map showing all sample positions and sites of wild fish capture is to be included.

For each individual sample: fish species, fish length, sample weight and gut contents (pellets / other) need to be specified, highlighting those fish that were/were not processed for residue analysis. The table should be presented in the layout shown below.

Water sampling:

Laboratory results are to be tabulated with reference to site name & purpose (in-lease, out-of-lease transects, ambient sites), **site coordinates**, sampling depth, sampling date, and number of days from completion of antibiotics treatment. A description of sample collection and processing must also be provided. A map showing all sampling sites and site names is to be included.

Congeaed fish oil sampling (if collected):

Laboratory results are to be tabulated with reference to site name & purpose (in-lease, out-of-lease transects, ambient sites), **site coordinates**, sampling depth, sampling date, and number of days from completion of antibiotics treatment. A description of sample collection and processing must also be provided. A map showing all sampling sites and site names is to be included.

Shellfish sampling (if required):

Laboratory results are to be tabulated with reference to site names & purpose, sampling date, site coordinates and number of days from completion of antibiotics treatment. A description of sample collection and processing to obtain pooled samples (if relevant) must also be provided. A map showing all sample positions is to be included.

Exceedances of the maximum residue levels (MRL) need to be highlighted.

Electronic data submission:

- Excel spreadsheet showing monitoring results, including sampling date & time, site names and quantifiers in relation to numeric results, utilising the layout in Table 7;
- GPS co-ordinates for all sampling sites.

Consolidated report:

- The consolidated report must provide a summary of all monitoring results and relevant contextual information.
- Interpretation of results, in written and graphical form, to show trends over time and against relevant benchmarks must be included in relation to sediment, wild fish and shellfish monitoring.
- Details identifying specific personnel involved in the medication and monitoring program can be omitted.
- A map showing all sampling sites is to be provided.

Table 7: Wild fish sampling - reporting layout

Sample Date	Specimen No.	Pooled Sample Bag No.	Sample Weight	Fish Species (e.g. flathead, cocky salmon)	Fish Length (mm)	Gut Contents (e.g. pellets, other)
	1					
	2					
	3					
	4					
	5					
	6					
	7					
	8					
	9					
	10					
	11					
	12					