MF94 The Sheppards

Therapeutant Residue Monitoring Program

FINAL REPORT

June 2026

Report to:
Tassal Operations Pty Ltd

Prepared by:
AQUENAL PTY LTD

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Contents

1 Introduction 5
2 Design Overview, Survey Sites and Laboratory Analysis 6
2.1 Initial sampling design 6
2.2 Weekly sampling design 6
2.3 Laboratory analysis 11
3 Sediment Sampling 12
3.1 Survey sites and frequency 12
3.1.1 Initial sampling design 12
3.1.2 Weekly sampling design 12
3.2 Sample collection, processing and storage 15
3.3 Results 16
3.3.1 Initial sampling design 16
3.3.2 Weekly sampling design 16
4 Water Sampling 19
4.1 Sample sites and frequency 19
4.1.1 Initial sampling design 19
4.1.2 Weekly sampling design 19
4.2 Sample collection, processing and storage 19
4.3 Results 22
4.3.1 Initial sampling design 22
4.3.2 Weekly sampling design 22
5 Wild Fish Sampling 24
5.1 Survey sites and Frequency 24
5.1.1 Initial sampling design 24
5.1.2 Weekly sampling design 24
5.2 Sample collection, processing and storage 28
5.3 Results 29
5.3.1 Initial sampling design 29
5.3.2 Weekly sampling design 29
6 Benthic Video Surveys 32
6.1 Survey sites and frequency 32
6.2 Sample collection, processing and storage 32
6.3 Results 32
7 Summary 33
Appendices 34

Operational Summary

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1 Introduction

A Therapeutant Residue Monitoring Program (TRMP) was undertaken for the Marine Farming Lease MF94 The Sheppards (Environmental Licence 9869/5) located within the D'Entrecasteaux Channel region. There were two therapeutant treatment events conducted across MF94 between 15 February 2026 and 03 March 2026 using the antibiotic florfenicol (Table 1 and Table 2; see Appendix 1 for treatment details).

The TRMP was conducted in accordance with the specifications stipulated by the Tasmanian Environment Protection Authority (EPA) in *Therapeutant Residue Monitoring Schedule – Marine Farming Lease No. 94* (herein TRM Schedule MF94), issued 20 February 2026 and *Addendum - Therapeutant Residue Monitoring Schedule – MF94* (herein TRM Schedule MF94 Addendum), issued 27 February 2026 (see Table 1).

The monitoring schedules were developed to assess potential environmental impacts from the treatment events and were based on the information provided in the Veterinary Authority (VA), the environment in which the lease is located, previous treatment history, and research into the active ingredient and its potential environmental fate. The TRMP included the following components: sediment sampling, water sampling, wild fish sampling, and benthic video surveys. Samples were taken at monitoring sites within the lease boundary (internal sites) and outside the lease boundary (external sites including compliance sites, transect sites, reference sites). The frequency of monitoring changed between the initial sampling schedule (TRM Schedule MF94; i.e. post-treatment on Day 1) and the subsequent addendum which stipulated weekly sampling (TRM Schedule MF94 Addendum; Table 1). The initial sampling design was developed for a single treatment event and the weekly sampling design was developed and issued to monitor multiple and any future treatment events on the lease.

This report provides survey details, treatment details, methods and results of the TRMP for MF94 The Sheppards. The report includes a summary of all monitoring results and relevant contextual information.

2 Design Overview, Survey Sites and Laboratory Analysis

There were two treatment events (T1, T2) at MF94 The Sheppards between 15 February 2026 and 3 March 2026 (Table 1; Table 2; Figure 1). This section provides an overview of the initial sampling design and the weekly sampling design stipulated in the relevant Therapeutant Residue Monitoring Schedules (Table 1). A detailed explanation of sampling sites, frequency of collection, methodology and results for the four survey components are documented iteratively in Sections 3-6. Field conditions for each sampling day are provided in Appendix 3.

2.1 Initial sampling design

The initial Therapeutant Residue Monitoring Schedule was associated with treatment T1 and incorporated four components (i.e. sediment, water quality, wild fish and benthic video). These components were sampled at a range of internal, external and reference sites in the vicinity of MF94 The Sheppards (Figure 2). Internal sites included treated pen-bays and the lease centroid (Figure 2b). External sites included compliance sites 35 m from the lease boundary and 100 m and 500 m transect sites associated with these compliance sites (Figure 2b). Reference sites were located further afield and were associated with Broadscale Environmental Monitoring Program (BEMP) sites (Figure 2a). Wild fish were sampled from zones defined by buffers at increasing distance from the lease boundary (e.g. 1 km, 3 km; Figure 3). Benthic video surveys associated with treatment T1 were conducted at all treated pen bays pre-treatment and post-treatment.

For surveys associated with treatment T1, the frequency of sampling was organised around the number of days post the conclusion of the treatment (e.g. Day 1; Table 1). Sampling for the T1 treatment shifted from the initial sampling design to the weekly sampling design outlined below after the Day 1 sampling event. Sites sampled in the initial sampling design carried through to the weekly sampling design.

2.2 Weekly sampling design

For surveys associated with treatments T1 and T2 there were four components (i.e. sediment, water quality, wild fish and benthic video). Samples were taken from a selection of internal, external and reference sites established under the initial survey design described in Section 2.1 (Figure 2; Figure 3). The frequency of sampling for sediments, water and wild fish was organised around weekly surveys (i.e. Week 1, Week 2…..Week 5; Table 1). Benthic video surveys associated with treatment T2 were conducted at all treated pen bays pre-treatment and post-treatment.

Subject to prior approval from the EPA, the initial and weekly schedules allowed for the discontinuation of some scheduled sampling events for sites returning results ‘below the Limit of Reporting’ (LoR) for two consecutive sampling events (see Section 2.3; Table 3). From 18 March 2026

onwards, sediment sampling requirements were altered by EPA to require a single result below the LoR before discontinuation.

It should be noted that the BEMP_M3 reference site for MF94 The Sheppards (3380 m from the lease, i.e. >3 km; Figure 2; Figure 3) was in relatively close proximity to the MF142 Roberts Point lease (1240 m from the lease; i.e. 1-3 km; Figure 2; Figure 3) which underwent treatment with florfenicol for three pen bays[1] during the same period as the T2 treatment at MF94 The Sheppards (i.e. 22/02/2026-03/03/2026; Figure 1). In this report, results from sediment, water and wild-fish surveys from BEMP_M3 are discussed in relation to the relative influence of treatment events from MF94 The Sheppards and MF142 Roberts Point.

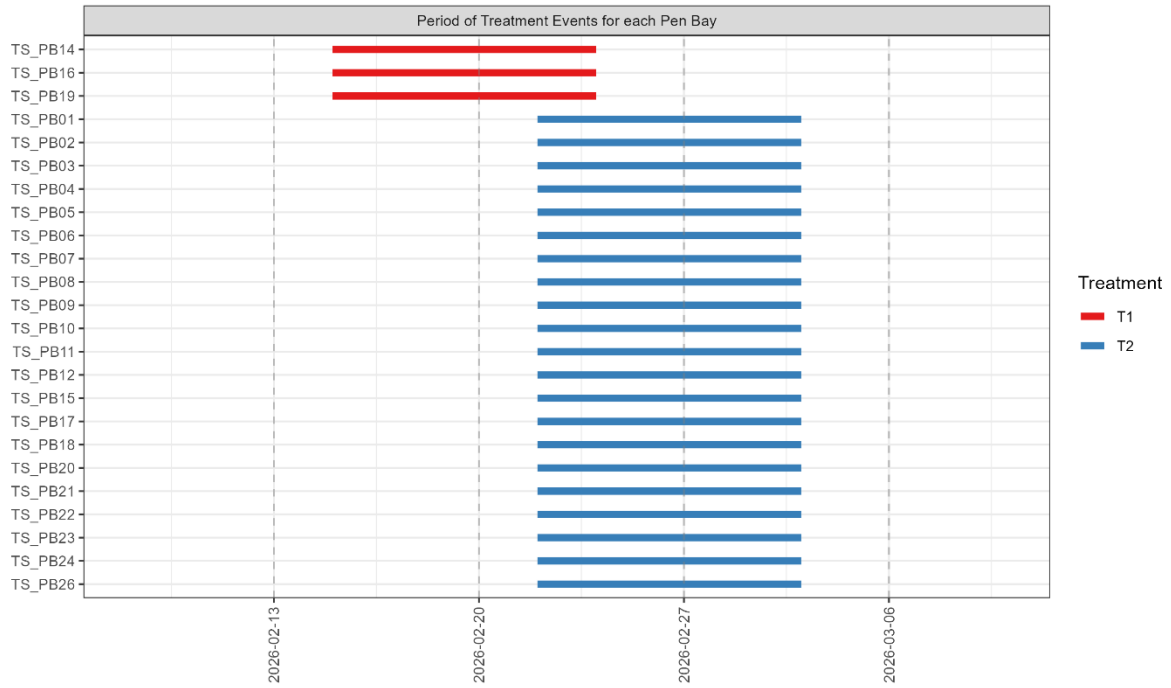

Figure 1: Date and duration of treatment events associated with pen bays at lease MF94 The Sheppards (see Table 2).

[1] Aquenal (2026) MF142 Roberts Point Therapeutant Residue Monitoring Program. Final Report. June 2026. Report to Tassal Operations Pty Ltd.

Table 1: Therapeutant treatments and associated Veterinary Authorities for treatment of finfish with florfenicol at MF94 The Sheppards. The schedule associated with each treatment event is indicated (*TRM Schedule MF94* = *Therapeutant Residue Monitoring Schedule MF94*; *TRM Schedule MF94 Addendum* = *Addendum - Therapeutant Residue Monitoring Schedule MF94*), along with the broad sampling design (i.e. 'initial' or 'weekly'). Additional details of treatment events are in Appendix 1.

Treatment	Veterinary Authority	Start Date	Schedule	Sampling Design
T1	26-Tx-M04	15/02/2026	*TRM Schedule MF94*	Initial sampling design: post- treatment (e.g. Day 1)
T2	26-Tx-M08	22/02/2026	*TRM Schedule MF94 Addendum*	Weekly sampling design

Table 2: Date and duration of treatment events associated with pen bays at lease MF94 The Sheppards. Sampling requirements (i.e. water, sediment and benthic video) are shown for each pen bay.

Treatment	Site	Start Date	Finish Date	Sampling	
				Water and Sediment	Benthic Video
T1	**TS_PB14**	15/02/2026	24/02/2026	Y	Y
	TS_PB16	15/02/2026	24/02/2026		Y
	TS_PB19	15/02/2026	24/02/2026	Y	Y
T2	**TS_PB01**	22/02/2026	3/03/2026	Y	Y
	TS_PB02	22/02/2026	3/03/2026		Y
	TS_PB03	22/02/2026	3/03/2026		Y
	TS_PB04	22/02/2026	3/03/2026	Y	Y
	TS_PB05	22/02/2026	3/03/2026	Y	Y
	TS_PB06	22/02/2026	3/03/2026		Y
	TS_PB07	22/02/2026	3/03/2026	Y	Y
	TS_PB08	22/02/2026	3/03/2026		Y
	TS_PB09	22/02/2026	3/03/2026	Y	Y
	TS_PB10	22/02/2026	3/03/2026		Y
	TS_PB11	22/02/2026	3/03/2026	Y	Y
	TS_PB12	22/02/2026	3/03/2026		Y
	TS_PB15	22/02/2026	3/03/2026	Y	Y
	TS_PB17	22/02/2026	3/03/2026		Y
	TS_PB18	22/02/2026	3/03/2026		Y
	TS_PB20	22/02/2026	3/03/2026	Y	Y
	TS_PB21	22/02/2026	3/03/2026		Y
	TS_PB22	22/02/2026	3/03/2026	Y	Y
	TS_PB23	22/02/2026	3/03/2026	Y	Y
	TS_PB24	22/02/2026	3/03/2026		Y
	TS_PB26	22/02/2026	3/03/2026	Y	Y

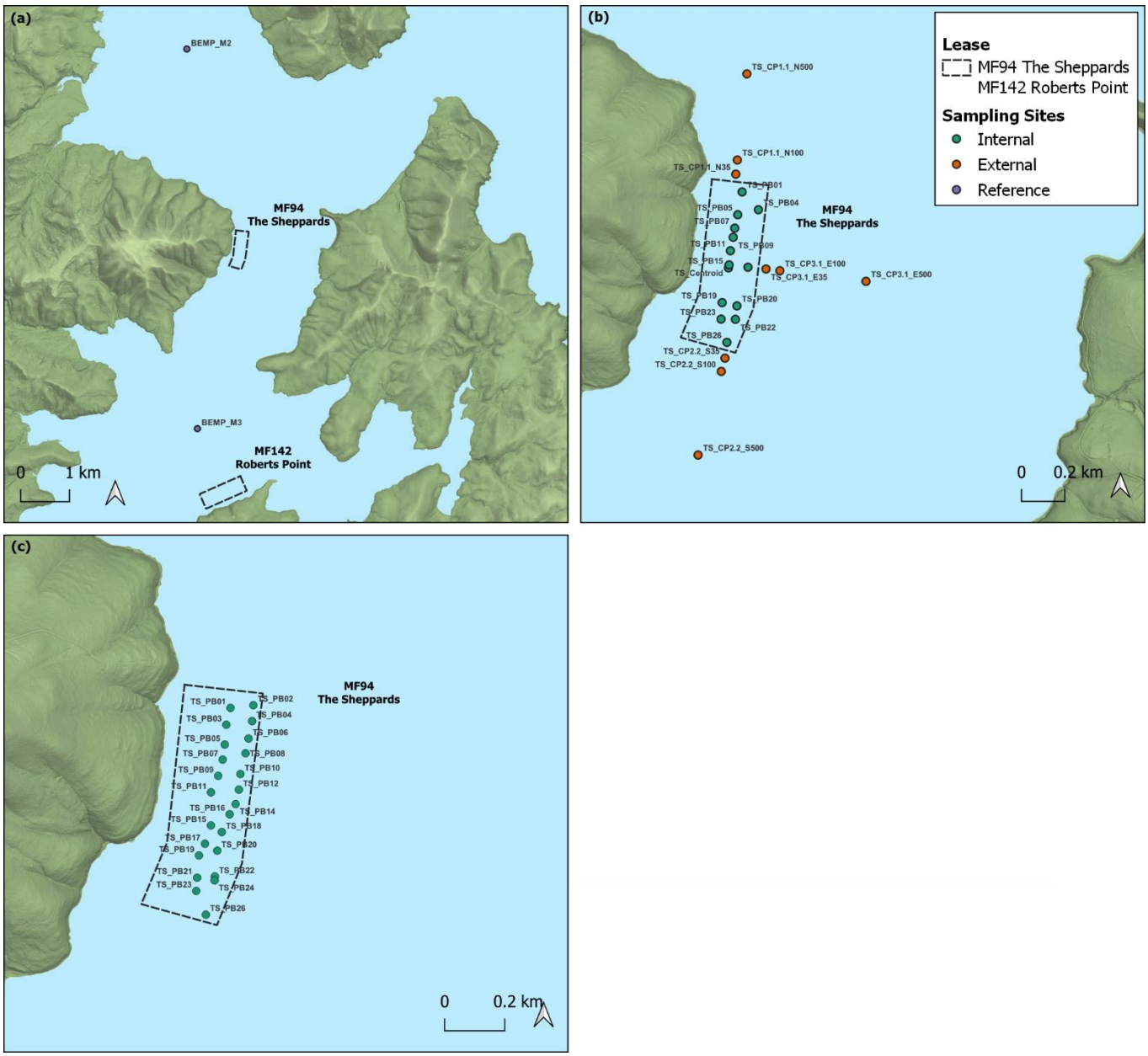

Figure 2: Sampling sites used in water quality sediment and benthic video surveys, including (a) overview of the lease and survey area in the D'Entrecasteaux Channel region, highlighting reference sites; (b) internal and external sites associated with MF94 The Sheppards; and (c) benthic video survey sites. In (a) and (b) mapped coordinates for external and reference sites are from the schedules and mapped coordinates for the pen bays are from the first instance of their sampling in the field (see Appendix 2). In (c) mapped coordinates are those sampled in pre-treatment surveys.

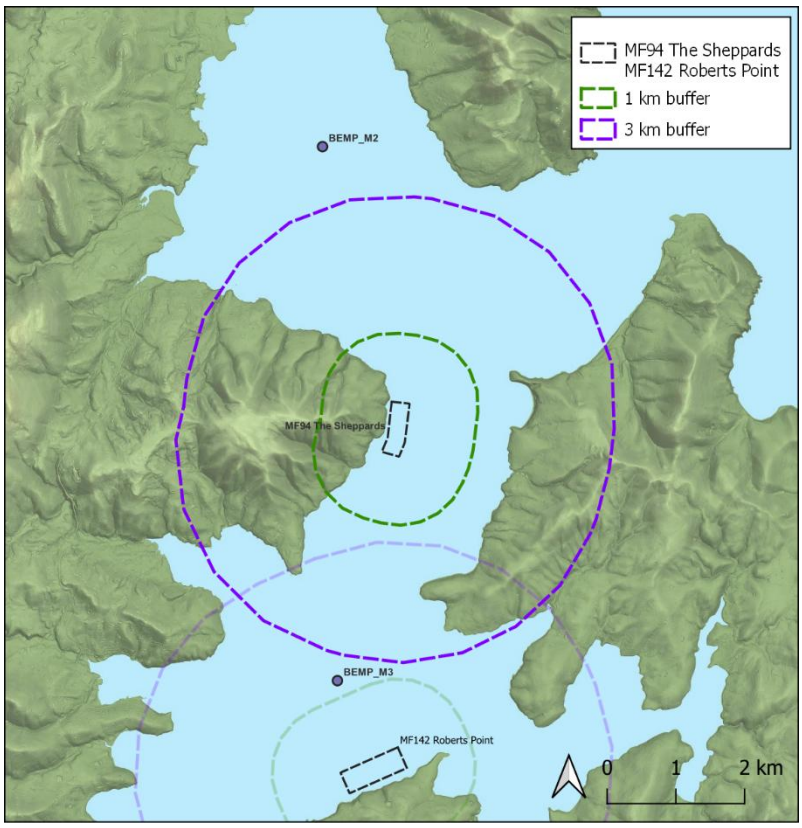

Figure 3: Reference sites and survey zones for wild fish sampling including zones defined by 1 km and 3 km buffers around MF94 The Sheppards. Faded buffer zones around MF142 Roberts Point illustrate the location of BEMP_M3 in the 1-3 km zone for this lease which was treated concurrently with T2 at MF94 The Sheppards.

