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MEADOWBANK HATCHERY OTC RESIDUE MONITORING INTERIM REPORT

JANUARY/FEBRUARY 2026 TREATMENT

prepared for

Huon Aquaculture Company

June 2026

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2	Marine Solutions	13/05/2026		
		14/05/2026	A. Smark	
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		19/05/2026	A. Smark	
4	Marine Solutions	21/05/2026		
		26/05/2026	EPA	
5	Marine Solutions	29/05/2026		
6	Marine Solutions	03/05/2026	C. Selkirk	
		16/06/2026	EPA	
7	Marine Solutions	19/06/2026	C. Selkirk	

Cover photo: Derwent River at Meadowbank Hatchery Interim Report (Marine Solutions, January 2026).

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1 Introduction and Scope

An Environmental Antibiotic Residue Monitoring Survey was undertaken in the vicinity of Meadowbank Hatchery (Environmental Licence 9954/3), located downstream of the Meadowbank Dam on the Derwent River. Sampling methodologies and analyses were in accordance with the monitoring requirements stipulated by the Tasmanian Environmental Protection Authority (EPA) in the Therapeutant Monitoring Schedule – Environmental License No. 9954/3 Meadowbank Hatchery – January 2026.

An increase in water temperature of 5°C, maintained over three days was identified as a contributing stressor associated with increased mortalities at Meadowbank Hatchery. Diagnostic investigation confirmed bacterial enteritis with intralesional bacteria. Given concerns for fish welfare arising from elevated, non-self-limiting mortalities and the progressive nature of the disease, a broad-spectrum antibiotic was administered to control the bacterial enteritis, with temperatures forecast to remain elevated. The requirement to administer antibiotic treatment to the fish is documented in Fish Medication Authorities 1113.2F and 1114, copies of which have previously been provided to the EPA.

Medication Authorities were issued on 20 January (MA-1113.2F – treatment commenced 21 January) and 30 January (MA-1114 – treatment commenced 31 January). These authorisations did not represent two treatments of the site, but rather reflected the staged response to disease progression across separate tank groups. Tanks 4 and 7 were not included under MA-1113.2F as they had lower mortality rates at the time of the initial treatment decision. Following a subsequent increase in mortalities in these tanks, further intervention was required, and MA-1114 was issued on 30 January to authorise treatment of Tanks 4 and 7 (Table 1).

Table 1. Treatment schedule for antibiotic oxytetracycline at Meadowbank Hatchery.

Treatment	Days	Dates prescribed	OTC administered (kg)	Tanks treated	
1 **(MA-1113.2F)**	10	21/01/2026 – 30/01/2026	38.28	MB5 MB6 MB8 MB9	MB10 MB11 MB12
2 **(MA 1114)**	10	31/01/2026 – 9/02/2026	13.42	MB4 MB7	

The monitoring design provided by the EPA included sampling of sediment, wild fish and water quality for the purpose of monitoring residue of the antibiotic oxytetracycline (OTC).

The monitoring survey design included:

- Sediment sampling at six locations
- Water quality sampling at five locations
- Wild fish sampling at one location (12 wild fish per sample)

Sampling was conducted at an interval of:

- Day 1 of treatment (sediment and water)
- Mid treatment (sediment and water)
- Day 1 after completion of treatment (sediment, water and wild fish)
- Weekly until two consecutive results were less than the laboratory Limit of Reporting (LoR) (sediment, water and wild fish).

Table 2 provides an overview of survey frequency and completed sampling rounds for sediments, water and wild fish. Additional sampling weeks were added to the Therapeutant Monitoring Schedule for wild fish monitoring.

Table 2. Overview of OTC residue monitoring program for Meadowbank Hatchery according to the Antibiotic Residue Monitoring Schedule

Survey component	Frequency	Status	Number of survey rounds	Date	Sampling Rounds
Sediment	• Day 1 of treatment • Mid-treatment • Day 1 after completion of treatment • Weekly thereafter until two consecutive results are <LoR.	Ongoing	14	21/01/2026	Day 1 of treatment
				27/01/2026	Mid treatment
				10/02/2026	Day 1 post treatment
				17/02/2026	Day 7 post treatment
				24/02/2016	Day 14 post treatment
				3/03/2026	Day 21 post treatment
				10/03/2026	Week 4 post treatment
				17/03/2026	Week 5 post treatment
				24/03/2026	Week 6 post treatment
				2/04/2026	Week 7 post treatment
				10/04/2026	Week 8 post treatment
				16/04/2026	Week 9 post treatment
				20/04/2026	Week 10 post treatment
				30/04/2026	Week 11 post treatment
Water Quality	• Day 1 of treatment • Mid-treatment • Day 1 after completion of treatment • Weekly thereafter until two consecutive results are <LoR.	Concluded	9	21/01/2026	Day 1 of treatment
				27/01/2026	Mid treatment
				10/02/2026	Day 1 post treatment
				17/02/2026	Day 7 post treatment
				24/02/2016	Day 14 post treatment
				3/3/2026	Day 21 post treatment
				10/03/2026	Week 4 post treatment
				17/03/2026	Week 5 post treatment
				24/03/2026	Week 6 post treatment
Wild Fish	• Day 1 after completion of treatment • Day 7 after completion of treatment (*Further weekly wild fish sampling was requested by EPA until two consecutive results were <LoR)*	Concluded	6	10/02/2026	Day 1 post treatment
				17/02/2026	Day 7 post treatment
				24/02/2016	Day 14 post treatment
				3/3/2026	Day 21 post treatment
				10/03/2026	Week 4 post treatment
				17/03/2026	Week 5 post treatment

2 Methods

2.1 Sediment sampling

The survey design for the residue monitoring included sediment sampling at six sites (Figure 1). Sampling sites were specified by the EPA (coordinates provided in). Due to access restrictions at the following sites: River Derwent (upstream), Derwent River at hatchery discharge to Water, sludge tank, downstream (near) of hatchery discharge, downstream (far) of hatchery discharge, samples were taken at the closest possible point to coordinates provided by the EPA Appendix 4. Restrictions occurred because of no river/water access due to dense vegetation, steep banks or in the case of the sludge tank, infrastructure being in a different location to locations outlined in Therapeutant Monitoring Schedule – Environmental License No. 9954/3 Meadowbank Hatchery – January 2026.

Sediment samples were collected using a 150 mL AST white jar. Prior to sampling each site, a new pair of nitrile gloves were worn and the sample jar was labelled. Sediment was collected from below the water, after which excess water was carefully decanted while retaining the sediment fraction. Sediment sample jars supplied by AST had a height of 95 mm, sediment samples were therefore collected to a depth of approximately 95 mm. The jar lid was then securely fastened. Gloves were removed by folding them over the sample jar during removal to minimise contamination risk. Each sample jar was subsequently placed into a sealed plastic ziplock bag, stored in an eski for transport and preservation. Sampling was undertaken in a sequential order, commencing at sites assessed as having the lowest potential for contamination and progressing towards sites of higher potential contamination from day 1 post treatment. Samples were delivered to Analytical Services Tasmania (AST) within 12 hours of collection for analysis.

Samples were analysed for oxytetracycline hydrochloride (OTC) in sediment and the LoR was 0.01 mg/kg.

2.2 Water sampling

The survey design for the residue monitoring included water sampling at five sites (Figure 1). Sampling sites were specified by the EPA (coordinates provided in Appendix 4). Due to access restrictions at the following sites: River Derwent (upstream), Derwent River at hatchery discharge to Water, sludge tank, downstream (near) of hatchery discharge, downstream (far) of hatchery discharge, samples were taken at the closest possible point to coordinates provided by the EPA Appendix 4. Restrictions occurred because of no river/water access due to dense vegetation, steep banks.

Surface water samples were collected into labelled 250mL amber glass bottles provided by AST. Prior to sampling each, a new pair of nitrile gloves were worn and each sample container was labelled. Water samples were collected directly below the water surface to avoid surface debris and contamination. Following collection, gloves were removed by folding them over the sample container during removal to minimise contamination risk. Each sample container was subsequently placed into a sealed plastic ziplock bag, stored in an eski for transport and preservation, and the sample collection time was recorded on the submission sheet. Samples were chilled and delivered to AST within 12 hours of collection for analysis. Sampling was undertaken in a sequential order, commencing at sites assessed as having the lowest potential for contamination and progressing towards sites of higher potential contamination from day 1 post treatment.