2.3 Laboratory analysis

Analysis of florfenicol and florfenicol amine in environmental samples was carried out by Analytical Services Tasmania (AST). Florfenicol is the antibiotic that was used to treat *Piscirickettsia salmonis*. Florfenicol amine is the primary, non-antibiotic metabolite and the principal breakdown product of florfenicol in the surrounding environment. When measuring florfenicol in fish, the fish samples were reported as the "sum of florfenicol and its metabolites measured as florfenicol amine", which is due to the method required to extract florfenicol from the fish tissues. The Limit of Reporting (LoR) that is referred to throughout this report is the lowest concentration of a substance that the laboratory can reliably and consistently measure (Table 3). Further information regarding the analytical methods for measuring florfenicol can be found on the AST website[2].

Table 3: AST laboratory Limits of Reporting for florfenicol.

Analyte	Sample Type		
	Water (µg/L*)	Sediment (mg/Kg#)	Fish Tissue (mg/Kg#)
Florfenicol	0.1	0.005	-
Florfenicol amine	0.1	0.01	-
Sum of florfenicol and its metabolites measured as florfenicol amine	-	-	0.01

* 1 µg/L is equal to 1 part per billion (ppb).
# 1 mg/kg is equal to 1 part per million (ppm).

[2] Florfenicol testing at Analytical Services Tasmania

3 Sediment Sampling

3.1 Survey sites and frequency

Sediment sampling was undertaken by Aquenal at an array of sites including internal pen bays, external and reference locations (Figure 2; Appendix 2) at a frequency defined by the initial and weekly sampling designs (Table 4; see Section 2.1-2.2).

3.1.1 Initial sampling design

Sampling associated with treatment T1 was organised around pre-defined surveys conducted post-treatment (e.g. Day 1; Table 4) at an array of sites including a subset of treated pen bays (designated by the EPA), external 35 m compliance and associated 100 m and 500 m transect sites, and reference sites (Table 4). Subject to prior approval from the EPA, any sediment sites returning results below the LoR (specified in Table 3) for two consecutive sampling events were discontinued. For sampling associated with T1 treatments, surface and depth sections of sediment cores were collected and analysed at all sites.

Sampling for the T1 treatment shifted from the initial sampling design to the weekly sampling design, outlined below, after the Day 1 sampling event. Sites sampled in the initial sampling design carried through to the weekly sampling design. Figure 4 illustrates the initial sampling design and the relationship with each treatment event. Note that the Day 1 sampling event was conducted during the T2 treatment (Figure 4).

3.1.2 Weekly sampling design

For sediment surveys associated with treatments T1 and T2**,** unless otherwise specified by the Director in writing, all sediment sites specified in Table 4 were sampled weekly, until all treatments on the Lease were completed AND two consecutive results below the LoR were recorded. Pen bays were sampled until the treatment on those pen bays had ended and until two consecutive results below LoR were returned. External and reference sites were sampled until all treatments on the lease were completed and two consecutive results below the LoR were recorded. Sediment sampling requirements were changed to require sampling until a single result below the LoR was returned, according to the EPA directive *Update to Requirements for Therapeutant Residue Sediment Sampling* (EPA email correspondence, 18 March 2026). This change was applicable to sampling from Week 3 onwards (see Figure 4).

Sediment samples were collected from 50% of the treated pen bays listed for each Veterinary Authority (VA). Where possible, pen bays selected for sediment sampling were spread spatially. The same pen bays were used for consecutive sampling events.

Week 1 and Week 2 samples for BEMP_M2 and BEMP_M3 were collected on a different date as part of the concomitant TRMP for MF142 Roberts Point (see Figure 4).

Each surface sample was analysed, while depth samples were retained (frozen) for future analysis if required. If a surface sample returned a florfenicol or florfenicol amine result greater than the LoR, the associated depth sample for that site and sample date was also analysed. If a result greater than the LoR was returned for a sample collected from a 35 m compliance site, the sediment samples from the 100 m and 500 m transect sites were also collected and analysed during the next sampling round. Sediment sampling at the 100 m and 500 m transect sites was not required unless a 35 m compliance site sediment result was greater than the LoR.

Figure 4 illustrates the weekly sampling design and the relationship with each treatment event.

Table 4: Sediment sampling design and frequency associated with treatment events at MF94 The Sheppards. Post treatment sampling under the initial sampling design included predefined surveys (e.g. Day 1). Sampling was not required for cells highlighted in blue.

Location	Site	Initial Design	Weekly Design
		T1	T1+T2
Internal (Pen Bay)	TS_PB14	Day 1	Weekly
	TS_PB19	Day 1	Weekly
	TS_PB01		Weekly
	TS_PB04		Weekly
	TS_PB05		Weekly
	TS_PB07		Weekly
	TS_PB09		Weekly
	TS_PB11		Weekly
	TS_PB15		Weekly
	TS_PB20		Weekly
	TS_PB22		Weekly
	TS_PB23		Weekly
	TS_PB26		Weekly
External (35 m)	TS_CP1.1_N35	Day 1	Weekly
	TS_CP3.1_E35	Day 1	Weekly
	TS_CP2.2_S35	Day 1	Weekly
External (100 m)	TS_CP1.1_N100	Day 1	
	TS_CP3.1_E100	Day 1	
	TS_CP2.2_S100	Day 1	
External (500 m)	TS_CP1.1_N500	Day 1	
	TS_CP3.1_E500	Day 1	
	TS_CP2.2_S500	Day 1	
Reference	BEMP_M2	Day 1	Weekly
	BEMP_M3	Day 1	Weekly

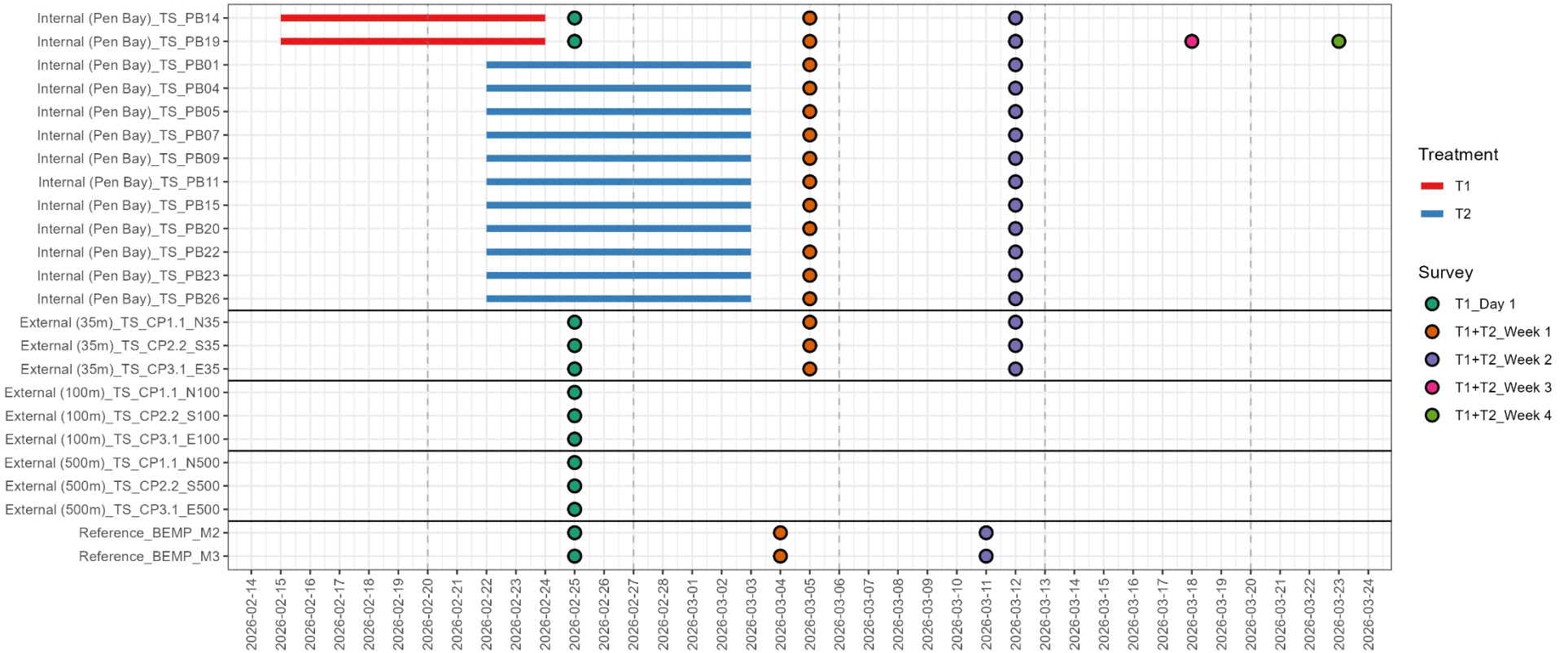

Figure 4: Sampling events for sediment surveys associated with the initial sampling design (treatment T1; i.e. post treatment on Day 1) and the weekly sampling design (i.e. treatments T1 and T2; weekly samples) at MF94 The Sheppards. Sites sampled included treated internal pen bays, external sites and reference sites. Horizontal bars illustrate the timing and duration of treatment events (see Table 2; Figure 1). Week 1 and Week 2 samples for BEMP_M2 and BEMP_M3 were collected on a different date as part of the concurrent TRMP for MF142 Roberts Point (see Figure 4).

3.2 Sample collection, processing and storage

Depending on sediment type, a Craib corer or Quad corer was used to collect triplicate sediment cores at each site. Core samples (75 mm minimum length) were collected in clean Perspex cores (Figure 5). The corer was thoroughly cleaned and rinsed with seawater between each site, with samples taken from the sites furthest from the lease boundary first (i.e. reference sites, 500 m, 100 m, 35 m and then internal (pen bay) sites to minimise contamination risk.

Cores were processed onboard following collection at each site. Samples were split into surface (top 25 mm) and bottom (bottom 50 mm) sections and composited according to the EPA schedule. For the duplicate composited samples, the top 25 mm from two replicate cores were combined into one jar. Similarly, the bottom 50 mm from the same two cores were combined into a single jar. The third replicate core sample was separated into the 25 mm and 50 mm sections. Samples were held in an Eski on board the work vessel before being transferred to frozen storage at the end of each field day. The composite samples were delivered to Analytical Services Tasmania (AST) as soon as practicable, while samples from the third replicate core were retained as archived samples in frozen storage.

Samples were tested for both florfenicol (FFC) and florfenicol-amine (FFA) residues by Analytical Services Tasmania.

Figure 5: (a) Craib corer; (b) Quad corer in the field with sediment samples collected in Perspex cores; (c) Perspex cores prior to processing; and (d) processing equipment onboard work vessel.

3.3 Results

3.3.1 Initial sampling design

For the Day 1 post-treatment survey associated with the T1 treatment event, florfenicol (FFC) and/or florfenicol amine (FFA) concentrations were below the LoR for all but one sediment sample, which was collected from the surface of pen bay TS_PB19 and recorded a FFC concentration above the LoR (Table 5; Table 6). The corresponding bottom sample collected from this site on Day 1 recorded a FFC concentration below the LoR (Table 5). After Day 1, sampling for the T1 treatment shifted from the initial sampling design to the weekly sampling design (see section 3.3.2).

3.3.2 Weekly sampling design

For weekly surveys associated with the T1 and T2 treatment events, florfenicol (FFC) and/or florfenicol amine (FFA) concentrations were below the LoR for the majority of sediment samples analysed at internal, external and reference sites (82 of 88 samples; Table 5; Table 6). All six samples with FFC and/or FFA concentrations above the LoR were collected at treated pen bays (Table 5; Table 6), with four samples (three surface, one bottom) collected in Week 1 (i.e. two days post-treatment; Figure 4), one sample collected in Week 2 (i.e. nine days post treatment; Figure 4) and one sample collected in Week 3 (i.e. fifteen days post treatment; Figure 4). All samples collected in weekly surveys from external and reference sites recorded FFC and FFA concentrations below the LoR (Table 5; Table 6).

Table 5: Florfenicol (FFC) concentration (mg/kg DMB) in sediment samples collected for the initial sampling design (treatment T1; Day 1) and weekly sampling design (treatments T1+T2; weekly samples) at the surface (S) and at depth (bottom, B) at internal, external, and reference sites at MF94 The Sheppards. Green shaded cells indicate sites where the sample was below the limit of reporting (LoR, i.e. <0.005). The LoR is the lowest concentration of a substance that the laboratory can reliably and consistently measure. Blue shaded cells indicate sites where sampling and analysis was not required (refer to Table 4 for details). Unshaded cells indicate the sample result was above the LoR for that sample.

Location	Site	Florfenicol (mg/kg DMB)									
		T1		T1+T2							
		Day 1		Week 1		Week 2		Week 3		Week 4	
		2026-02-25		2026-03-04* 2026-03-05		2026-03-11* 2026-03-12		2026-03-18		2026-03-23	
		S	B	S	B	S	B	S	B	S	B
Internal (Pen Bay)	TS_PB14	<0.005	<0.005	0.015	0.006	<0.005	<0.005				
	TS_PB19	0.030	<0.005	<0.005		0.007	<0.005	0.006	<0.005	<0.005	
	TS_PB01			<0.005		<0.005					
	TS_PB04			<0.005		<0.005					
	TS_PB05			<0.005		<0.005					
	TS_PB07			<0.005		<0.005					
	TS_PB09			<0.005		<0.005					
	TS_PB11			<0.005		<0.005					
	TS_PB15			<0.005		<0.005					
	TS_PB20			<0.005	<0.005	<0.005					
	TS_PB22			<0.005		<0.005					
	TS_PB23			<0.005	<0.005	<0.005					
	TS_PB26			<0.005		<0.005					
External (35 m)	TS_CP1.1_N35	<0.005	<0.005	<0.005		<0.005					
	TS_CP2.2_S35	<0.005	<0.005	<0.005		<0.005					
	TS_CP3.1_E35	<0.005	<0.005	<0.005		<0.005					
External (100 m)	TS_CP1.1_N100	<0.005	<0.005								
	TS_CP2.2_S100	<0.005	<0.005								
	TS_CP3.1_E100	<0.005	<0.005								
External (500 m)	TS_CP1.1_N500	<0.005	<0.005								
	TS_CP2.2_S500	<0.005	<0.005								
	TS_CP3.1_E500	<0.005	<0.005								
Reference	BEMP_M2	<0.005	<0.005	<0.005		<0.005					
	BEMP_M3	<0.005	<0.005	<0.005		<0.005					

***** *Samples from BEMP_M2 and BEMP_M3 were collected on a different date as part of the concurrent TRMP for MF142 Roberts Point (Figure 4).*

Table 6: Florfenicol amine (FFA) concentration (mg/kg DMB) in sediment samples collected for the initial sampling design (treatment T1; Day 1) and weekly sampling design (treatments T1+T2; weekly samples) at the surface (S) and at depth (bottom, B) at internal, external, and reference sites at MF94 The Sheppards. Green shaded cells indicate sites where the sample was below the limit of reporting (LoR, i.e. <0.01). The LoR is the lowest concentration of a substance that the laboratory can reliably and consistently measure. Blue shaded cells indicate sites where sampling and analysis was not required (refer to Table 4 for details). Unshaded cells indicate the sample result was above or equal to the LoR for that sample.

Location	Site	Florfenicol amine (mg/kg DMB)									
		T1		T1+T2							
		Day 1		Week 1		Week 2		Week 3		Week 4	
		2026-02-25		2026-03-04* 2026-03-05		2026-03-11* 2026-03-12		2026-03-18		2026-03-23	
		S	B	S	B	S	B	S	B	S	B
Internal (Pen Bay))	TS_PB14	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01				
	TS_PB19	<0.01	<0.01	<0.01		<0.01	<0.01	<0.01	<0.01	<0.01	
	TS_PB01			<0.01		<0.01					
	TS_PB04			<0.01		<0.01					
	TS_PB05			<0.01		<0.01					
	TS_PB07			<0.01		<0.01					
	TS_PB09			<0.01		<0.01					
	TS_PB11			<0.01		<0.01					
	TS_PB15			<0.01		<0.01					
	TS_PB20			0.01	<0.01	<0.01					
	TS_PB22			<0.01		<0.01					
	TS_PB23			0.01	<0.01	<0.01					
	TS_PB26			<0.01		<0.01					
External (35 m)	TS_CP1.1_N35	<0.01	<0.01	<0.01		<0.01					
	TS_CP2.2_S35	<0.01	<0.01	<0.01		<0.01					
	TS_CP3.1_E35	<0.01	<0.01	<0.01		<0.01					
External (100 m)	TS_CP1.1_N100	<0.01	<0.01								
	TS_CP2.2_S100	<0.01	<0.01								
	TS_CP3.1_E100	<0.01	<0.01								
External (500 m)	TS_CP1.1_N500	<0.01	<0.01								
	TS_CP2.2_S500	<0.01	<0.01								
	TS_CP3.1_E500	<0.01	<0.01								
Reference	BEMP_M2	<0.01	<0.01	<0.01		<0.01					
	BEMP_M3	<0.01	<0.01	<0.01		<0.01					

** Samples from BEMP_M2 and BEMP_M3 were collected on a different date as part of the concurrent TRMP for MF142 Roberts Point (Figure 4).*

4 Water Sampling

4.1 Sample sites and frequency

Water sampling was undertaken by Marine Solutions at an array of sites including internal, external and reference sites (Figure 2; Appendix 2) at a frequency defined by the initial and weekly sampling designs (Table 7; see Section 2.1-2.2).