Samples were analysed for oxytetracycline hydrochloride (OTC) in water and the LoR was 0.01 µg/L

Figure 1. Sediment and water sampling sites, and wild fish sampling zone for Meadowbank OTC monitoring. Sediment, water, and wild fish sampling locations are labelled, with GPS points collected during sampling shown as red circles. The wild fish sampling location is shown with a 50 m buffer zone

2.3 Wild fish sampling

The wild fish survey requirement for residue monitoring included an area of the Derwent River between the Meadowbank Hatchery discharge to water 1 km downstream (TMS for EL No. 9954/3, EPA 2026). Restricted access to the Derwent River due to deep water and high flows resulted in the sampling being conducted at intersection of the hatchery discharge channel and the Derwent River. Sampling was conducted at the same entrance point on each sampling occasion, and a buffer of a 50 m radius was implemented for the sample location (Figure 1). Sampling coordinates are provided in Appendix 4.

Wild fish sampling was conducted by electrofishing[1] on foot. Twelve fish were sampled on each monitoring occasion. Fish species, length and weight were recorded. Length (longest measurable point) was measured with a ruler, and weight was measured using calibrated fish scales. The tail (posterior to the anus) was retained for laboratory flesh analysis. Flesh samples were bagged and labelled (site, sample day, date, fish species and number) and delivered to AST within 12 hours of collection for analysis. Samples that could not be delivered within 12 hours were frozen as soon as possible after collection and analysed later.

Samples were analysed for oxytetracycline hydrochloride (OTC) in the fish tissue and the LoR was 0.01 mg/kg. On some occasions, as a result of the small sample size of wild fish the laboratory (AST) increased the LoR (Table 4).

[1] Electrofishing conducted under Environmental Monitoring Permit D26-27492 issued by the Aquaculture Branch of the Department of Natural Resources and Environment Tasmania.

3 Results

3.1 Sediment

Results of sediment sampling are summarised in Table 3. Huon Aquaculture only received the monitoring schedule on 23/01/2026, after treatment had already commenced (Day 1 of treatment was 21/01/2026). Therefore, only the sludge tank and settling tank were sampled on Day 1 of treatment, and the other sites from mid-treatment onwards (except where detailed below). OTC residue at the sludge tank and settlement pond was <LoR on Day 1 of treatment.

The upstream Derwent River site was sampled mid-treatment and every week until Week 12 post-treatment and returned OTC concentrations <LoR on every occasion.

OTC concentration was above LoR in the sludge tank from mid-treatment (290 mg/kg DMB) with peak concentrations occurring at Day 7 post-treatment (1800 mg/kg DMB) (Table 3). During post-treatment sampling the contents of the sludge tank were removed and the tank systems thoroughly cleaned. The sludge tank is the holding area for any solid matter removed from the system as water flows from the hatchery tanks to the settling ponds. Sludge from the tank was taken to Southern Waste Solutions (Copping) for disposal. The cleaning process commenced after sampling on Day 14 and was completed on 3 March 2026 (Day 21) and is considered to have contributed to the reduced OTC concentrations observed after Day 21 (Table 3).

OTC was present in sediment in the settlement pond inlet from mid-treatment to Day 21 post-treatment, with the highest elevation recorded on Day 1 post-treatment (3.5 mg/kg DMB). The settlement pond inlet sample returned OTC values <LoR consistently from Week 4.

At mid treatment there was a sampling oversight. The sample originally identified as the hatchery discharge was in fact collected from the downstream near location, while the downstream near sample was collected from the site approximately 500 m downstream (Figure 1). Therefore, OTC concentration was recorded in the Derwent River hatchery discharge to water from Day 1 post

treatment (0.23 mg/kg DMB) with the highest values recorded on Day 14 post-treatment (46 mg/kg DMB). OTC was present in the hatchery discharge at lower levels from Week 5 to Week 8 (inclusive). The hatchery discharge sample returned OTC values <LoR in Week 9 and Week 10. OTC concentrations were above LoR in Week 11 (0.04 mg/kg DMB) and Week 12 (0.06 mg/kg DMB).

OTC residue was <LoR at the downstream near site from Week 4 to Week 10. Detection above the LoR was recorded in Week 11 (0.04 mg/kg DMB).

Table 3. OTC concentration in sediments (in mg/kg DMB) sampled during the monitoring program at Meadowbank Hatchery. Results above the LoR (<0.01 mg/kg) are shaded in blue. Weeks where sampling did not occur are shaded in grey.

Site	Day 1 of treatment	Mid treatment	Post treatment												
			Day 1	Day 7	Day 14	Day 21	Week 4	Week 5	Week 6	Week 7	Week 8	Week 9	Week 10	Week 11	Week 12
	21/01/2026	27/01/2026	10/02/2026	17/02/2026	24/02/2026	3/03/2026	10/03/2026	17/03/2026	24/03/2026	2/04/2026	10/04/2026	16/04/2026	20/04/2026	30/04/2026	7/05/2026
River Derwent (upstream)		<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
Sludge tank	<0.01	290	950	1800	68	2.2	1.7	0.81	0.55	0.21	0.65	1.4	1.7	1.8	1.9
Settlement pond inlet	<0.01	0.19	3.5	0.37	0.13	0.02	<0.01	<0.01							
Derwent River hatchery discharge to water			0.23	0.31	46	18	5.7	0.21	0.48	0.16	0.04	<0.01	<0.01	0.04	0.06
Downstream (near) of hatchery discharge		1.5	0.02	0.02	0.09	0.04	<0.01	<0.01	<0.01		<0.01	<0.01	<0.01	0.01	<0.01
Downstream 500 m		0.02													
Downstream (far) of hatchery discharge		<0.01	0.02	0.01	<0.01	<0.01	0.02	<0.01	<0.01						

3.2 Water quality

Results from nine water sampling events as part of the OTC residue monitoring program are presented in Table 4.

The hatchery intake site was sampled from Day 1 of treatment and every week to Week 6 post-treatment and returned OTC concentrations of <LoR on every occasion.

OTC concentration was detected in the settlement pond inlet water on Day 1 of treatment (30 µg/L) and the highest value recorded mid treatment (49 µg/L). Concentration of OTC in the settlement pond dropped post-treatment with values <LoR from Day 14.

Hatchery discharge was sampled from Day 1 post-treatment and returned the highest results at this site on this occasion (3.9 µg/L), with concentration dropping progressively each week with values <LoR from Week 4 post-treatment.

Samples were taken at a site, 500 m downstream from the outfall during Day 1 of sampling (0.5 mg/kg DMB) and mid treatment (<0.01 mg/kg DMB). This occurred on Day 1 due to Huon Aquaculture having not received the monitoring schedule when treatment commenced. At mid treatment there was a sampling oversight. The sample originally identified as the hatchery discharge was in fact collected from the downstream near location, while the downstream near sample was collected from the site approximately 500 m downstream (Table 4). Therefore, Hatchery discharge was sampled from Day 1 post-treatment and returned the highest results at this site on this occasion (3.9 µg/L), with concentration dropping progressively each week with values <LoR from Week 4 post-treatment.

OTC concentrations at the near downstream site in the Derwent River was highest mid treatment (0.5 µg/L) with concentrations decreasing progressively to <LoR from Day 14.

OTC was not detected above the LoR in samples from the far downstream (Derwent River) site on any sampling occasion.

Water sampling continued to 24 March 2026, representing Week 6 post treatment when OTC concentration was <LoR at all sites for three consecutive weeks.

In summary, OTC residue in water samples was highest in the settling pond during treatment. OTC residue was retained in the hatchery discharge for longer than at any other sampling location.

Table 4. OTC concentration of water samples collected during the monitoring program at Meadowbank Hatchery. Results above the laboratory LoR (<0.01 µg/L) are shaded in blue. Weeks where sampling did not occur are shaded in grey.

Site	Day 1 of treatment	Mid treatment	Post treatment						
			Day 1	Day 7	Day 14	Day 21	Week 4	Week 5	Week 6
	21/01/2026	27/01/2026	10/02/2026	17/02/2026	24/02/2026	3/03/2026	10/03/2026	17/03/2026	24/03/2026
Hatchery Inlet	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
Settlement pond inlet	30	49	9	0.2	<0.1	<0.1	<0.1	<0.1	<0.1
Derwent River Hatchery discharge to water			3.9	1.9	0.6	0.2	<0.1	<0.1	<0.1
Downstream (near) of hatchery discharge		0.5	0.4	0.3	<0.1	<0.1	<0.1	<0.1	<0.1
500 m Downstream	<0.01	0.5							
Downstream (far) of hatchery discharge	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1

3.3 Wild fish

Results of the six wild fish sampling events are summarised in Table 5, including fish species and OTC concentration in mg/kg. Details on fish species, length and weight ins Appendix 3.