4.1.1 Initial sampling design

For sampling associated with treatment T1, water samples were collected at a predefined frequency (e.g. post-treatment on Day 1; Table 7) from three depth strata: (i) the surface (0.1 m below the surface); (ii) 10 m below the surface; and (iii) the bottom (2 m above the seabed). Samples were collected at an array of sites including a subset of treated pen bays (designated in the EPA schedule), the lease centroid, external 35 m compliance sites and reference sites (Figure 2; Table 7).

Sampling for the T1 treatment shifted from the initial sampling design to the weekly sampling design, outlined below, after the Day 1 sampling event. Sites sampled in the initial sampling design carried through to the weekly sampling design.

Figure 6 illustrates water sampling under the initial sampling design and the relationship with each treatment event. Note that the Day 1 sampling event was conducted during the T2 treatment (Figure 6).

4.1.2 Weekly sampling design

For sampling associated with treatments T1 and T2, water samples were collected weekly from the three depth strata at internal (pen bay and centroid), external (35 m compliance site) and reference sites (Table 7). Samples were taken from the same 50% of treated pen bays as those sampled for sediment.

All internal, external and reference sites were sampled weekly until the treatment on the lease ended and until two consecutive results below LoR were returned for each internal (lease centroid, and pen bays), external and reference site, respectively.

Figure 6 illustrates the water sampling regime under the weekly sampling design and the relationship with each treatment event. The Week 1 survey was conducted on the final day of the T2 treatment, with all remaining weekly surveys occurring post-treatment (Figure 6).

4.2 Sample collection, processing and storage

Samples were collected in labelled 250 mL amber glass bottles provided by AST, kept cool in a dark environment and frozen as soon as possible after collection (unless they were delivered to the

laboratory within 12 hours of collection). Water samples were taken from the sites furthest from the lease boundary first (i.e. reference sites, external 35 m compliance sites and then internal sites) to minimise contamination risk. Water samples were delivered to AST, within 24 hours where possible. When this was not possible, samples were frozen for preservation and delivered promptly thereafter.

Samples were tested for both florfenicol (FFC) and florfenicol-amine (FFA) residues.

Table 7: Water sampling design and frequency associated with treatment events at MF94 The Sheppards. Post treatment sampling under the initial sampling design included predefined surveys (e.g. Day 1). Sampling was not required for cells highlighted in blue.

Location	Site	Initial Design	Weekly Design
		T1	T1+T2
Internal (Pen Bay)	TS_PB14	Day 1	Weekly
	TS_PB19	Day 1	Weekly
	TS_PB01		Weekly
	TS_PB04		Weekly
	TS_PB05		Weekly
	TS_PB07		Weekly
	TS_PB09		Weekly
	TS_PB11		Weekly
	TS_PB15		Weekly
	TS_PB20		Weekly
	TS_PB22		Weekly
	TS_PB23		Weekly
	TS_PB26		Weekly
Internal (Centroid)	TS_Centroid	Day 1	Weekly
External (35 m)	TS_CP1.1_N35	Day 1	Weekly
	TS_CP3.1_E35	Day 1	Weekly
	TS_CP2.2_S35	Day 1	Weekly
Reference	BEMP_M2	Day 1	Weekly
	BEMP_M3	Day 1	Weekly

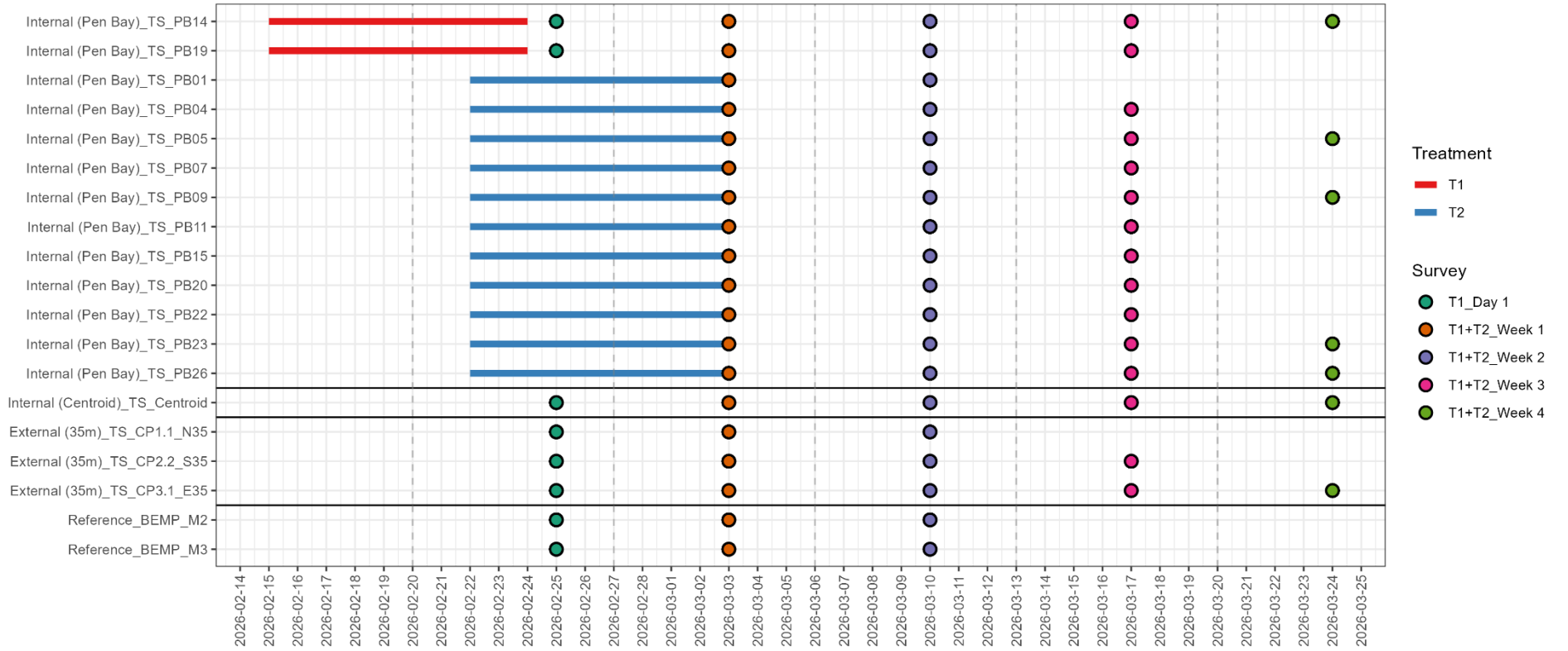

Figure 6: Sampling events for water surveys associated with the initial design (treatment T1; Day 1) and weekly design (treatments T1 and T2; weekly sampling). Sites sampled included internal pen bays, the lease centroid, external 35 m compliance sites and reference sites. Horizontal bars illustrate the timing and duration of treatment events (see Table 1; Figure 1).

4.3 Results

4.3.1 Initial sampling design

During T1 post-treatment surveys on Day 1, florfenicol (FFC) and/or florfenicol amine (FFA) concentrations were below the LoR for 62% of surface, middle and bottom water samples analysed (30 of 48 samples; Table 8; Table 9). There were eighteen samples above or equal to the LoR, which were collected from internal sites (i.e. the two pen bay sites and lease centroid) and two external 35 m compliance sites (Table 8; Table 9). It should be noted that while this was Day 1 post-treatment for pen bays associated treatment T1, treatment T2 was underway on the lease at this time (Figure 6). After Day 1, sampling for the T1 treatment shifted from the initial sampling design to the weekly sampling design (see Section 4.3.2).

4.3.2 Weekly sampling design

During weekly sampling associated with treatments T1 and T2, florfenicol (FFC) and/or florfenicol amine (FFA) concentrations were below the LoR for 75% of water samples analysed across surface, middle and bottom depths (270 of 360 samples; Table 8; Table 9). The ninety samples above or equal to the LoR were collected from internal sites (i.e. pen bays and lease centroid) and external 35 m compliance sites (Table 8; Table 9), with eighty samples collected in Week 1 (i.e. on the final day of treatment; Figure 6) and ten samples collected in Week 2 (i.e. seven days post-treatment; Figure 6; all FFA and equal to the LoR; Table 9). All samples collected from pen bay and external and reference sites in the following Week 3 and Week 4 surveys recorded FFC and/or FFA concentrations below the LoR (Table 8; Table 9). At reference sites, FFC and FFA concentrations were below the LoR during all weekly sampling events (Week 1 – Week 2; Table 8; Table 9).

4.3.3 Patterns of FFC and FFA across depths

Records of florfenicol (FFC) and/or florfenicol amine (FFA) concentrations above the LoR were distributed relatively evenly throughout the water column (surface = 38%, middle = 35%, bottom = 27%; Table 8; Table 9).

Table 8: Florfenicol (FFC) concentration (µg/L) in surface (S), middle (M) and bottom (B) water samples collected for initial sampling design (treatment T1; post-treatment Day 1) and weekly sampling design (treatments T1 and T2; weekly samples) at internal, external and reference sites at MF94 The Sheppards. Green shaded cells indicate sites where the sample was below the limit of reporting (LoR, i.e. <0.1 µg/L). The LoR is the lowest concentration of a substance that the laboratory can reliably and consistently measure. Blue shaded cells indicate sites where sampling and analysis was not required (refer to Table 7 for details). Unshaded cells indicate the sample result was above or equal to the LoR for that sample.

Location	Site	Florfenicol (µg/L)														
		T1			T1+T2											
		Day 1			Week 1			Week 2			Week 3			Week 4		
		2026-02-25			2026-03-03			2026-03-10			2026-03-17			2026-03-24		
		S	M	B	S	M	B	S	M	B	S	M	B	S	M	B
Internal (Pen Bay)	TS_PB01				<0.1	<0.1	<0.1	<0.1	<0.1	<0.1						
	TS_PB04				0.2	0.3	0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_PB05				0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
	TS_PB07				<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_PB09				<0.1	0.2	0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
	TS_PB11				0.8	0.4	0.5	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_PB14	<0.1	<0.1	<0.1	0.3	0.3	0.3	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
	TS_PB15				0.9	0.5	0.5	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_PB19	0.6	<0.1	0.2	0.2	0.1	0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_PB20				0.3	0.1	0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_PB22				0.4	0.4	0.5	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_PB23				0.2	0.3	0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
	TS_PB26				0.3	0.6	0.4	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
Internal (Centroid)	TS_Centroid	0.3	0.2	<0.1	0.2	0.2	0.3	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
External (35 m)	TS_CP1.1_N35	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1						
	TS_CP2.2_S35	0.2	0.2	<0.1	0.2	0.1	0.3	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_CP3.1_E35	<0.1	<0.1	<0.1	0.4	0.4	0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
Reference	BEMP_M2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1						
	BEMP_M3	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1						

Table 9: Florfenicol amine (FFA) concentration (µg/L) in surface (S), middle (M) and bottom (B) water samples collected for initial sampling design (treatment T1; post-treatment Day 1) and weekly sampling design (treatments T1 and T2; weekly samples) at internal, external and reference sites at MF94 The Sheppards. Green shaded cells indicate sites where the sample was below the limit of reporting (LoR, i.e. <0.1 µg/L). The LoR is the lowest concentration of a substance that the laboratory can reliably and consistently measure. Blue shaded cells indicate sites where sampling and analysis was not required (refer to Table 7 for details). Unshaded cells indicate the sample result was above or equal to the LoR for that sample.

Location	Site	Florfenicol amine (µg/L)														
		T1			T1+T2											
		Day 1			Week 1			Week 2			Week 3			Week 4		
		2026-02-25			2026-03-03			2026-03-10			2026-03-17			2026-03-24		
		S	M	B	S	M	B	S	M	B	S	M	B	S	M	B
Internal (Pen Bay)	TS_PB01				<0.1	<0.1	<0.1	<0.1	<0.1	<0.1						
	TS_PB04				0.4	0.4	0.4	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_PB05				0.4	<0.1	<0.1	0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
	TS_PB07				<0.1	0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_PB09				0.2	0.4	0.4	<0.1	0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
	TS_PB11				1.4	0.7	0.7	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_PB14	0.1	0.2	<0.1	0.6	0.6	0.6	0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
	TS_PB15				0.8	0.8	0.7	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_PB19	1.2	0.3	0.7	0.2	0.2	0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_PB20				0.5	0.2	0.3	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_PB22				0.8	0.6	0.8	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_PB23				0.4	0.6	0.5	0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
	TS_PB26				0.8	1.1	0.7	0.1	0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
Internal (Centroid)	TS_Centroid	0.3	0.9	0.3	0.4	0.4	0.4	0.1	0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
External (35 m)	TS_CP1.1_N35	0.9	0.4	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1						
	TS_CP2.2_S35	0.5	0.6	<0.1	0.5	0.3	0.3	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1			
	TS_CP3.1_E35	<0.1	<0.1	<0.1	0.9	0.7	0.4	0.1	0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
Reference	BEMP_M2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1						
	BEMP_M3	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1						

5 Wild Fish Sampling

5.1 Survey sites and Frequency

All wild fish sampling was undertaken by Marine Solutions at an array of sites within zones defined by buffers at increasing distance from the lease boundary (Figure 7; Appendix 2) at a frequency defined by the initial and weekly sampling designs (Table 10).

5.1.1 Initial sampling design

Wild fish samples associated with treatment T1 were collected from external zones (i.e. <1 km, 1-3 km) and specified reference sites (i.e. >3 km) post-treatment on Day 1 (Figure 7; Table 10). Note that the <1 km zone included the area inside and outside the lease (Figure 7; Appendix 2, Table A2.2). Twelve fish were collected from each site. Samples were submitted as individual fish and were composed of a mix of benthic and pelagic species. Note that the Day 1 sampling event on 25/02/2026 was conducted during the T2 treatment (22/02/2026 to 03/03/2026; Table 2).

5.1.2 Weekly sampling design

For surveys associated with treatments T1 and T2, samples from twelve wild fish were collected weekly from sites located within the same zones (i.e. <1 km, 1-3 km) and reference sites (i.e. >3 km) defined for the initial sampling (Figure 7; Table 10). For some sampling events, sufficient catch at prescribed BEMP reference sites could not be achieved due to low fish numbers. Under these circumstances fish sampling was instead conducted at one or two alternative sites that were equivalent in distance to the lease but had more fish present due to more favourable benthic habitat. Samples from the weekly sampling design were submitted as individual fish and were composed of a mix of benthic and pelagic species. Despite significant effort, only 11 fish were caught from the BEMP_M2 reference site on Week 1 due to low catch rates following a ban on capturing sand flathead by the Department of Natural Resources and Environment Tasmania (NRE Tas) after March 1 2026.

Importantly, the BEMP_M3 reference site for MF94 The Sheppards (3380 m from the lease, i.e. >3 km; Figure 3) was in relatively close proximity to the MF142 Roberts Point lease (1240 m from the lease; i.e. <3 km; Figure 3) which underwent treatment with florfenicol for three pen bays during the same period as the T2 treatment at MF94 The Sheppards (i.e. 22/02/2026-03/03/2026; Figure 1). The TRMP schedule for MF142 Roberts Point included a reference to BEMP_M3 in the monitoring requirements for the 1-3 km zone (Figure 3; Figure 7) and noted that samples collected at this site provide "*an additional 12 secondary fish samples within this zone*". The alternative sites for BEMP_M3 used in weekly surveys were located at Little Oyster Cove and Apollo Bay and were also within the 1-3 km zone for MF142 Roberts Point (see faded buffers in Figure 7). Results of the wild fish survey from samples collected BEMP_M3 are therefore discussed in relation to both leases.

Subject to prior approval from the EPA Director, all wild fish samples from an individual site or zone were sampled weekly until the treatment on the lease had ended and until two consecutive results below the LoR were returned for all samples within that site or zone. Based on this requirement, the EPA approved the discontinuation of wild fish sampling and analysis for the 1-3 km zone after Week 2, the <1 km zone and BEMP_M2 after Week 3, and BEMP_M3 after Week 7.