A total of five different wild fish species were sampled over the monitoring period, including the short finned eel (*Anguilla australis*), Freshwater sandy (*Pseudaphritis urvillii*), Jollytail galaxiid (*Galaxias maculatus*), Spotted galaxiid (*Galaxias truttaceus*), and Brown trout (*Salmo trutta*).

On some occasions AST increased the LoR due to small sample sizes (full body length of some fish was <100 mm, resulting in very small flesh samples collected). In response to this, the EPA requested that larger fish were targeted when possible. The LoR value used for each wild fish sample on each sampling occasion is documented in Table 5.

OTC was not detected above the laboratory LoR in wild fish samples from Day 1 following completion of treatment. OTC was detected above the LoR in a single sample (short finned eel, 0.02 mg/kg WMB) on Day 7 after treatment.

On Day 14 post-treatment the reporting limit was increased for some samples, from 0.01 mg/kg WMB to 0.02 or 0.1 mg/kg WMB, due to the small size of six of the 12 wild fish samples (Table 5). There were no detections of OTC above the LoR in Day 14 post-treatment samples.

Two fish samples (both short finned eels) returned elevated OTC concentrations on Day 21 post treatment, at concentrations of 0.05 and 0.02 mg/kg WMB. On Day 21 and Week 4 (Day 28) post-treatment, the LoR was increased for three samples on each sampling occasion due to small sample size (<5 grams). In Week 5, all sample sizes were sufficient for testing to LoR of 0.01 mg/kg WMB. In Week 4 and Week 5 post-treatment, OTC concentration was not detected above the laboratory LoR in any wild fish sample.

In summary, OTC was detected above the laboratory LoR in three wild fish samples over the six-week sampling period at Day 7 (one sample) and Day 21 (two samples). Of the five different

freshwater fish species sampled, OTC was only recorded above the LoR in short finned eels *A. australis*.

Table 5. OTC concentration (mg/kg WMB) in wild fish samples collected downstream of Meadowbank Hatchery during the monitoring program. Results above the LoR are shaded in blue. LoR typically = <0.01 mg/kg, unless sample was <5 g.

	Days / weeks post treatment					
	Day 1 10/02/2026	**Day 7 17/02/2026**	**Day 14 24/02/2026**	**Day 21 03/03/2026**	**Week 4 10/03/2026**	**Week 5 17/03/2026**
Fish 1	<0.01 Brown Trout	<0.01 Jollytail (galaxiid)	<0.01 Short Finned Eel	<0.01 Short Finned Eel	<0.01 Short Finned Eel	<0.01 Freshwater Sandy
Fish 2	<0.01 Short Finned Eel	<0.01 Spotted Galaxiid	<0.01 Freshwater Sandy	0.05 Short Finned Eel	<0.01 Short Finned Eel	<0.01 Short Finned Eel
Fish 3	<0.01 Short Finned Eel	<0.01 Short Finned Eel	<0.01 Freshwater Sandy	0.02 Short Finned Eel	<0.01 Short Finned Eel	<0.01 Short Finned Eel
Fish 4	<0.01 Short Finned Eel	<0.01 Short Finned Eel	<0.01 Short Finned Eel	<0.01 Short Finned Eel	<0.01 Short Finned Eel	<0.01 Short Finned Eel
Fish 5	<0.01 Short Finned Eel	<0.01 Short Finned Eel	<0.01 Freshwater Sandy	<0.10* Jollytail (galaxiid)	<0.01 Short Finned Eel	<0.01 Short Finned Eel
Fish 6	<0.01 Short Finned Eel	0.02 Short Finned Eel	<0.02* Spotted galaxiid	<0.01 Short Finned Eel	<0.01 Short Finned Eel	<0.01 Short Finned Eel
Fish 7	<0.01 Freshwater Sandy	<0.01 Freshwater Sandy	<0.1* Jollytail (galaxiid)	<0.10* Jollytail (galaxiid)	<0.01 Short Finned Eel	<0.01 Short Finned Eel
Fish 8	<0.01 Freshwater Sandy	<0.01 Freshwater Sandy	<0.1* Jollytail (galaxiid)	<0.01 Freshwater Sandy	<0.1* Jollytail (galaxiid)	<0.01 Short Finned Eel
Fish 9	<0.01 Freshwater Sandy	<0.01 Freshwater Sandy	<0.1* Jollytail (galaxiid)	<0.01 Short Finned Eel	<0.01 Freshwater Sandy	<0.01 Short Finned Eel
Fish 10	<0.01 Freshwater Sandy	<0.01 Freshwater Sandy	<0.02* Short Finned Eel	<0.01 Short Finned Eel	<0.02* Freshwater Sandy	<0.01 Short Finned Eel
Fish 11	<0.01 Freshwater Sandy	<0.01 Freshwater Sandy	<0.02* Freshwater Sandy	<0.05* Short Finned Eel	<0.02* Freshwater Sandy	<0.01 Short Finned Eel
Fish 12	<0.01 Freshwater Sandy	<0.01 Freshwater Sandy	<0.01 Freshwater Sandy	<0.01 Short Finned Eel	<0.01 Short Finned Eel	<0.01 Short Finned Eel