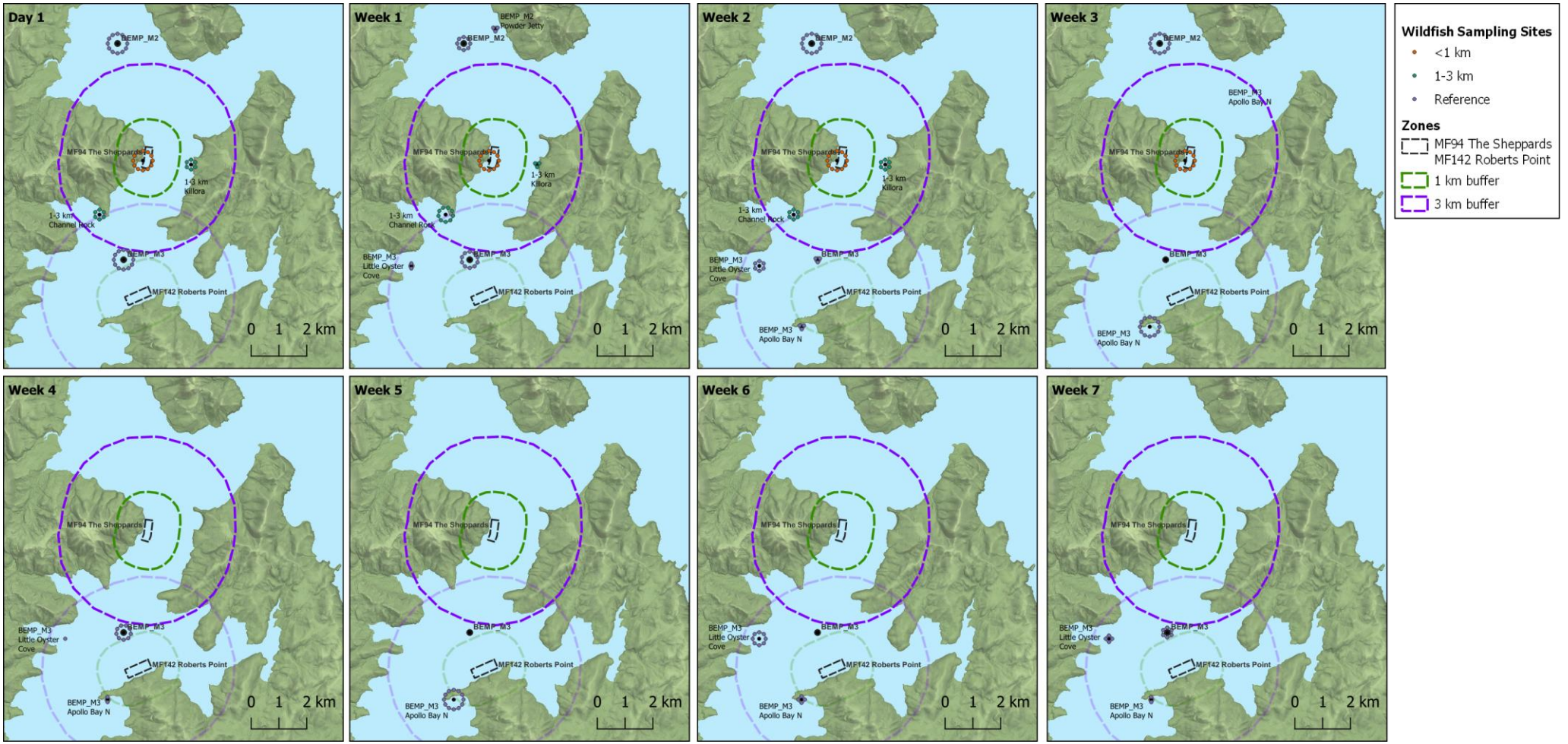

Figure 7: Location of sites where wild fish were collected within the <1 km (including area inside and outside the lease), 1-3 km zones and reference sites for the initial survey design (Day 1) and the weekly survey design (Week 1-Week 7). Twelve fish were collected from each zone in each survey. Overlapping samples were offset to concentric rings around a central point where the sample was taken. Coordinates are provided in Appendix 2; Table A2.2. Wild fish sampling details are provided in Appendix 5. Faded buffer zones around MF142 Roberts Point illustrate the location of BEMP_M3 in the 1-3 km zone for this lease which was treated concurrently to MF94 The Sheppards.

Table 10: Wild fish sampling sites and frequency associated with the initial design (treatment T1) and weekly design (treatments T1 and T2) at MF94 The Sheppards. Location of sites in their respective zones is illustrated in Figure 7. Coordinates and distance from lease are provided in Appendix 2; Table A2.2.

Treatment	Category	Zone	Site(s)	Fish Count	Frequency
T1	External	<1 km	<1 km	12	Day 1
		1-3 km	1-3 km Channel Rock 1-3 km Killora	12	Day 1
	Reference	>3 km	BEMP_M2	12	Day 1
		>3 km	BEMP_M3	12	Day 1
T1+T2	External	<1 km	<1 km	12	Weekly
		1-3 km	1-3 km Channel Rock 1-3 km Killora	12	Weekly
	Reference	>3 km	BEMP_M2 BEMP_M2 Powder Jetty	12	Weekly
		>3 km	BEMP_M3 BEMP_M3 Apollo Bay N BEMP_M3_Little Oyster Cove	12	Weekly

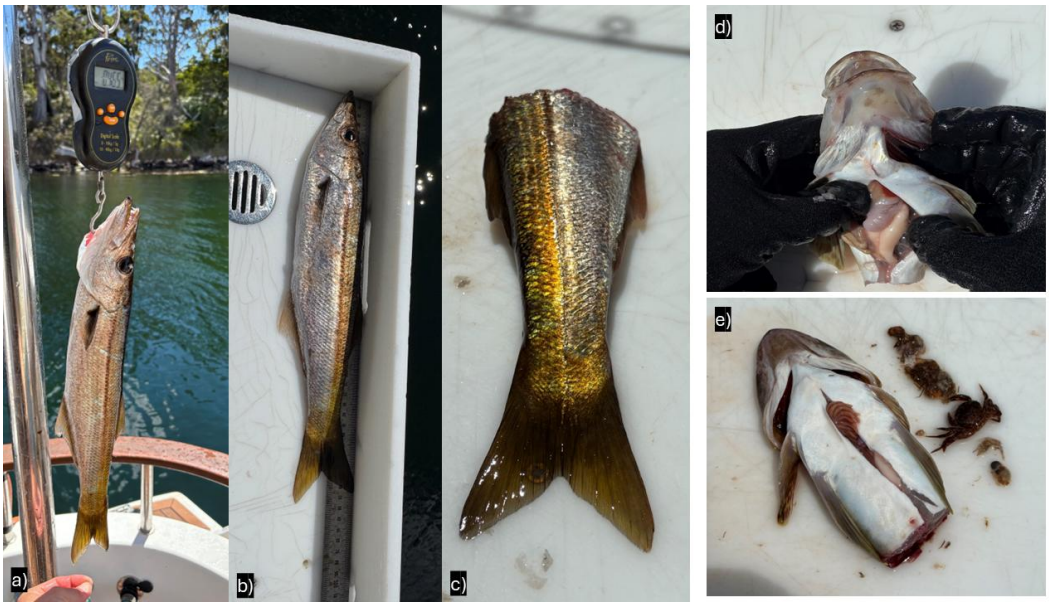

Figure 8: Representative photos taken during the processing of fish, showing (a) weight, (b) measurement, (c) tail sample for laboratory analysis, (d) gut processing and (e) gut content analysis.

5.2 Sample collection, processing and storage

Wild fish samples were collected first from reference sites (i.e. >3 km) before moving progressively to the zones closer to the lease. This method was used to manage contamination risk between samples closest to the pens treated with florfenicol. Ideally, the fish collected within each zone were to consist of 50% pelagic and 50% benthic species, but due to practical constraints there were instances where only one category of fish could be captured in a zone. Note that sand flathead could not be captured as part of wild fish sampling after March 1 2026, due to the catch ban implemented by the Department of Natural Resources and Environment Tasmania (NRE Tas).

GPS points were recorded at locations where fish were sampled (Appendix 2; Table A2.2). Once all fish for a location had been collected and the zones were complete, the full set of samples were processed.

Processing the samples involved collecting information on fish length, weight, and gut contents (Figure 8). Length was recorded using the longest measurable point, and weight was recorded using scales. A designated knife was then used to remove the tail posterior to the anus, which was retained for laboratory flesh analysis. A separate knife was used to dissect the fish to examine gut contents, and all observations were recorded. Between each dissection, all equipment used was thoroughly cleaned, and gloves were worn throughout the process. Flesh samples were bagged, labelled, and frozen for laboratory analysis. The lab analysis tested the sum of florfenicol, and its metabolites measured as florfenicol amine in the flesh tissue of each fish.

Note that results for wild fish flesh analysis were analysed and reported by AST as combined ‘florfenicol & metabolites as florfenicol amine’. This differed from the water and sediment samples which reported florfenicol and florfenicol amine components separately.

5.3 Results

Wild fish collected as part of the initial sampling design and weekly sampling design included fourteen benthic and pelagic species of which blue mackerel, blue-throat wrasse and Australian salmon were the species most commonly captured (Appendix 4). Details of fish caught and their gut contents are provided in Appendix 5.

5.3.1 Initial sampling design

For wild fish surveys conducted on Day 1 in association with the T1 treatment, florfenicol & metabolites as florfenicol amine (FFC/FFA) for wild fish samples were below the LoR for the majority of samples collected (45 of 48 samples; Table 11). Of the three fish samples with FFC/FFA concentrations above or equal to the LoR, one was collected from the <1 km zone and two were collected from reference site BEMP_M2 (Table 11; Figure 9). The three wild fish samples above or equal to the LoR were from blue mackerel (Table 12). After Day 1, sampling for the T1 treatment shifted from the initial sampling design to the weekly sampling design (see below).

5.3.2 Weekly sampling design

For weekly wild fish surveys associated with the T1 and T2 treatments, florfenicol & metabolites as florfenicol amine (FFC/FFA) for wild fish samples were below the LoR for the majority of samples collected between Week 1 and Week 7 (166 of 179 samples; Table 11). Of the thirteen weekly samples above or equal to the LoR, eleven were collected in Week 1 (conducted on the final day of treatment) from the <1 km zone and reference sites BEMP_M2 and BEMP_M3 (Table 12; Figure 9). The remaining two samples above or equal to the LoR were collected in Week 3 (14 days post-treatment) and Week 5 (28 days post-treatment) from the reference site BEMP_M3 Apollo Bay (Table 12; Figure 9). While the frequency was very low, some samples above or equal to the LoR were recorded up to 3.8 km MF94 The Sheppards and 1.2 km from MF142 Roberts Point (Table 12; Figure 9). Most of the wild fish that recorded FFC/FFA above or equal to the LoR were blue mackerel (Table 12). Blue mackerel are a mobile pelagic species which may explain the presence of FFC/FFA residues in wild fish samples at more distant reference sites.

The results for BEMP_M3 reference sites should also be interpreted in relation to their relatively close proximity to MF142 Roberts Point (i.e. within 1-3 km zone; Figure 7) which underwent treatment with florfenicol concurrently with MF94 The Sheppards[3]. It is possible that samples with FFC/FFA concentrations above or equal to the LoR in the five wild fish samples collected from the BEMP_M3 reference sites may have been influenced by florfenicol from the MF142 lease treatment (Table 12; Figure 9).

[3] Aquenal (2026) MF145 Roberts Point Therapeutant Residue Monitoring Program. Final Report. June 2026. Report to Tassal Operations Pty Ltd.

Table 11: Florfenicol & metabolites as florfenicol amine (FFC/FFA; mg/kg WMB) in wild fish tissue for wild fish caught from external zones (i.e. <1 km, 1-3 km) and specified reference sites (i.e. >3 km) as part of the initial sampling design (treatment T1) and weekly sampling design (T1 and T2) at MF94 The Sheppards. Green shaded cells indicate sites where the sample was below the limit of reporting (LoR, i.e. <0.01 mg/kg WMB). The LoR is the lowest concentration of a substance that the laboratory can reliably and consistently measure. Blue shaded cells indicate sites where sampling and analysis was not required (refer to Table 7 for details). Unshaded cells indicate the sample result was above or equal to the LoR for that sample.

Zone	Fish ID	Florfenicol & metabolites as Florfenicol Amine (mg/kg WMB)							
		T1	T1+T2						
		Day 1	Week 1	Week 2	Week 3	Week 4	Week 5	Week 6	Week 7
		2026-02-25	2026-03-03 2026-03-04 2026-03-05	2026-03-10 2026-03-11 2026-03-12	2026-03-17	2026-03-24 2026-03-25 2026-03-26	2026-04-01 2026-04-02	2026-04-08	2026-04-15
<1 km	Fish 01	2.1	<0.01	<0.01	<0.01				
	Fish 02	<0.01	<0.01	<0.01	<0.01				
	Fish 03	<0.01	<0.01	<0.01	<0.01				
	Fish 04	<0.01	<0.01	<0.01	<0.01				
	Fish 05	<0.01	<0.01	<0.01	<0.01				
	Fish 06	<0.01	<0.01	<0.01	<0.01				
	Fish 07	<0.01	0.04	<0.01	<0.01				
	Fish 08	<0.01	0.07	<0.01	<0.01				
	Fish 09	<0.01	<0.01	<0.01	<0.01				
	Fish 10	<0.01	0.17	<0.01	<0.01				
	Fish 11	<0.01	0.11	<0.01	<0.01				
	Fish 12	<0.01	0.89	<0.01	<0.01				
1-3 km	Fish 01	<0.01	<0.01	<0.01					
	Fish 02	<0.01	<0.01	<0.01					
	Fish 03	<0.01	<0.01	<0.01					
	Fish 04	<0.01	<0.01	<0.01					
	Fish 05	<0.01	<0.01	<0.01					
	Fish 06	<0.01	<0.01	<0.01					
	Fish 07	<0.01	<0.01	<0.01					
	Fish 08	<0.01	<0.01	<0.01					
	Fish 09	<0.01	<0.01	<0.01					
	Fish 10	<0.01	<0.01	<0.01					
	Fish 11	<0.01	<0.01	<0.01					
	Fish 12	<0.01	<0.01	<0.01					
BEMP_M2 (>3 km)	Fish 01	<0.01	<0.01	<0.01	<0.01				
	Fish 02	<0.01	0.04	<0.01	<0.01				
	Fish 03	<0.01	<0.01	<0.01	<0.01				
	Fish 04	<0.01	0.04	<0.01	<0.01				
	Fish 05	<0.01	0.01	<0.01	<0.01				
	Fish 06	<0.01	<0.01	<0.01	<0.01				
	Fish 07	<0.01	<0.01	<0.01	<0.01				
	Fish 08	0.01	<0.01	<0.01	<0.01				
	Fish 09	<0.01	<0.01	<0.01	<0.01				
	Fish 10	<0.01	<0.01	<0.01	<0.01				
	Fish 11	0.45	<0.01	<0.01	<0.01				
	Fish 12	<0.01	No Sample	<0.01	<0.01				
BEMP_M3[#] (>3 km)	Fish 01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
	Fish 02	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
	Fish 03	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
	Fish 04	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
	Fish 05	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
	Fish 06	<0.01	<0.01	<0.01	<0.01	<0.01	0.02	<0.01	<0.01
	Fish 07	<0.01	<0.01	<0.01	0.01	<0.01	<0.01	<0.01	<0.01
	Fish 08	<0.01	0.03	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
	Fish 09	<0.01	0.03	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
	Fish 10	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
	Fish 11	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
	Fish 12	<0.01	0.02	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01

[#] *Reference sites sampled for BEMP_M3 were > 3 km from MF94 The Sheppards lease but approximately 1 km from the MF142 Roberts Point lease, which underwent FFC treatment at the same time.*

Table 12: Description of fish species and location of sites (see Figure 7) for samples where florfenicol & metabolites as florfenicol amine (mg/kg WMB) in wild fish tissue was above the Limit of Reporting (see Table 11). Straight line distance from monitoring sites to the lease boundary was calculated for MF94 The Sheppards and MF142 Roberts Point.

Survey	Date	Zone	Site	Fish ID	Fish species	Easting	Northing	Distance from MF94 The Sheppards (m)	Distance from MF142 Roberts Point (m)
Day 1	25/02/2026	<1 km	<1 km	Fish 01	Blue mackerel	524447	5228827	40	4540
	25/02/2026	Reference	BEMP_M2	Fish 08	Blue mackerel	523535	5233037	3850	8810
	25/02/2026	Reference	BEMP_M2	Fish 11	Blue mackerel	523535	5233037	3850	8810
Week 1	3/03/2026	<1 km	<1 km	Fish 07	Blue mackerel	524447	5228827	40	4540
	3/03/2026	<1 km	<1 km	Fish 08	Blue mackerel	524447	5228827	40	4540
	3/03/2026	<1 km	<1 km	Fish 10	Blue mackerel	524447	5228827	40	4540
	3/03/2026	<1 km	<1 km	Fish 11	Blue mackerel	524447	5228827	40	4540
	3/03/2026	<1 km	<1 km	Fish 12	Blue mackerel	524447	5228827	40	4540
	3/03/2026	Reference	BEMP_M2	Fish 02	Snapper	523535	5233037	3850	8810
	3/03/2026	Reference	BEMP_M2	Fish 04	Blue mackerel	523535	5233037	3850	8810
	3/03/2026	Reference	BEMP_M2	Fish 05	Blue mackerel	523535	5233037	3850	8810
	3/03/2026	Reference	BEMP_M3	Fish 08	Blue mackerel	523752	5225263	3380	1240
	3/03/2026	Reference	BEMP_M3	Fish 09	Blue mackerel	523752	5225263	3380	1240
	3/03/2026	Reference	BEMP_M3	Fish 12	Blue mackerel	523752	5225263	3380	1240
Week 3	17/03/2026	Reference	BEMP_M3 Apollo Bay N	Fish 07	Bluethroat wrasse	523177	5222852	5860	1080
Week 5	1/04/2026	Reference	BEMP_M3 Apollo Bay N	Fish 06	Blue mackerel	523177	5222852	5860	1080

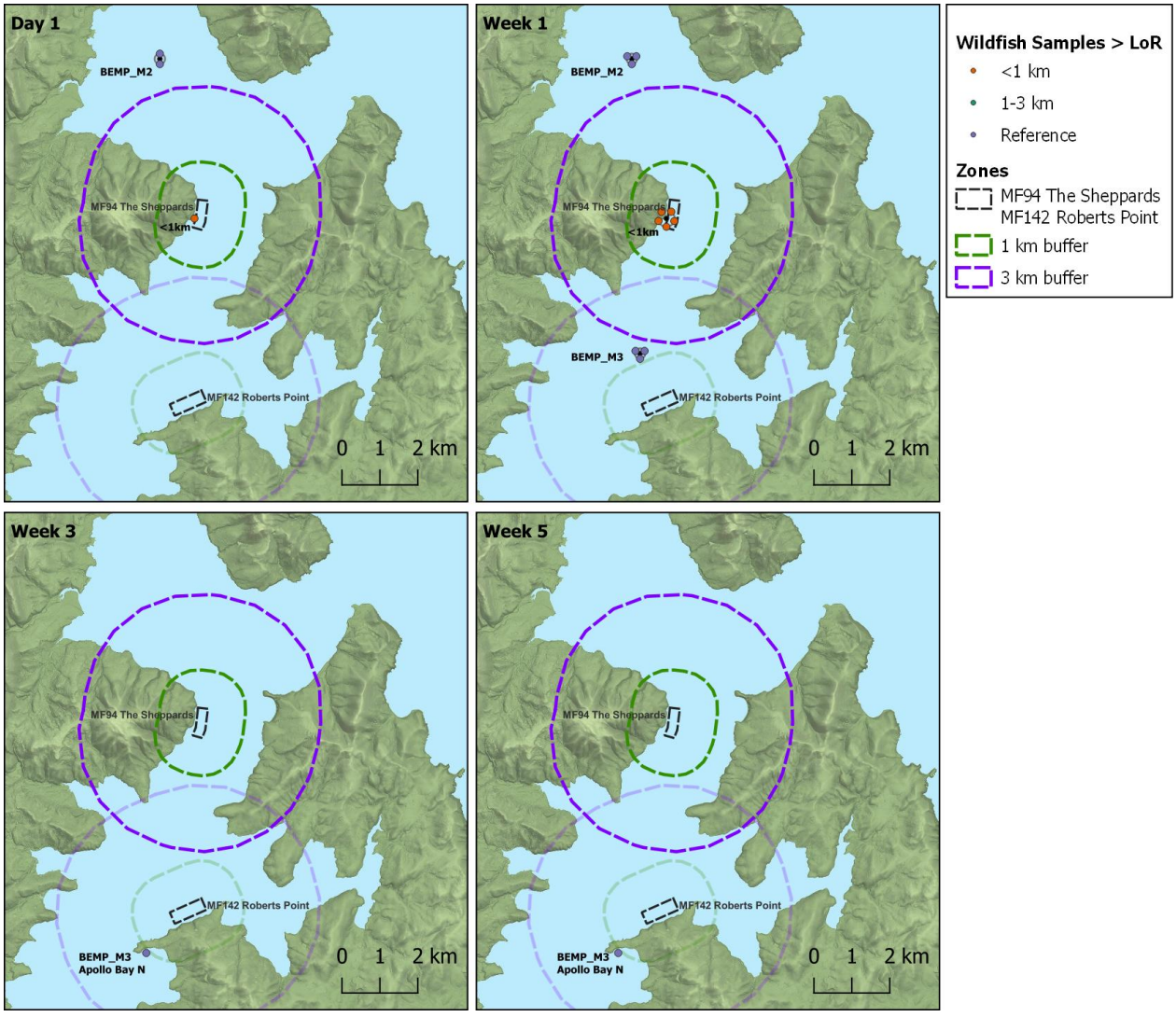

Figure 9: Location of sites where wild fish samples recorded florfenicol & metabolites as florfenicol amine (FFC/FFA) concentrations above the Limit of Reporting (LoR). Note that for display purposes, overlapping samples were offset slightly to concentric rings around a central point where the sample was taken. Faded buffer zones around MF142 Roberts Point illustrate the location of BEMP_M3 in the 1-3 km zone for this lease which was treated concurrently to MF94 The Sheppards.

6 Benthic Video Surveys

6.1 Survey sites and frequency

Benthic video surveys and interpretation of footage was undertaken as follows:

- Pre-treatment and post-treatment surveys of all pen bays treated as part of treatment T1 (Figure 2; Table A2.3 in Appendix 2).
- Pre-treatment and post-treatment surveys of all pen bays treated as part of treatment T2 (Figure 2; Table A2.3 in Appendix 2).

6.2 Sample collection, processing and storage

The pre-treatment and post-treatment benthic video surveys for treatment T1 and T2 were conducted by Fulcrum robotics. Fulcrum's surveys were carried out using a Deep Trekker Revolution® vectored inspection class Remotely Operated Vehicle (ROV). The ROV has an internal 4K colour video camera that is relayed to the topside via a tether. HD recording was also captured using a GoPro® Hero5 with the settings medium view, NTSC 2.7K, 60FPS.

All video footage from the two treatments was reviewed and databased by Fulcrum, using the same methodology as specified in the Environmental Licence. Survey coordinates for the benthic video survey are provided in Appendix 2.

6.3 Results

A description of seabed conditions for the pre-treatment and post-treatment surveys for each pen bay is provided in Appendix 6. The benthic habitats of pen bay sites at The Sheppards (MF94) were typically characterised by silty or sandy sediments, typically with shells and shell fragments present. There was no sign of gas bubbles emanating from sediments under pen bays. Patchy *Beggiatoa* sp. were observed at the majority of pens bays during pre-treatment and post-treatment surveys. Capitellid worms (many or few) were observed in seven of the forty-eight surveys, mainly in the pre-treatment surveys. Pellets (mostly 'few') and faeces (mostly 'many') were observed under the majority of pen bays during pre-treatment and post-treatment surveys. A variable flora consisting mainly of drift algae was recorded under most pen bays. A range of fauna was observed, with anemones, ascidians, seastars, side-gilled slugs, and a range of crustaceans most commonly recorded. Overall, benthic condition was consistent between pre-treatment and post-treatment surveys.

7 Summary

A Therapeutant Residue Monitoring Program (TRMP) was undertaken in the vicinity of MF94 The Sheppards, located in the D'Entrecasteaux Channel and Huon River Marine Farming Development Plan area. The TRMP was conducted in accordance with the specifications stipulated by the Tasmanian EPA in *Therapeutant Residue Monitoring Schedule – Marine Farming Lease No. 94,* issued 20 February 2026 and *Addendum - Therapeutant Residue Monitoring Schedule – MF94*, issued 27 February 2026.

The TRMP included a suite of monitoring components (sediment, water, wild fish and benthic video surveys). Water, sediment and benthic video surveys sampled an array of monitoring sites including internal sites (i.e. pen bays and lease centroid), external sites (i.e. compliance sites and transects) and reference sites. Wild fish were sampled from zones defined by buffers at increasing distance from the lease boundary (i.e. <1 km, 1-3 km) and two reference sites. The frequency of monitoring changed between the initial sampling schedule (TRM Schedule MF94; e.g. post-treatment on Day 1) and the subsequent addendum which required weekly sampling (TRM Schedule MF94 Addendum).

For sediment samples, florfenicol (FFC) and/or florfenicol amine (FFA) concentrations were below the limit of reporting (LoR) for the overwhelming majority (>95%) of samples taken on Day 1 after the T1 treatment and during weekly surveys associated with treatments T1 and T2. All seven samples with FFC/FFA above or equal to the LoR were from treated pen bays with the majority collected within two days post-treatment. All sediment samples collected from external and reference sites recorded FFC and FFA concentrations below the LoR.

For water samples, florfenicol (FFC) and/or florfenicol amine (FFA) concentrations were below the LoR for the majority (74%) of samples collected under the initial and weekly sampling designs. Samples with FFC and/or FFA concentrations above or equal to the LoR included internal (pen bay and lease centroid) and external compliance sites (but not reference sites) and the majority were collected on the final day of treatment or one day post-treatment.

For wild fish samples, florfenicol & metabolites as florfenicol amine (FFC/FFA) concentrations were below the LoR for the majority of samples (93%) associated with Day 1 and weekly surveys for treatments T1 and T2. Wild fish samples with FFC/FFA concentrations above or equal to the LoR were collected from the <1 km zone and at reference sites. While the frequency was low, in some cases samples with concentrations above the LoR were recorded at reference sites 1.2-3.8 km from the nearest treated lease. Of the sixteen wild fish with FFC/FFA concentrations above (or equal to) the LoR, fourteen were collected on the final day of treatment or one day post-treatment. Instances of FFC/FFA above the LoR in wild fish samples collected from BEMP_M3 reference sites may also have been influenced by concurrent treatment events applied to the nearby MF142 Roberts Point lease.

Appendices

Appendix 1: Antibiotic Florfenicol treatment details for MF94 The Sheppards lease. Treatment was administered by top coated feed, as prescribed by the Veterinary Authority (VA). Information supplied by Tassal Operations Pty Ltd.

Type	VA	Lease	Pen Bays	Days	Treatment start date	Actual quantity of active ingredient (kg) used
Florfenicol	26-Tx-M04	94	TS_PB14, TS_PB16, TS_PB19	10	15/02/2026	77
Florfenicol	26-Tx-M08	94	TS_PB01, TS_PB02, TS_PB03, TS_PB04, TS_PB05, TS_PB06, TS_PB07, TS_PB08, TS_PB09, TS_PB10, TS_PB11, TS_PB12, TS_PB15, TS_PB17, TS_PB18, TS_PB20, TS_PB21, TS_PB22, TS_PB23, TS_PB24, TS_PB26	10	22/02/2026	496

In accordance with section 28(2(d)) of the *Environmental Standards for Tasmanian Marine Finfish Farming 2023*, the Therapeutant Residue Monitoring Program report must include an evaluation of the factors contributing to the need for medication, including (i) potential seasonal/environmental factors; (ii) history of therapeutant use and its cause on the lease area in the previous 10 year period; and (iii) Any known issues with batches of fish involved in the medication event, including vaccination status with reference to the disease required. A summary of these factors is provided below:

(i) The main triggering issue is likely to be extended periods of elevated water temperatures.

(ii) In the last 10 years antibiotics have been used once previously on the lease. Medicated feed containing antibiotics (active ingredient: Oxytetracycline) was applied in 2022 for the treatment of Vibriosis in Atlantic Salmon stock.

(iii) There were no known issues with batches of fish in the medication event. Waning immunity from vaccination likely coincided with the observed temperature rises

Appendix 2: Survey coordinates for the residue monitoring survey, based on the Mapping Grid of Australia Zone 55 (Datum GDA94). Coordinates from all surveys have been supplied to EPA in electronic format.

Table A2.1: Coordinates for sampling sites for water and sediment surveys. Coordinates are from the EPA schedule or from the first instance a site was visited in the field by Aquenal or Marine Solutions.

Location	Site	Easting	Northing
Internal (Pen Bay)	TS_PB14	524707	5228918
	TS_PB19	524588	5228755
	TS_PB01	524680	5229265
	TS_PB04	524755	5229183
	TS_PB05	524660	5229160
	TS_PB07	524646	5229098
	TS_PB09	524639	5229056
	TS_PB11	524626	5228994
	TS_PB15	524617	5228915
	TS_PB20	524656	5228740
	TS_PB22	524649	5228678
	TS_PB23	524583	5228679
	TS_PB26	524610	5228572
Internal (Centroid)	TS_Centroid	524620	5228929
External (35 m)	TS_CP1.1_N35	524651	5229347
	TS_CP3.1_E35	524790	5228910
	TS_CP2.2_S35	524601	5228499
External (100 m)	TS_CP1.1_N100	524658	5229412
	TS_CP3.1_E100	524854	5228902
	TS_CP2.2_S100	524584	5228438
External (500 m)	TS_CP1.1_N500	524702	5229809
	TS_CP3.1_E500	525251	5228853
	TS_CP2.2_S500	524477	5228053
Reference	BEMP_M2	523535	5233037
	BEMP_M3	523752	5225263

Table A2.2: Coordinates for sites sampled during each wild fish survey (Marine Solutions) and the straight line distance of monitoring sites to the lease boundary for MF94 The Sheppards and MF142 Roberts Point.

Survey	Zone	Location	Easting	Northing	Distance to MF94 The Sheppards (m)	Distance to MF142 Roberts Point (m)
Day 1	<1 km	<1 km	524447	5228827	40	4540
	1-3 km	1-3 km Channel Rock	522886	5226886	2280	3080
		1-3 km Killora	526180	5228683	1440	4660
	Reference	BEMP_M2	523535	5233037	3850	8810
		BEMP_M3	523752	5225263	3380	1240
Week 1	<1 km	<1 km	524447	5228827	40	4540
	1-3 km	1-3 km Channel Rock	522886	5226886	2280	3080
		1-3 km Killora	526180	5228683	1440	4660
	Reference	BEMP_M2	523535	5233037	3850	8810
		BEMP_M2	523535	5233037	3850	8810
		BEMP_M2 Powder Jetty	524675	5233561	4240	9270
		BEMP_M3	523752	5225263	3380	1240
		BEMP_M3 Little Oyster Cove	521653	5225040	4490	2420
Week 2	<1 km	<1 km	524447	5228827	40	4540
	1-3 km	1-3 km Channel Rock	522886	5226886	2280	3080
		1-3 km Killora	526180	5228683	1440	4660
	Reference	BEMP_M2	523535	5233037	3850	8810
		BEMP_M3	523752	5225263	3380	1240
		BEMP_M3 Apollo Bay N	523177	5222852	5860	1080
		BEMP_M3 Little Oyster Cove	521653	5225040	4490	2420
Week 3	<1 km	<1 km	524447	5228827	40	4540
	1-3 km	1-3 km Channel Rock	522886	5226886	2280	3080
		1-3 km Killora	526180	5228683	1440	4660
	Reference	BEMP_M2	523535	5233037	3850	8810
		BEMP_M3 Apollo Bay N	523177	5222852	5860	1080
Week 4	<1 km	<1 km	524447	5228827	40	4540
	1-3 km	1-3 km Channel Rock	522886	5226886	2280	3080
		1-3 km Killora	526180	5228683	1440	4660
	Reference	BEMP_M2	523535	5233037	3850	8810
		BEMP_M3	523752	5225263	3380	1240
		BEMP_M3 Apollo Bay N	523177	5222852	5860	1080
		BEMP_M3 Little Oyster Cove	521653	5225040	4490	2420
Week 5	<1 km	<1 km	524447	5228827	40	4540
	1-3 km	1-3 km Channel Rock	522886	5226886	2280	3080
	Reference	BEMP_M2	523535	5233037	3850	8810
		BEMP_M3 Apollo Bay N	523177	5222852	5860	1080
Week 6	<1 km	<1 km	524447	5228827	40	4540
	Reference	BEMP_M2	523535	5233037	3850	8810
		BEMP_M3 Apollo Bay N	523177	5222852	5860	1080
		BEMP_M3 Little Oyster Cove	521653	5225040	4490	2420
Week 7	<1 km	<1 km	524447	5228827	40	4540
	Reference	BEMP_M2	523535	5233037	3850	8810
		BEMP_M3	523752	5225263	3380	1240
		BEMP_M3 Apollo Bay N	523177	5222852	5860	1080
		BEMP_M3 Little Oyster Cove	521653	5225040	4490	2420

Table A2.3: GPS coordinates and depth for pre-treatment and post-treatment benthic video surveys conducted by Fulcrum Robotics

Treatment	Survey	Date	Site	Easting	Northing
T1	Pre-treatment	6/02/2026	TS_PB14	524711	5228927
		6/02/2026	TS_PB16	524691	5228893
		6/02/2026	TS_PB19	524589	5228756
	Post-treatment	25/02/2026	TS_PB14	524694	5228952
		25/02/2026	TS_PB16	524675	5228854
		25/02/2026	TS_PB19	524584	5228755
T2	Pre-treatment	21/02/2026	TS_PB01	524694	5229247
		21/02/2026	TS_PB02	524770	5229256
		21/02/2026	TS_PB03	524680	5229191
		21/02/2026	TS_PB04	524766	5229203
		21/02/2026	TS_PB05	524675	5229125
		21/02/2026	TS_PB06	524754	5229145
		21/02/2026	TS_PB07	524668	5229075
		21/02/2026	TS_PB08	524744	5229096
		21/02/2026	TS_PB09	524653	5229021
		21/02/2026	TS_PB10	524727	5229027
		21/02/2026	TS_PB11	524629	5228966
		21/02/2026	TS_PB12	524722	5228975
		21/02/2026	TS_PB15	524629	5228856
		21/02/2026	TS_PB17	524609	5228795
		21/02/2026	TS_PB18	524665	5228834
		21/02/2026	TS_PB20	524650	5228772
		21/02/2026	TS_PB21	524583	5228682
		21/02/2026	TS_PB22	524642	5228686
		21/02/2026	TS_PB23	524580	5228638
		21/02/2026	TS_PB24	524641	5228673
		21/02/2026	TS_PB26	524612	5228559
	Post-treatment	4/03/2026	TS_PB01	524693	5229246
		4/03/2026	TS_PB02	524769	5229256
		4/03/2026	TS_PB03	524680	5229190
		4/03/2026	TS_PB04	524765	5229203
		4/03/2026	TS_PB05	524675	5229124
		4/03/2026	TS_PB06	524753	5229144
		4/03/2026	TS_PB07	524668	5229074
		4/03/2026	TS_PB08	524743	5229095
		4/03/2026	TS_PB09	524652	5229020
		4/03/2026	TS_PB10	524727	5229026
		4/03/2026	TS_PB11	524628	5228966
		4/03/2026	TS_PB12	524721	5228974
		4/03/2026	TS_PB15	524628	5228856
		4/03/2026	TS_PB17	524609	5228795
		4/03/2026	TS_PB18	524665	5228834
		4/03/2026	TS_PB20	524650	5228772
		4/03/2026	TS_PB21	524583	5228682
		4/03/2026	TS_PB22	524641	5228685
		4/03/2026	TS_PB23	524579	5228638
		4/03/2026	TS_PB24	524640	5228672
		4/03/2026	TS_PB26	524612	5228558

Appendix 3: Weather conditions during field surveys.