*Reporting limit increased due to the small sample size available.

Appendices

Appendix 1. Treatment Schedule

Treatment	Type	Days	Dates Prescribed	Active Ingredient (kg)
Antibiotic	Oxytetracycline	10	21/01/2026 – 31/01/2026	38.28
Antibiotic	Oxytetracycline	10	31/01/2026 = 9/02/2026	13.42

The treatments listed above did not overlap, as each fish received only one treatment. The two treatments were applied to separate tanks within the Meadowbank Hatchery.

Appendix 2. Operational Summary

Date	Sampling team	Works conducted
21/01/2026	Huon Aquaculture	Water sampling – Day 1 of treatment
	Huon Aquaculture	Sediment sampling – Day 1 of treatment
27/01/2026	Huon Aquaculture	Water sampling – Mid treatment
	Huon Aquaculture	Sediment sampling – Mid treatment
10/02/2026	Huon Aquaculture	Water sampling – Day 1 post treatment
	Huon Aquaculture	Sediment sampling – Day 1 post treatment
	Marine Solutions / Entura	Wild fish sampling – Day 1 post treatment
17/02/2026	Huon Aquaculture	Water sampling – Day 7 post treatment
	Huon Aquaculture	Sediment sampling – Day 7 post treatment
	Marine Solutions / Entura	Wild fish sampling – Day 7 post treatment
24/02/2026	Huon Aquaculture	Water sampling – Day 14 post treatment
	Huon Aquaculture	Sediment sampling – Day 14 post treatment
	Marine Solutions / Entura	Wild fish sampling – Day 14 post treatment
03/03/2026	Huon Aquaculture	Water sampling – Day 21 post treatment
	Huon Aquaculture	Sediment sampling – Day 21 post treatment
	Marine Solutions / Entura	Wild fish sampling – Day 21 post treatment
10/03/2026	Huon Aquaculture	Water sampling – Week 4 post treatment
	Huon Aquaculture	Sediment sampling – Week 4 post treatment
	Marine Solutions / Entura	Wild fish sampling – Week 4 post treatment
17/03/2026	Huon Aquaculture	Water sampling – Week 5 post treatment
	Huon Aquaculture	Sediment sampling – Week 5 post treatment
	Marine Solutions / Entura	Wild fish sampling – Week 5 post treatment
24/03/2026	Huon Aquaculture	Sediment sampling – Week 6 post treatment
	Huon Aquaculture	Water sampling – Week 6 post treatment
02/04/2026	Huon Aquaculture	Sediment sampling – Week 7 post treatment
10/04/2026	Huon Aquaculture	Sediment sampling – Week 8 post treatment
16/04/2026	Huon Aquaculture	Sediment sampling – Week 9 post treatment
20/04/2026	Huon Aquaculture	Sediment sampling – Week 10 post treatment
30/04/2026	Huon Aquaculture	Sediment sampling – Week 11 post treatment