Treatment	Sampling Event	Sampler	Field Date	Wind (knots, direction)	Sky
T1	**Day 1**	Marine Solutions	25/02/2026	10 knots S	Cloudy
		Aquenal	25/02/2026	5-10 knots SW	Cloudy
T1+T2	**Week 1**	Aquenal	4-5/03/2026	10-15 knots NW	Partly Cloudy
		Marine Solutions	3/03/2026	Light and variable	Sunny
		Marine Solutions	4/03/2026	5 knots S	Partly Cloudy
		Marine Solutions	5/03/2026	10 knots NW	Partly Cloudy
	Week 2	Aquenal	11-12/03/2026	10 knots W	Cloudy
		Marine Solutions	10/03/2026	Light and variable	Sunny
		Marine Solutions	11/03/2026	10 - 20 knots S	Cloudy
		Marine Solutions	13/03/2026	15 knots NW	Partly Cloudy
	Week 3	Marine Solutions	17/03/2026	5 - 15 knots SW	Partly Cloudy
		Aquenal	18/03/2026	5-10 knots NE	Sunny
	Week 4	Aquenal	23/03/2026	10 knots SW	Cloudy
		Marine Solutions	24/03/2026	10 knots SW	Cloudy
		Marine Solutions	25/03/2026	Light and variable	Sunny
		Marine Solutions	26/03/2026	10 - 15 knots SW	Cloudy
	Week 5	Marine Solutions	1/04/2026	5 - 10 knots NW	Cloudy
		Marine Solutions	2/04/2026	Light and variable	Partly Cloudy
	Week 6	Marine Solutions	8/04/2026	Light and variable	Partly Cloudy
	Week 7	Marine Solutions	15/04/2026	5-15 knots W	Cloudy

Appendix 4: Wild fish sampling – species collected during surveys.

Figure A4: Number of fish collected for each species caught during surveys conducted under the initial sampling design (treatment T1) and the weekly sampling design (treatments T1 and T2). Sampling of 1-3 km zone was discontinued after Week 2 and sampling of <1 km and BEMP_M2 sites were discontinued after Week 3.

Appendix 5: Wild fish sampling field data.

Table A5.1: Wild fish sample data, Initial sampling design: Day 1 survey (T1).

Site	Date	Fish No.	Species	Length (mm)	Weight (g)	Gut Content	Easting	Northing
<1 km	25/02/2026	1	Blue mackerel	270	195	Digested material	524447	5228827
<1 km	25/02/2026	2	Blue mackerel	270	195	Digested material	524447	5228827
<1 km	25/02/2026	3	Blue mackerel	280	220	Digested material	524447	5228827
<1 km	25/02/2026	4	Blue mackerel	230	95	Digested material	524447	5228827
<1 km	25/02/2026	5	Sand flathead	235	65	Empty	524447	5228827
<1 km	25/02/2026	6	Sand flathead	205	65	Amphipod	524447	5228827
<1 km	25/02/2026	7	Sand flathead	235	65	Empty	524447	5228827
<1 km	25/02/2026	8	Sand flathead	145	30	Empty	524447	5228827
<1 km	25/02/2026	9	Bluethroat wrasse	395	1120	Shell grit	524447	5228827
<1 km	25/02/2026	10	Bluethroat wrasse	300	525	Empty	524447	5228827
<1 km	25/02/2026	11	Bluethroat wrasse	315	595	Shell grit	524447	5228827
<1 km	25/02/2026	12	Bluethroat wrasse	360	870	Mussel + shells	524447	5228827
1-3 km Channel Rock	25/02/2026	1	Sand flathead	260	110	Empty	522886	5226886
1-3 km Channel Rock	25/02/2026	2	Sand flathead	265	105	Amphipod	522886	5226886
1-3 km Channel Rock	25/02/2026	3	Sand flathead	270	120	Brown algae	522886	5226886
1-3 km Channel Rock	25/02/2026	4	Sand flathead	225	80	Amphipod	522886	5226886
1-3 km Channel Rock	25/02/2026	5	Sand flathead	225	80	Empty	522886	5226886
1-3 km Channel Rock	25/02/2026	6	Bluethroat wrasse	330	620	Empty	522886	5226886
1-3 km Killora	25/02/2026	7	Sand flathead	305	185	Empty	526180	5228683
1-3 km Killora	25/02/2026	8	Sand flathead	260	125	Shell grit	526180	5228683
1-3 km Killora	25/02/2026	9	Sand flathead	255	120	Amphipod	526180	5228683
1-3 km Killora	25/02/2026	10	Sand flathead	220	75	Amphipod	526180	5228683
1-3 km Killora	25/02/2026	11	Sand flathead	225	75	Empty	526180	5228683
1-3 km Killora	25/02/2026	12	Sand flathead	240	75	Empty	526180	5228683
BEMP_M2	25/02/2026	1	Sand flathead	244	100	Empty	523535	5233037
BEMP_M2	25/02/2026	2	Sand flathead	230	70	Empty	523535	5233037
BEMP_M2	25/02/2026	3	Sand flathead	220	70	Empty	523535	5233037
BEMP_M2	25/02/2026	4	Sand flathead	237	85	Empty	523535	5233037
BEMP_M2	25/02/2026	5	Sand flathead	228	70	Empty	523535	5233037
BEMP_M2	25/02/2026	6	Sand flathead	195	45	Amphipod	523535	5233037
BEMP_M2	25/02/2026	7	Sand flathead	199	50	Empty	523535	5233037
BEMP_M2	25/02/2026	8	Blue mackerel	225	115	Digested material	523535	5233037
BEMP_M2	25/02/2026	9	Blue mackerel	206	80	Digested material	523535	5233037
BEMP_M2	25/02/2026	10	Blue mackerel	195	60	Digested material	523535	5233037
BEMP_M2	25/02/2026	11	Blue mackerel	272	200	Digested material	523535	5233037
BEMP_M2	25/02/2026	12	Blue mackerel	285	200	Digested material	523535	5233037
BEMP_M3	25/02/2026	1	Sand flathead	210	55	Empty	523752	5225263
BEMP_M3	25/02/2026	2	Sand flathead	215	55	Empty	523752	5225263
BEMP_M3	25/02/2026	3	Sand flathead	223	75	White bait	523752	5225263
BEMP_M3	25/02/2026	4	Sand flathead	255	120	White bait	523752	5225263
BEMP_M3	25/02/2026	5	Sand flathead	215	60	Philinid	523752	5225263
BEMP_M3	25/02/2026	6	Sand flathead	220	60	Empty	523752	5225263
BEMP_M3	25/02/2026	7	Sand flathead	145	30	Digested material	523752	5225263
BEMP_M3	25/02/2026	8	Sand flathead	150	25	Empty	523752	5225263
BEMP_M3	25/02/2026	9	Sand flathead	250	100	Empty	523752	5225263
BEMP_M3	25/02/2026	10	Sand flathead	140	30	Empty	523752	5225263
BEMP_M3	25/02/2026	11	Sand flathead	140	25	Empty	523752	5225263
BEMP_M3	25/02/2026	12	Sand flathead	150	30	Empty	523752	5225263

Table A5.2: Wild fish sample data: Week 1 (T1+T2).

Site	Date	Fish No.	Species	Length (mm)	Weight (g)	Gut Content	Easting	Northing
<1 km	3/03/2026	1	Bluethroat wrasse	250	530	Urchin, crab	524447	5228827
<1 km	3/03/2026	2	Bluethroat wrasse	345	580	Crab, shell grit	524447	5228827
<1 km	3/03/2026	3	Bluethroat wrasse	290	490	Shell grit	524447	5228827
<1 km	3/03/2026	4	Bluethroat wrasse	270	355	Crab, shell grit	524447	5228827
<1 km	3/03/2026	5	Pike	200	60	Fish	524447	5228827
<1 km	3/03/2026	6	Blue mackerel	310	350	Fish	524447	5228827
<1 km	3/03/2026	7	Blue mackerel	300	320	Unidentifiable digested material	524447	5228827
<1 km	3/03/2026	8	Blue mackerel	300	320	Unidentifiable digested material	524447	5228827
<1 km	3/03/2026	9	Blue mackerel	295	255	Empty	524447	5228827
<1 km	3/03/2026	10	Blue mackerel	290	280	Algae	524447	5228827
<1 km	3/03/2026	11	Blue mackerel	285	250	Unidentifiable digested material	524447	5228827
<1 km	3/03/2026	12	Blue mackerel	350	400	Unidentifiable digested material	524447	5228827
1-3 km Channel Rock	4/03/2026	1	Pike	225	75	Fish	522886	5226886
1-3 km Channel Rock	4/03/2026	2	Pike	240	85	Empty	522886	5226886
1-3 km Channel Rock	4/03/2026	3	Barber Perch	195	135	Unidentifiable digested material	522886	5226886
1-3 km Channel Rock	4/03/2026	4	Barber Perch	205	170	Empty	522886	5226886
1-3 km Channel Rock	4/03/2026	5	Barber Perch	195	160	Algae	522886	5226886
1-3 km Channel Rock	4/03/2026	6	Barber Perch	160	100	Empty	522886	5226886
1-3 km Channel Rock	4/03/2026	7	Barber Perch	160	100	Empty	522886	5226886
1-3 km Channel Rock	4/03/2026	8	Barber Perch	130	75	Empty	522886	5226886
1-3 km Killora	5/03/2026	9	Bluethroat wrasse	305	385	Shell grit	526180	5228683
1-3 km Killora	5/03/2026	10	Bluethroat wrasse	485	310	Empty	526180	5228683
1-3 km Killora	5/03/2026	11	Bluethroat wrasse	350	995	Shell grit, mussel shells	526180	5228683
1-3 km Channel Rock	5/03/2026	12	Barber Perch	158	85	Unidentifiable digested material	522886	5226886
BEMP_M2	3/03/2026	1	Blue mackerel	283	200	Empty	523535	5233037
BEMP_M2	3/03/2026	2	Snapper	220	185	Empty	523535	5233037
BEMP_M2	3/03/2026	3	Blue mackerel	285	225	Empty	523535	5233037
BEMP_M2	3/03/2026	4	Blue mackerel	287	220	Unidentifiable digested material	523535	5233037
BEMP_M2	3/03/2026	5	Blue mackerel	315	295	Empty	523535	5233037
BEMP_M2	3/03/2026	6	Blue mackerel	290	220	Unidentifiable digested material	523535	5233037
BEMP_M2	3/03/2026	7	Jack Mackerel	225	105	Unidentifiable digested material	523535	5233037
BEMP_M2	3/03/2026	8	Jack Mackerel	230	115	Empty	523535	5233037
BEMP_M2 Powder Jetty	3/03/2026	9	Bluethroat wrasse	220	185	Shell grit	524675	5233561
BEMP_M2 Powder Jetty	3/03/2026	10	Bluethroat wrasse	330	530	Shell grit, unidentifiable digested material	524675	5233561
BEMP_M2 Powder Jetty	3/03/2026	11	Bluethroat wrasse	210	178	Mussel shell	524675	5233561
BEMP_M3	4/03/2026	1	Australian Salmon	240	210	Unidentifiable digested material	523752	5225263
BEMP_M3 Little Oyster Cove	4/03/2026	2	Australian Salmon	300	375	Fish	521653	5225040
BEMP_M3	4/03/2026	3	Australian Salmon	260	225	Empty	523752	5225263
BEMP_M3 Little Oyster Cove	4/03/2026	4	Australian Salmon	295	340	Fish	521653	5225040
BEMP_M3	4/03/2026	5	Australian Salmon	280	300	Fish	523752	5225263
BEMP_M3	4/03/2026	6	Blue mackerel	275	305	Fish	523752	5225263
BEMP_M3	4/03/2026	7	Blue mackerel	320	380	Fish	523752	5225263
BEMP_M3	4/03/2026	8	Blue mackerel	305	285	Fish	523752	5225263
BEMP_M3	4/03/2026	9	Blue mackerel	330	285	Fish	523752	5225263
BEMP_M3	4/03/2026	10	Blue mackerel	320	290	Empty	523752	5225263
BEMP_M3	4/03/2026	11	Blue mackerel	295	270	Fish	523752	5225263
BEMP_M3	4/03/2026	12	Blue mackerel	305	330	Fish	523752	5225263

Table A5.3: Wild fish sample data: Week 2 (T1+T2).

Site	Date	Fish No.	Species	Length (mm)	Weight (g)	Gut Content	Easting	Northing
<1 km	10/03/2026	1	Bluethroat wrasse	325	570	Unidentifiable digested material	524447	5228827
<1 km	10/03/2026	2	Bluethroat wrasse	417	1400	Green algae and shell grit	524447	5228827
<1 km	10/03/2026	3	Bluethroat wrasse	280	355	Empty	524447	5228827
<1 km	10/03/2026	4	Bluethroat wrasse	310	500	Unidentifiable digested material	524447	5228827
<1 km	10/03/2026	5	Bluethroat wrasse	310	635	Unidentifiable digested material	524447	5228827
<1 km	10/03/2026	6	Bluethroat wrasse	278	400	Shell grit	524447	5228827
<1 km	10/03/2026	7	Bluethroat wrasse	273	320	Unidentifiable digested material	524447	5228827
<1 km	10/03/2026	8	Bluethroat wrasse	250	260	Shell grit	524447	5228827
<1 km	10/03/2026	9	Bluethroat wrasse	242	260	Shell grit	524447	5228827
<1 km	10/03/2026	10	Bluethroat wrasse	220	200	Shell grit	524447	5228827
<1 km	10/03/2026	11	Barber Perch	230	200	Algae	524447	5228827
<1 km	10/03/2026	12	Barber Perch	230	225	Empty	524447	5228827
1-3 km Killora	10/03/2026	1	Bluethroat wrasse	285	295	Empty	526180	5228683
1-3 km Killora	10/03/2026	2	Bluethroat wrasse	305	560	Empty	526180	5228683
1-3 km Killora	10/03/2026	3	Bluethroat wrasse	390	790	Empty	526180	5228683
1-3 km Killora	10/03/2026	4	Bluethroat wrasse	230	185	Shell grit	526180	5228683
1-3 km Killora	10/03/2026	5	Bluethroat wrasse	455	1720	Crab, algae	526180	5228683
1-3 km Killora	10/03/2026	6	Red Cod	309	370	Unidentifiable digested material	526180	5228683
1-3 km Channel Rock	10/03/2026	7	Snapper	234	185	Unidentifiable digested material	522886	5226886
1-3 km Channel Rock	10/03/2026	8	Barber Perch	223	155	Bait fish	522886	5226886
1-3 km Channel Rock	10/03/2026	9	Pike	235	75	Shell grit	522886	5226886
1-3 km Channel Rock	10/03/2026	10	Pike	207	55	Empty	522886	5226886
1-3 km Channel Rock	10/03/2026	11	Pike	227	75	Shell grit, mussel shells	522886	5226886
1-3 km Channel Rock	10/03/2026	12	Jackass Morwong	240	195	Unidentifiable digested material	522886	5226886
BEMP_M2	10/03/2026	1	Blue mackerel	205	250	Empty	523535	5233037
BEMP_M2	10/03/2026	2	Blue mackerel	220	55	Empty	523535	5233037
BEMP_M2	10/03/2026	3	Blue mackerel	220	85	Empty	523535	5233037
BEMP_M2	10/03/2026	4	Blue mackerel	195	65	Unidentifiable digested material	523535	5233037
BEMP_M2	10/03/2026	5	Blue mackerel	215	85	Empty	523535	5233037
BEMP_M2	10/03/2026	6	Blue mackerel	210	85	Unidentifiable digested material	523535	5233037
BEMP_M2	10/03/2026	7	Blue mackerel	220	85	Empty	523535	5233037
BEMP_M2	10/03/2026	8	Blue mackerel	210	85	Unidentifiable digested material	523535	5233037
BEMP_M2	10/03/2026	9	Blue mackerel	195	85	Empty	523535	5233037
BEMP_M2	10/03/2026	10	Blue mackerel	200	65	Unidentifiable digested material	523535	5233037
BEMP_M2	10/03/2026	11	Australian Salmon	305	250	Bait fish	523535	5233037
BEMP_M2	10/03/2026	12	Australian Salmon	320	295	Unidentifiable digested material	523535	5233037
BEMP_M3	10/03/2026	1	Australian Salmon	275	340	Empty	523752	5225263
BEMP_M3	10/03/2026	2	Australian Salmon	300	305	Empty	523752	5225263
BEMP_M3 Little Oyster Cove	10/03/2026	3	Australian Salmon	295	305	Bait fish	521653	5225040
BEMP_M3 Little Oyster Cove	10/03/2026	4	Australian Salmon	310	400	Bait fish	521653	5225040
BEMP_M3 Little Oyster Cove	10/03/2026	5	Australian Salmon	290	355	Empty	521653	5225040
BEMP_M3 Little Oyster Cove	10/03/2026	6	Blue mackerel	310	260	Bait fish	521653	5225040
BEMP_M3 Little Oyster Cove	10/03/2026	7	Australian Salmon	285	300	Bait fish	521653	5225040
BEMP_M3 Little Oyster Cove	11/03/2026	8	Australian Salmon	270	275	Bait fish	521653	5225040
BEMP_M3	13/03/2026	9	Gurnard	280	390	Crab	523752	5225263
BEMP_M3 Apollo Bay N	13/03/2026	10	Bluethroat wrasse	250	285	Crab	523177	5222852
BEMP_M3 Apollo Bay N	13/03/2026	11	Bluethroat wrasse	246	250	Algae	523177	5222852
BEMP_M3 Apollo Bay N	13/03/2026	12	Barber Perch	170	80	Bait fish	523177	5222852

Table A5.4: Wild fish sample data: Week 3 (T1+T2).

Site	Date	Fish No.	Species	Length (mm)	Weight (g)	Gut Content	Easting	Northing
<1 km	17/03/2026	1	Red Cod	350	540	Bait fish	524447	5228827
<1 km	17/03/2026	2	Red Cod	335	435	Green algae	524447	5228827
<1 km	17/03/2026	3	Red Cod	355	485	Crustacean remains	524447	5228827
<1 km	17/03/2026	4	Red Cod	350	455	Bait fish, shrimp	524447	5228827
<1 km	17/03/2026	5	Senator Wrasse	250	215	Green algae	524447	5228827
<1 km	17/03/2026	6	Senator Wrasse	220	130	Empty	524447	5228827
<1 km	17/03/2026	7	Bluethroat wrasse	275	360	White flesh	524447	5228827
<1 km	17/03/2026	8	Bluethroat wrasse	275	330	Limpet shells	524447	5228827
<1 km	17/03/2026	9	Bluethroat wrasse	225	190	Shell grit	524447	5228827
<1 km	17/03/2026	10	Bluethroat wrasse	235	180	Shell grit, eggs	524447	5228827
<1 km	17/03/2026	11	Bluethroat wrasse	230	170	Empty	524447	5228827
<1 km	17/03/2026	12	Barber Perch	190	100	Empty	524447	5228827
BEMP_M2	17/03/2026	1	Blue mackerel	205	85	Whitebait	523535	5233037
BEMP_M2	17/03/2026	2	Blue mackerel	220	110	Whitebait	523535	5233037
BEMP_M2	17/03/2026	3	Blue mackerel	210	95	Empty	523535	5233037
BEMP_M2	17/03/2026	4	Blue mackerel	210	95	Empty	523535	5233037
BEMP_M2	17/03/2026	5	Blue mackerel	217	95	Unidentifiable digested material	523535	5233037
BEMP_M2	17/03/2026	6	Jack Mackerel	225	95	Unidentifiable digested material	523535	5233037
BEMP_M2	17/03/2026	7	Jack Mackerel	240	135	Unidentifiable digested material	523535	5233037
BEMP_M2	17/03/2026	8	Jack Mackerel	250	135	Empty	523535	5233037
BEMP_M2	17/03/2026	9	Jack Mackerel	245	150	Empty	523535	5233037
BEMP_M2	17/03/2026	10	Jack Mackerel	230	125	Unidentifiable digested material	523535	5233037
BEMP_M2	17/03/2026	11	Jack Mackerel	250	155	Empty	523535	5233037
BEMP_M2	17/03/2026	12	Jack Mackerel	235	120	Empty	523535	5233037
BEMP_M3 Apollo Bay N	17/03/2026	1	Australian Salmon	440	1060	Bait fish	523177	5222852
BEMP_M3 Apollo Bay N	17/03/2026	2	Australian Salmon	370	640	Empty	523177	5222852
BEMP_M3 Apollo Bay N	17/03/2026	3	Australian Salmon	410	775	Empty	523177	5222852
BEMP_M3 Apollo Bay N	17/03/2026	4	Australian Salmon	435	955	Empty	523177	5222852
BEMP_M3 Apollo Bay N	17/03/2026	5	Australian Salmon	440	1010	Bait fish	523177	5222852
BEMP_M3 Apollo Bay N	17/03/2026	6	Australian Salmon	415	855	Empty	523177	5222852
BEMP_M3 Apollo Bay N	17/03/2026	7	Bluethroat wrasse	395	1095	Shell grit	523177	5222852
BEMP_M3 Apollo Bay N	17/03/2026	8	Bluethroat wrasse	425	1450	Crab, shell grit	523177	5222852
BEMP_M3 Apollo Bay N	17/03/2026	9	Bluethroat wrasse	407	1150	Empty	523177	5222852
BEMP_M3 Apollo Bay N	17/03/2026	10	Bluethroat wrasse	320	540	Shell grit	523177	5222852
BEMP_M3 Apollo Bay N	17/03/2026	11	Bluethroat wrasse	300	450	Shell grit	523177	5222852
BEMP_M3 Apollo Bay N	17/03/2026	12	Bluethroat wrasse	290	410	Shell grit	523177	5222852

Table A5.5: Wild fish sample data: Week 4 (T1+T2).

Site	Date	Fish No.	Species	Length (mm)	Weight (g)	Gut Content	Easting	Northing
BEMP_M3 Apollo Bay N	24/03/2026	1	Bluethroat wrasse	300	335	Shell grit	523177	5222852
BEMP_M3 Apollo Bay N	24/03/2026	2	Bluethroat wrasse	400	845	Shell grit	523177	5222852
BEMP_M3	24/03/2026	3	Australian Salmon	370	505	Unidentifiable digested material	523752	5225263
BEMP_M3	24/03/2026	4	Australian Salmon	360	550	Unidentifiable digested material	523752	5225263
BEMP_M3	24/03/2026	5	Australian Salmon	330	360	Unidentifiable digested material	523752	5225263
BEMP_M3	25/03/2026	6	Australian Salmon	271	160	Empty	523752	5225263
BEMP_M3	25/03/2026	7	Australian Salmon	285	160	Bait fish	523752	5225263
BEMP_M3 Little Oyster Cove	25/03/2026	8	Australian Salmon	240	320	Bait fish	521653	5225040
BEMP_M3	25/03/2026	9	Australian Salmon	240	332	Empty	523752	5225263
BEMP_M3	26/03/2026	10	Australian Salmon	389	750	Empty	523752	5225263
BEMP_M3	26/03/2026	11	Australian Salmon	365	590	Empty	523752	5225263
BEMP_M3	26/03/2026	12	Australian Salmon	410	880	Unidentifiable digested material	523752	5225263

Table A5.6: Wild fish sample data: Week 5 (T1+T2).

Site	Date	Fish No.	Species	Length (mm)	Weight (g)	Gut Content	Easting	Northing
BEMP_M3 Apollo Bay N	1/04/2026	1	Barracouta	220	35	Empty	523177	5222852
BEMP_M3 Apollo Bay N	1/04/2026	2	Barracouta	208	35	Empty	523177	5222852
BEMP_M3 Apollo Bay N	1/04/2026	3	Jack Mackerel	243	125	Unidentifiable digested material	523177	5222852
BEMP_M3 Apollo Bay N	1/04/2026	4	Jack Mackerel	236	125	Unidentifiable digested material	523177	5222852
BEMP_M3 Apollo Bay N	1/04/2026	5	Jack Mackerel	220	90	Unidentifiable digested material	523177	5222852
BEMP_M3 Apollo Bay N	1/04/2026	6	Jack Mackerel	285	215	Unidentifiable digested material	523177	5222852
BEMP_M3 Apollo Bay N	1/04/2026	7	Jack Mackerel	218	100	Unidentifiable digested material	523177	5222852
BEMP_M3 Apollo Bay N	1/04/2026	8	Jack Mackerel	212	85	Unidentifiable digested material	523177	5222852
BEMP_M3 Apollo Bay N	1/04/2026	9	Jack Mackerel	230	115	Unidentifiable digested material	523177	5222852
BEMP_M3 Apollo Bay N	1/04/2026	10	Jack Mackerel	212	100	Unidentifiable digested material	523177	5222852
BEMP_M3 Apollo Bay N	1/04/2026	11	Jack Mackerel	226	100	Unidentifiable digested material	523177	5222852
BEMP_M3 Apollo Bay N	1/04/2026	12	Jack Mackerel	226	100	Unidentifiable digested material	523177	5222852

Table A5.7: Wild fish sample data: Week 6 (T1+T2).

Site	Date	Fish No.	Species	Length (mm)	Weight (g)	Gut Content	Easting	Northing
BEMP_M3 Little Oyster Cove	8/04/2026	1	Barracouta	270	35	Empty	521653	5225040
BEMP_M3 Little Oyster Cove	8/04/2026	2	Barracouta	269	30	Empty	521653	5225040
BEMP_M3 Little Oyster Cove	8/04/2026	3	Barracouta	246	30	Empty	521653	5225040
BEMP_M3 Little Oyster Cove	8/04/2026	4	Barracouta	265	40	Unidentifiable digested material	521653	5225040
BEMP_M3 Little Oyster Cove	8/04/2026	5	Barracouta	238	25	Bait fish	521653	5225040
BEMP_M3 Little Oyster Cove	8/04/2026	6	Barracouta	218	25	Empty	521653	5225040
BEMP_M3 Little Oyster Cove	8/04/2026	7	Barracouta	258	35	Fish	521653	5225040
BEMP_M3 Little Oyster Cove	8/04/2026	8	Barracouta	204	10	Empty	521653	5225040
BEMP_M3 Apollo Bay N	8/04/2026	9	Blue mackerel	230	60	Unidentifiable digested material	523177	5222852
BEMP_M3 Apollo Bay N	8/04/2026	10	Blue mackerel	228	60	Unidentifiable digested material	523177	5222852
BEMP_M3 Apollo Bay N	8/04/2026	11	Blue mackerel	218	60	Unidentifiable digested material	523177	5222852
BEMP_M3 Apollo Bay N	8/04/2026	12	Blue mackerel	224	60	White flesh	523177	5222852

Table A5.8: Wild fish sample data: Week 7 (T1+T2).

Site	Date	Fish No.	Species	Length (mm)	Weight (g)	Gut Content	Easting	Northing
BEMP_M3 Little Oyster Cove	15/04/2026	1	Australian Salmon	300	520	Empty	521653	5225040
BEMP_M3	15/04/2026	2	Australian Salmon	320	550	Bait fish	523752	5225263
BEMP_M3 Little Oyster Cove	15/04/2026	3	Australian Salmon	330	545	Empty	521653	5225040
BEMP_M3	15/04/2026	4	Australian Salmon	280	275	White flesh	523752	5225263
BEMP_M3	15/04/2026	5	Australian Salmon	320	505	White flesh	523752	5225263
BEMP_M3	15/04/2026	6	Australian Salmon	325	510	Unidentifiable digested material	523752	5225263
BEMP_M3 Little Oyster Cove	15/04/2026	7	Australian Salmon	320	515	Empty	521653	5225040
BEMP_M3	15/04/2026	8	Australian Salmon	330	555	Whitebait	523752	5225263
BEMP_M3 Little Oyster Cove	15/04/2026	9	Australian Salmon	320	660	Empty	521653	5225040
BEMP_M3	15/04/2026	10	Australian Salmon	340	600	White flesh	523752	5225263
BEMP_M3 Apollo Bay N	15/04/2026	11	Bluethroat wrasse	240	245	Shell grit	523177	5222852
BEMP_M3	15/04/2026	12	Rock Cod	380	520	White flesh	523177	5222852

Appendix 6: Benthic video survey – summary data.

Table A6.1: Benthic video observations for pre-treatment and post-treatment surveys conducted by Fulcrum Robotics for Treatment T1.

Treatment	Survey	Site	Date	Comments
T1	**Pre-treatment**	**TS_PB14**	**6/02/2026**	Seabed: Marine snow, silty/sandy sediment, shells and shell fragments, feed and faeces, light patchy *Beggiatoa* sp. Fauna: NP seastar, ascidian sp., biscuit star sp., fire crab, swarming crustacea, side-gilled slug, *Nebalia* sp., vase tunicate, smooth stingray, egg masses, seal interaction, capitellid sp. Flora: Benthic red and brown algae sp., green and red algae fragments, benthic epiphytic growth Other: Terrestrial plant material. Pellets: Many
T1	**Pre-treatment**	**TS_PB16**	**6/02/2026**	Seabed: Marine snow, silty/sandy sediment, feed and faeces, shells and shell fragments, light patchy *Beggiatoa* sp. Fauna: Egg mass, *Nebalia* sp., sponge sp. fragments, crab sp., side-gilled slug Flora: Benthic epiphytic growth, benthic brown and red algae sp., red and green algae fragments. Other: Terrestrial plant material. Pellets: Few
T1	**Pre-treatment**	**TS_PB19**	**6/02/2026**	Seabed: Marine snow, sandy/silty sediment, shells and shell fragments, patchy *Beggiatoa* sp., feed and faces Fauna: NP seastar, ascidian sp., crab spp., fire crab, *Nebalia* sp., egg masses, vase tunicate, side-gilled slug, goby sp., biscuit star Flora: Green algae fragments, benthic red and brown algae sp. Other: NA. Pellets: Few
T1	**Post-treatment**	**TS_PB14**	**25/02/2026**	Seabed: Marine snow, silty sediment, shells and shell fragments, feed and faeces, biofouling debris Fauna: NP seastar, vase tunicate, crab sp., fire crab, anemone sp., ascidian sp., sponge sp., goby sp., swarming crustacea, whiting sp., pelagic baitfish sp., *Nebalia* sp., southern spindle Flora: Various algae fragments Other: Tape (GoPro 1:27), rope (GoPro 2:29), debris? (GoPro 3:00), chain (GoPro 3:17), rope coil (GoPro 3:43), terrestrial plant material. Pellets: Few
T1	**Post-treatment**	**TS_PB16**	**25/02/2026**	Seabed: Marine snow, silty sediment, biofouling debris, shells and shell fragments, NZ screwshell, feed and faeces, patchy *Beggiatoa* sp. Fauna: NP seastar, anemone sp., ascidian sp., crab spp., fire crab, side-gilled slug, swarming crustacea, whiting sp., vase tunicate Flora: Benthic epiphytes, red and green algae fragments, benthic red algae fragments, filamentous green algae sp. Other: Terrestrial plant material, rope (GoPro 3:21, 3:57), seal interaction. Pellets: Few
T1	**Post-treatment**	**TS_PB19**	**25/02/2026**	Seabed: Marine snow, silty sediment, biofouling debris, shells and shell fragments, feed and faeces, patchy *Beggiatoa* sp., NZ screwshell Fauna: NP seastar, ascidian sp., crab spp., fire crab, egg mass, vase tunicate, side-gilled slug, granular seastar, biscuit star Flora: Green and red algae fragments Other: Terrestrial plant material, blue debris (GoPro 2:53). Pellets: Few

Table A6.2: Benthic video observations for pre-treatment surveys conducted by Fulcrum Robotics for Treatment T2.

Treatment	Survey	Site	Date	Comments
T2	**Pre-treatment**	**TS_PB01**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, feed and faeces, mussel shells, patches of *Beggiatoa* sp. Fauna: NP seastar, oyster sp., ascidian sp., sponge sp., goby sp., egg mass, fire crab, vase tunicate. Flora: Red benthic algae, various drift algae, brown algal growth on substrate. Other: Strong current, rope (GoPro 1:45), seal interaction, terrestrial plant material, unknown debris? (ROV 3:49). Pellets: Few
T2	**Pre-treatment**	**TS_PB02**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, feed and faeces, burrows and mounds, mussel shells, patchy *Beggiatoa* sp. Fauna: NP seastar, seastar sp., goby sp., side gilled slug, ascidian spp., egg mass, NZ screwshell, swarming crustacea, vase tunicate, sponge sp. Flora: Various drift algae, brown algal growth on substrate, red benthic algae. Other: Branch, rope and bullet weight (GoPro 2:24), stick, tyre (GoPro 4:23). Pellets: Few
T2	**Pre-treatment**	**TS_PB03**	**21/02/2026**	Seabed: Sandy sediment, marine snow, feed and faeces, shells and shell fragments, mussel shells, patchy *Beggiatoa* sp., urchin test fragment. Fauna: NP seastar, anemone sp., vase tunicate, ascidian sp., egg mass, fire crab, oyster sp., bryozoan sp., goby sp., seastar sp. Flora: Various drift algae, red and green benthic algae, brown algal growth on substrate. Other: Rope (GoPro 2:04, 5:13), dive reset, terrestrial plant material, strop? (GoPro 6:39), tyre (GoPro 7:42), strong current. Pellets: Few
T2	**Pre-treatment**	**TS_PB04**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, mussel shells, ascidian stalks, feed and faeces, patches of *Beggiatoa* sp. Fauna: Ascidian sp., seastar sp., ascidian sp., NZ screwshell, fire crab, vase tunicate, side gilled slug, goby sp., swarming crustacea, southern hulafish. Flora: Brown algal growth on substrate, red and green benthic algae, various drift algae. Other: Bullet weight (GoPro 3:07, 4:11). Pellets: Few
T2	**Pre-treatment**	**TS_PB05**	**21/02/2026**	Seabed: Sandy sediment, marine snow, feed and faeces, shells and shell fragments, mussel shells, patchy *Beggiatoa* sp. Fauna: NP seastar, fire crab, side gilled slug, crab sp., vase tunicate, biscuit star, sponge sp., bryozoan sp., anemone sp. Flora: Red and green benthic algae, brown algal growth on substrate, various drift algae. Other: Strong current, terrestrial plant material, tyre (GoPro 2:59), net (GoPro 3:49), rope (GoPro 4:20), seal interaction. Pellets: Few
T2	**Pre-treatment**	**TS_PB06**	**21/02/2026**	Seabed: Sandy sediment, marine snow, feed and faeces, shells and shell fragments, mussel shells, patchy *Beggiatoa* sp. Fauna: NP seastar, seastar sp., ascidian sp., fire crab, egg mass, bryozoan sp., side gilled slug, goby sp., swarming crustacea, spindle shell, vase tunicate. Flora: Red and green benthic algae, brown algal growth on substrate, various drift algae. Other: Gate rope on descent, terrestrial plant material. Pellets: Few
T2	**Pre-treatment**	**TS_PB07**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, patches of *Beggiatoa* sp., mussel and oyster shells. Fauna: NP seastar, vase tunicate, fire crab, egg mass, crab sp., anemone sp., capitellid sp., bryozoan sp., crowned nudibranch. Flora: Brown algal growth on substrate, red benthic algae, various drift algae. Other: Hose? (GoPro 1:17), terrestrial plant material, debris? (GoPro 2:03), rope (GoPro 2:52, 4:18). Pellets: Few
T2	**Pre-treatment**	**TS_PB08**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, feed and faeces, mussel shells, Fauna: NP seastar, ascidian sp., seastar sp., fire crab, bryozoan sp., egg mass, vase tunicate, side gilled slug, crab sp., swarming crustacea, goby sp. Flora: Red and green benthic algae, brown algal growth on substrate, various drift algae. Other: Terrestrial plant material, bottle (GoPro 3:30), bullet weight (GoPro 3:34), red tag (GoPro 4:08), rope (GoPro 4:18, 4:45), rope? (GoPro 4:33). Pellets: Few
T2	**Pre-treatment**	**TS_PB09**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, feed and faeces, patches of *Beggiatoa* sp. Fauna: Vase tunicate, fire crab, NP seastar, crab sp., anemone sp., egg mass, crowned nudibranch, bryozoan sp., sponge sp., ascidian sp., capitellid sp? (ROV 4:21). Flora: Brown algal growth on substrate, red and green benthic algae, various drift algae. Other: Rope (GoPro 1:50, 2:40), terrestrial plant material, debris (GoPro 3:39), strop (GoPro 4:10).. Pellets: Few
T2	**Pre-treatment**	**TS_PB10**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, feed and faeces, patches of *Beggiatoa* sp. Fauna: Ascidian sp., NP seastar, seastar sp., side gilled slug, bryozoan sp., crab spp., fire crab, goby sp., vase tunicate, swarming crustacea, spindle shell, blenny sp., piecrust crab. Flora: Green and red benthic algae, various drift algae, brown algal growth of substrate. Other: Rope (GoPro 1:44, 3:05), terrestrial plant material, net interaction, debris? (GoPro 5:00). Pellets: Few

Table A6.2 (continued): Benthic video observations for pre-treatment surveys conducted by Fulcrum Robotics for Treatment T2.