Appendix 3. Wild Fish Sampling Length and Weight

	Number	Species	Length (mm)	Weight (g)
Day 1 Post Treatment (10/02/2026)	1	Brown Trout	105	10
	2	Short Finned eel	530	250
	3	Short Finned eel	400	97
	4	Short Finned eel	560	358
	5	Short Finned eel	455	171
	6	Short Finned eel	520	281
	7	Fresh Water Sandy	105	7
	8	Fresh Water Sandy	125	16
	9	Fresh Water Sandy	155	30
	10	Fresh Water Sandy	120	13
	11	Fresh Water Sandy	95	6
	12	Fresh Water Sandy	90	5
Day 7 Post Treatment (17/02/2026)	1	Jolly Tail (galaxiid)	75	<5
	2	Spotted Galaxiid	70	<5
	3	Short Finned Eel	560	387
	4	Short Finned Eel	510	311
	5	Short Finned Eel	530	317
	6	Short Finned Eel	160	6
	7	Fresh Water Sandy	150	31
	8	Fresh Water Sandy	97	7
	9	Fresh Water Sandy	115	14
	10	Fresh Water Sandy	95	7
	11	Fresh Water Sandy	90	6
	12	Fresh Water Sandy	97	6
Day 14 Post Treatment (24/02/2026)	1	Short Finned Eel	605	510
	2	Fresh Water Sandy	94	70
	3	Fresh Water Sandy	82	60
	4	Short Finned Eel	495	245
	5	Fresh Water Sandy	139	40
	6	Spotted Galaxiid	91	5
	7	Jolly Tail (galaxiid)	73	<5
	8	Jolly Tail (galaxiid)	77	<5
	9	Jolly Tail (galaxiid)	72	<5
	10	Short Finned Eel	150	5
	11	Fresh Water Sandy	94	5

	12	Fresh Water Sandy	106	10
Day 21 Post Treatment (03/03/2026)	1	Short Finned Eel	580	400
	2	Short Finned Eel	560	350
	3	Short Finned Eel	484	225
	4	Short Finned Eel	610	495
	5	Jolly Tail (galaxiid)	130	20
	6	Fresh Water Sandy	1540	25
	7	Jolly Tail (galaxiid)	83	15
	8	Fresh Water Sandy	130	25
	9	Short Finned Eel	183	20
	10	Short Finned Eel	556	385
	11	Short Finned Eel	135	5
	12	Short Finned Eel	479	235
Week 4 Post Treatment (10/03/2026)	1	Short Finned Eel	480	234
	2	Short Finned Eel	540	260
	3	Short Finned Eel	620	580
	4	Short Finned Eel	530	302
	5	Short Finned Eel	495	282
	6	Short Finned Eel	445	261
	7	Short Finned Eel	580	426
	8	Galaxias Maculatus	95	5
	9	Fresh Water Sandy	120	13
	10	Fresh Water Sandy	105	8
	11	Fresh Water Sandy	90	5
	12	Short Finned Eel	600	427
Week 5 Post Treatment (17/03/2026)	1	Fresh Water Sandy	120	5
	2	Short Finned Eel	560	305
	3	Short Finned Eel	405	205
	4	Short Finned Eel	540	340
	5	Short Finned Eel	580	375
	6	Short Finned Eel	555	290
	7	Short Finned Eel	470	225
	8	Short Finned Eel	430	175
	9	Short Finned Eel	470	190
	10	Short Finned Eel	500	215
	11	Short Finned Eel	495	245
	12	Short Finned Eel	410	135

Appendix 4. GPS Positions of sampling locations and photos

	Site Name	Chart Zone	Easting	Northing	Easting	Northing	
			Provided by EPA		Sample Location		Photo ID
Fishing	Discharge channel	GDA 94	488328E	5279492N			
	Hatchery outfall	GDA 94	488382E	5279473N			
	Maximum downstream extent of fishing	GDA 94	488423E	5279304N			
	Wild fish sample location	GDA 94			488359.36	5279472.01	F
Sediments	Hatchery Inlet / River Derwent (upstream)	GDA 94	488310E	5280094N	488360	5279791	A
	Derwent River at Hatchery discharge to water	GDA 94	488356E	5279475N	488336	5279482	D
	Settlement Pond adjacent to inlet	GDA 94	488222E	5279773N	488226	5279778	C
	Sludge Tank	GDA 94	488280E	5279791N	488258	5279807	B
	Downstream (near) of hatchery discharge	GDA 94	488377E	5279401N	488391	5279334	E
	Downstream (far) of hatchery discharge	GDA 94	489554E	5277521N	488751	5276524	
Water Quality	Hatchery Inlet / River Derwent (upstream)	GDA 94	488310E	5280094N	488360	5279791	A
	Derwent River at Hatchery discharge to water	GDA 94	488356E	5279475N	488336	5279482	D
	Settlement Pond adjacent to inlet	GDA 94	488222E	5279773N	488226	5279778	C
	Downstream (near) of hatchery discharge	GDA 94	488377E	5279401N	488391	5279334	E
	Downstream (far) of hatchery discharge	GDA 94	489554E	5277521N	488751	5276524	G
	500 m Downstream				488459.34	5279161.59	

A
B
C
G
D
E
F