Treatment	Survey	Site	Date	Comments
T2	**Pre-treatment**	**TS_PB11**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, mussel shells, feed and faeces, patchy *Beggiatoa* sp. Fauna: Ascidian spp., NP seastar, vase tunicate, fire crab, egg mass, side gilled slug. Flora: Various drift algae, red benthic algae, brown algal growth on substrate. Other: Terrestrial plant material, hose (GoPro 4:00). Pellets: Few
T2	**Pre-treatment**	**TS_PB12**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, feed and faeces, patches of *Beggiatoa* sp., Fauna: Seastar sp., NP seastar, ascidian sp., fire crab, egg mass, vase tunicate, bryozoan sp., goby sp., crab sp., sponge sp., capitellid sp.? (e.g. ROV 3:15) Flora: Brown algal growth on substrate, red and green benthic algae, various drift algae. Other: Terrestrial plant material, rope? (GoPro 2:06, 4:50, ROV 3:21), rope (GoPro 2:13, 2:27, 2:33).. Pellets: Few
T2	**Pre-treatment**	**TS_PB15**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, patches of *Beggiatoa* sp., faeces. Fauna: NP seastar, anemone sp., vase tunicate, egg mass, fire crab, sponge sp., capitellid sp. Flora: Various drift algae, brown algal growth on substrate, red and green benthic algae. Other: Terrestrial plant material, rope (GoPro 4:51). Pellets: No
T2	**Pre-treatment**	**TS_PB17**	**21/02/2026**	Seabed: Silty sediment, marine snow, feed and faeces, shells and shell fragments, patches of *Beggiatoa* sp. Fauna: NP seastar, ascidian sp., smooth ray, side gilled slug, fire crab, goby sp., vase tunicate, egg mass, bryozoan sp., swarming crustacea. Flora: Red and green benthic algae, various drift algae, brown algal growth on substrate. Other: Rope (GoPro 2:13, 5:16), rope? (GoPro 2:24), terrestrial plant material. Pellets: Few
T2	**Pre-treatment**	**TS_PB18**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, mussel shells, feed and faeces, patchy *Beggiatoa* sp. Fauna: NP seastar, mackerel sp., ascidian spp., crab sp., vase tunicate, fire crab, egg mass, anemone sp., little weed whiting. Flora: Various drift algae, red and green benthic algae, brown algal growth on substrate. Other: Terrestrial plant material, rope (GoPro 3:18, 3:27, 4:50), seal interaction. Pellets: Few
T2	**Pre-treatment**	**TS_PB20**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, mussel shells, patches of *Beggiatoa* sp., feed and faeces. Fauna: NP seastar, anemone sp., ascidian sp., fire crab, bryozoan sp., vase tunicate, silver trevally, side gilled slug, flathead sp., seastar sp., egg mass. Flora: Various drift algae, red and green benthic algae, brown algal growth on substrate. Other: Terrestrial plant material, strop (GoPro 2:12), tyre (GoPro 4:44), rope and tyre (GoPro 5:26), bubble from ROV (GoPro 5:33).. Pellets: Few
T2	**Pre-treatment**	**TS_PB21**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, mussel shells, patches of *Beggiatoa* sp., feed and faeces, unidentified organic (GoPro 3:22). Fauna: NP seastar, ascidian sp., side gilled slug, fire crab, eight-armed seastar, regular seastar, vase tunicate, capitellid sp. Flora: Red and green benthic algae, brown algal growth on substrate, various drift algae. Other: Terrestrial plant material, rope (GoPro 3:32, 3:57), debris? (GoPro 3:38), tyre (GoPro 4:21).. Pellets: Few
T2	**Pre-treatment**	**TS_PB22**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, patches of *Beggiatoa* sp., feed and faeces, mussel shells. Fauna: NP seastar, ascidian sp., fire crab, egg mass, side gilled slug, swarming crustacea, vase tunicate, anemone sp., capitellid sp? (e.g. ROV 4:42). Flora: Various drift algae, benthic red and green algae, brown algal growth on substrate. Other: Rope? (GoPro 1:58), terrestrial plant material, rope (GoPro 2:20), bullet weight (GoPro 4:57, 5:05). . Pellets: Few
T2	**Pre-treatment**	**TS_PB23**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, mussel shells, patches of *Beggiatoa* sp., feed and faeces. Fauna: Flathead sp., ascidian sp., NP seastar, crab sp., fire crab, bryozoan sp., swarming crustacea, capitellid sp., vase tunicate, side gilled slug. Flora: Brown algal growth on substrate, red and green benthic algae, various drift algae. Other: Terrestrial plant material, bullet weights (GoPro 2:45), rope (GoPro 3:03, 3:42), duct tape (GoPro 4:47).. Pellets: Few
T2	**Pre-treatment**	**TS_PB24**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, mussel shells, patches of *Beggiatoa* sp., feed and faeces. Fauna: NP seastar, ascidian sp., egg mass, fire crab, vase tunicate, side gilled slug, octopus sp., flathead sp., swarming crustacea, capitellid sp? (e.g. ROV 5:25). Flora: Brown algal growth on substrate, red and green benthic algae. Other: Rope (GoPro 2:20, 5:20), mesh bag (GoPro 3:03), terrestrial plant material, debris (GoPro 4:16), hose (GoPro 4:57), debris? (GoPro 5:02).. Pellets: Few
T2	**Pre-treatment**	**TS_PB26**	**21/02/2026**	Seabed: Sandy sediment, marine snow, shells and shell fragments, mussel shells, patches of *Beggiatoa* sp., feed and faeces, crab carapace. Fauna: NP seastar, ascidian sp., NZ screwshell, fire crab, capitellid sp., sponge sp? (GoPro 3:53). Flora: Brown algal growth on substrate, various drift algae, red and green benthic algae. Other: Terrestrial plant material, bullet weight (GoPro 3:48, 4:03), rope (GoPro 4:33). Pellets: Few

Table A6.3: Benthic video observations for post-treatment surveys conducted by Fulcrum Robotics for Treatment T2.

Treatment	Survey	Site	Date	Comments
T2	**Post-treatment**	**TS_PB01**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, shells and shell fragments (mussel, scallops, oyster), feed and faeces Fauna: NP seastar, egg mass, ascidian sp., fire crab, vase tunicate, swarming crustacea, *Nebalia* sp., side-gilled slug, goby sp. Flora: Benthic epiphytes, red algae and fragments Other: Terrestrial plant material, rope (GoPro 1:50). Pellets: Few
T2	**Post-treatment**	**TS_PB02**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, scallop), patches of *Beggiatoa* sp. Fauna: NP seastar, regular seastar, vase tunicate, side-gilled slug, egg mass, fire crab, *Nebalia* sp., goby sp., regular seastar, shore crab, blenny sp., ascidian sp. Flora: Benthic epiphytes, benthic brown algae sp. Other: Seal interaction, bullet weight and rope (GoPro 5:15). Pellets: Few
T2	**Post-treatment**	**TS_PB03**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, scallop, oyster), patches of *Beggiatoa* sp. Fauna: Ascidian sp., NP seastar, fire crab, vase tunicate, side-gilled slug, biscuit star, swarming crustacea Flora: Benthic epiphytes, red algae and fragments Other: Terrestrial plant material, rope (GoPro 3:05, 4:25), bottle (ROV 5:30). Pellets: Few
T2	**Post-treatment**	**TS_PB04**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, faeces, shells and shell fragments (mussel) Fauna: NP seastar, ascidian sp., *Nebalia* sp., egg mass, side-gilled slug, sponge sp., fire crab, goby sp., vase tunicate, regular seastar Flora: Benthic epiphytes, brown algae fragments Other: Bullet weight (GoPro 1:35), rope (GoPro 4:13), mesh (GoPro 4:55), terrestrial plant material. Pellets: No
T2	**Post-treatment**	**TS_PB05**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, oyster, scallop), patchy *Beggiatoa* sp. Fauna: NP seastar, ascidian spp., fire crab, side-gilled slug, crab sp., vase tunicate, pelagic fish sp., anemone sp., egg mass Flora: Benthic epiphytes, red and green algae fragments, benthic red algae sp. Other: Terrestrial plant material, bottle and rope (GoPro 2:01), rope (GoPro 2:28, 4:02). Pellets: Few
T2	**Post-treatment**	**TS_PB06**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, scallop), patches of *Beggiatoa* sp. Fauna: NP seastar, egg mass, side-gilled slug, fire crab, *Nebalia* sp. Flora: Benthic epiphytes, green and red algae fragments. Other: Rope (GoPro 2:34), terrestrial plant material. Pellets: Few
T2	**Post-treatment**	**TS_PB07**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, scallop, oyster), patches of *Beggiatoa* sp., crab carapace Fauna: NP seastar, vase tunicate, crab sp., fire crab, egg mass, anemone sp., side-gilled slug, ascidian sp., crowned nudibranch Flora: Brown and red algae fragments, benthic epiphytes Other: Terrestrial plant material, strop (GoPro 1:36), hose (GoPro 2:25), rope (GoPro 2:27, 2:55, 3:11), seal interaction. Pellets: Few
T2	**Post-treatment**	**TS_PB08**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, scallop), patches of *Beggiatoa* sp. Fauna: NP seastar, ascidian sp., side-gilled slug, vase tunicate, fire crab, goby sp., regular seastar, egg mass, swarming crustacea Flora: Benthic epiphytes, red and green algae fragments, benthic red algae sp. Other: Terrestrial plant material, rope (ROV 2:36, 3:57), bullet weight and ropes (GoPro 3:02), debris? (ROV 03:35). Pellets: Few
T2	**Post-treatment**	**TS_PB09**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, scallop) Fauna: NP seastar, crab sp., fire crab, vase tunicate, ascidian sp., side-gilled slug, egg mass, anemone sp., southern spindle Flora: Benthic epiphyte, red algae fragments Other: Rope (GoPro 2:38, 2:55, 3:38, 5:08), blue tape (GoPro 4:17). Pellets: Few
T2	**Post-treatment**	**TS_PB10**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, scallop, screwshell sp.), sea tulip stalks Fauna: NP seastar, ascidian sp., side-gilled slug, fire crab, crab sp., vase tunicate, egg mass, *Nebalia* sp. Flora: Benthic epiphytes, benthic red algae spp., green and red algae fragments Other: Terrestrial plant material. Pellets: Few
T2	**Post-treatment**	**TS_PB11**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, scallop, oyster), patches of *Beggiatoa* sp. Fauna: NP seastar, ascidian spp., vase tunicate, fire crab, granular seastar, egg mass, side-gilled slug, crab sp., whiting sp., shore crab Flora: Benthic epiphytes, various algae fragments, filamentous brown algae sp. Other: Terrestrial plant material, weight (GoPro 1:35), hose (3:58), rope (5:05)], seal interaction. Pellets: Few
T2	**Post-treatment**	**TS_PB12**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, scallop) Fauna: Ascidian spp., NP seastar, fire crab, vase tunicate, crab sp., egg mass, side-gilled slug, regular seastar, goby sp., *Nebalia* sp. Flora: Benthic epiphytes, red and green algae fragments, benthic red algae sp. Other: Terrestrial plant material. Pellets: No

Table A6.3 (continued): Benthic video observations for post-treatment surveys conducted by Fulcrum Robotics for Treatment T2.

Treatment	Survey	Site	Date	Comments
T2	**Post-treatment**	**TS_PB15**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, oyster, scallop), patches of *Beggiatoa* sp. Fauna: NP seastar, ascidian sp., fire crab, crab sp., vase tunicate, side-gilled slug, egg mass, anemone sp. Flora: Benthic epiphytes, various algae fragments, benthic brown algae Other: Terrestrial plant material, snood (GoPro 2:22), rope (GoPro 4:59). Pellets: Few
T2	**Post-treatment**	**TS_PB17**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, faeces, shells and shell fragments (mussel, scallop), patches of *Beggiatoa* sp. Fauna: NP seastar, fire crab, vase tunicate, ascidian sp., side-gilled slug, regular seastar, egg mass, crab sp. Flora: Benthic epiphytes, various algae fragments. Other: Terrestrial plant material. Pellets: No
T2	**Post-treatment**	**TS_PB18**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, scallop, oyster), patches of *Beggiatoa* sp. Fauna: NP seastar, anemone sp., *Nebalia* sp., mackerel sp., swarming crustacea, ascidian sp., crab sp., southern spindle, regular seastar Flora: Benthic epiphytes, benthic red algae sp., red algae fragments Other: Terrestrial plant material, log, hose (GoPro 4:10). Pellets: Few
T2	**Post-treatment**	**TS_PB20**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, scallop, oyster, screwshell), patches of *Beggiatoa* sp. Fauna: NP seastar, fire crab, anemone sp., ascidian sp., cod sp. (GoPro 4:18), gurnard sp., vase tunicate, flathead sp., regular seastar Flora: Benthic epiphytes, green and red algae fragments, benthic red algae sp. Other: Terrestrial plant material. Pellets: Few
T2	**Post-treatment**	**TS_PB21**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, scallop), patches of *Beggiatoa* sp. Fauna: NP seastar, ascidian sp., vase tunicate, fire crab, egg mass, side-gilled slug Flora: Benthic epiphytes, various algae fragments, benthic red algae sp. Other: Bottle? (GoPro 2:17), rope (GoPro 2:19, 3:31). Pellets: Few
T2	**Post-treatment**	**TS_PB22**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, scallop), sea tulip stalks, patches of *Beggiatoa* sp. Fauna: NP seastar, ascidian sp., fire crab, side-gilled slug, flathead sp., *Nebalia* sp., swarming crustacea Flora: Benthic epiphytes, benthic red and brown algae sp., various algae fragments, seagrass fragment Other: Rope (GoPro 1:58, 2:47), terrestrial plant material. Pellets: Few
T2	**Post-treatment**	**TS_PB23**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, faeces, shells and shell fragments (mussel, scallop, oyster), patches of *Beggiatoa* sp. Fauna: NP seastar, ascidian sp., side-gilled slug, fire crab, vase tunicate, egg mass, crab sp., crowned nudibranch Flora: Benthic epiphytes, red and green algae fragments, benthic red algae sp. Other: Terrestrial plant material . Pellets: No
T2	**Post-treatment**	**TS_PB24**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, feed and faeces, shells and shell fragments (mussel, scallop), patchy *Beggiatoa* sp. Fauna: NP seastar, ascidian sp., unknown fish sp. (GoPro 3:28), fire crab Flora: Benthic epiphytes, various algae fragments, benthic brown algae sp. Other: Terrestrial plant material, ropes (GoPro 3:30, 3:50, 4:36, 4:42, 5:53), bag (GoPro 4:27), unknown debris (GoPro 5:21), cable? (GoPro 5:26), seal interaction. Pellets: Few
T2	**Post-treatment**	**TS_PB26**	**4/03/2026**	Seabed: Marine snow, sandy/silty sediment, faeces, shells and shell fragments (mussel, oyster, scallop, screwshell), patches of *Beggiatoa* sp. Fauna: NP seastar, ascidian sp., fire crab, unknown fish sp. (GoPro 6:00) Flora: Benthic epiphytes, red and green algae fragments, benthic brown and red algae sp. Other: Terrestrial plant material. Pellets: No